А. Н. Цыганов, К. В. Бабешко, Ю. А. Мазей

Определитель родов раковинных амеб

Монография

A. N. Tsyganov, K. V. Babeshko, Yu. A. Mazei

A Guide to Testate Amoebae with the Keys to Genera

Monograph

Пенза Издательство ПГУ 2016

Рецензенты (Reviewers):

доктор биологических наук, профессор кафедры географии почв Московского государственного университета имени М. В. Ломоносова

А. А. Бобров

(Dr. A. A. Bobrov – professor at the Lomonosov Moscow State University);

доктор биологических наук, главный научный сотрудник лаборатории микробиологии Института биологии внутренних вод имени И. Д. Папанина Российской академии наук

оссийской академии наун

А. П. Мыльников

(Dr. A. P. Mylnikov – principal researcher at the Papanin Institute for Biology of Inland Waters, Russian Academy of Sciences)

Цыганов, А. Н.

Ц94

Определитель родов раковинных амеб : моногр. / A. H. Цыганов, K. B. Бабешко, Ю. A. Maзей = A Guide to Testate Amoebae with the Keys to Genera : monograph / A. N. Tsyganov, K. V. Babeshko, Yu. A. Mazei. – Пенза : Изд-во ПГУ, 2016. – 132 с.

ISBN 978-5-906913-19-7

Приводится современный обзор систематики раковин-ных амеб до уровня рода с определительными ключами и кратким описанием 107 родов. Представлены иллюстрации наиболее типичных представителей каждого рода. Издание предназначено для сотрудников научно-исследовательских организаций и студентов высших учебных заведений.

The book provides the modern synthesis on testate amoebae taxonomy at the genus level with identification keys and short description of 107 genera. Illustrations of the most typical taxa for each genus are included. The book is intended for professional researches and students.

УДК 593.11

Работа выполнена при финансовой поддержке Российского научного фонда (проект № 14-14-00891) и в рамках государственного задания Министерства образования и науки Российской Федерации (проект № 1315)

Financial support for the work was provided by the Russian Science Foundation (grant № 14-14-00891) and by the Ministry of Education and Science of the Russian Federation (project № 1315)

ISBN 978-5-906913-19-7

© А. Н. Цыганов, К. В. Бабешко, Ю. А. Мазей, текст, иллюстрации, 2016

TABLE OF CONTENTS

Introduction	7
Chapter 1. Systematics, biology and ecology of testate	
amoebae	10
Chapter 2. Order Arcellinida Kent, 1880	
[= Testacealobosia de Saedeleer, 1934]	16
Family Arcellidae Ehrenberg, 1843	20
Genus Arcella Ehrenberg, 1832	20
Genus Antarcella Deflandre, 1928, emend. Deflandre, 1953	28
Family Netzeliidae Kosakyan et al., 2016	28
Genus Netzelia Ogden, 1979	28
Family Cucurbitellidae Gomaa et al., 2017	29
Genus Cucurbitella Penard, 1902	29
Family Centropyxidae Jung, 1942	29
Genus Armipyxis Dekhtiar, 2009	31
Genus Centropyxis Stein, 1857	31
Genus Conicocassis Nasser et Patterson, 2015	31
Genus Proplagiopyxis Schönborn, 1964	37
Family Cryptodifflugiidae Jung, 1942	38
Genus Cryptodifflugia Penard, 1890	38
Genus Meisterfeldia Bobrov, 2016	42
Genus Wailesella Deflandre, 1928	42
Family Distomatopyxidae Bonnet, 1970	43
Genus Distomatopyxis Bonnet, 1964	43
Family Hyalospheniidae Schultze, 1877,	
emend. Kosakyan et Lara, 2012	43
Genus Alocodera Jung, 1942	45
Genus Apodera Loeblich et Tappan, 1961	46
Genus Certesella Loeblich et Tappan, 1961	46
Genus Cornutheca Kosakyan et al., 2016	46
Genus Gibbocarina Kosakyan et al., 2016	48
Genus Hyalosphenia Stein, 1859	48
Genus Mrabella Kosakyan et al., 2016	50
Genus Nebela (Leidy, 1874) Kosakyan et al., 2016	51
Genus Longinebela Kosakyan et al., 2016	52
Genus Padaungiella Lara et Todorov, 2012	52
Genus Planocarina Kosakyan et al., 2016	53

Genus Porosia (Jung, 1942) Bobrov et Kosakyan, 2015	. 53
Genus Quadrulella (Cockerell, 1909) Kosakyan et al., 2016	. 54
Family Lamtopyxidae Bonnet, 1974	. 54
Genus Lamtopyxis Bonnet, 1974	. 54
Family Microchlamyiidae Ogden, 1985,	
emend. Kudryavtsev et Hausmann, 2007	. 56
Genus Microchlamys Cockerell, 1911	. 56
Genus Spumochlamys Kudryavtsev et Hausmann, 2007	. 56
Family Microcoryciidae de Saedeleer, 1934	. 57
Genus Amphizonella Greeff, 1866	. 57
Genus Diplochlamys Greeff, 1888	. 58
Genus Microcorycia Cockerell, 1911	. 58
Genus Parmulina Penard, 1902	. 59
Genus Penardochlamys Deflandre, 1953	. 59
Genus Zonomyxa Nüsslin, 1882	. 60
Family Plagiopyxidae Bonnet et Thomas, 1960	. 60
Genus Bullinularia Deflandre, 1953	. 61
Genus Geoplagiopyxis Chardez, 1961	. 61
Genus Hoogenraadia Gauthier-Lièvre et Thomas, 1958	. 62
Genus Paracentropyxis Bonnet, 1960	. 62
Genus Plagiopyxis Penard, 1910	. 63
Genus Planhoogenraadia Bonnet, 1977	. 64
Genus Protoplagiopyxis Bonnet, 1962	. 64
Family Paraquadrulidae Deflandre, 1953	. 64
Genus Lamtoquadrula Bonnet, 1974	. 65
Genus Paraquadrula Deflandre, 1932	. 65
Family Phryganellidae Jung, 1942	. 65
Genus Phryganella Penard, 1902	. 65
Family Trigonopyxidae Loeblich et Tappan, 1964	. 66
Genus Cornuapyxis Coûteaux et Chardez, 1981	. 67
Genus Cyclopyxis Deflandre, 1929	. 67
Genus Geopyxella Bonnet et Thomas, 1955	. 67
Genus Trigonopyxis Penard, 1912	. 70
INCERTAE SEDIS Arcellinida	. 70
Genus Argynnia Vucetich, 1974	. 70
Genus Awerintzewia Schouteden, 1906	. 72
Genus Difflugia Leclerc, 1815	. 72
Genus Geamphorella Bonnet, 1959	. 75
Genus Ellipsopyxella Bonnet, 1975	. 76
Genus Ellipsopyxis Bonnet, 1965	. 76

Genus <i>Heleopera</i> Leidy, 1879	77
Genus Jungia Loeblich et Tappan, 1961	77
Genus Lagenodifflugia Medioli et Scott, 1983	78
Genus Leptochlamys West, 1901	78
Genus Lesquereusia Schlumberger, 1845	79
Genus Maghrebia Gauthier-Lièvre et Thomas, 1960	79
Genus Mediolus Patterson, 2014	80
Genus Microquadrula Golemansky, 1968	80
Genus Oopyxis Jung, 1942	81
Genus Pentagonia Gauthier-Lièvre et Thomas, 1960	81
Genus Physochilla Jung, 1942	81
Genus Pomoriella Golemansky, 1970	82
Genus Pontigulasia Rhumbler, 1896	83
Genus Protocucurbitella Gauthier-Lièvre et Thomas, 1960	84
Genus Pseudocucurbitella Gauthier-Lièvre et Thomas, 1960.	84
Genus Pseudawerintzewia Bonnet, 1959	84
Genus Pseudonebela Gauthier-Lièvre, 1953	84
Genus Pyxidicula Ehrenberg, 1838	85
Genus Schoenbornia Décloître, 1964	85
Genus Schwabia Jung, 1942	85
Genus Sexangularia Awerintzew, 1906	86
Genus Suiadifflugia Green, 1975	86
Genus Zivkovicia Ogden, 1987	87
Chapter 3. Order Euglyphida Copeland, 1956, emend.	
Cavalier-Smith, 1997	88
Family Assulinidae Lara et at., 2007	89
Genus Assulina Leidy, 1879	89
Genus <i>Placocista</i> Leidy, 1879	89
Genus Valkanovia Tappan, 1966	91
Family Euglyphidae Wallich, 1864, emend. Lara et al., 2007	91
Genus Euglypha Dujardin, 1841	91
Genus <i>Scutiglypha</i> Foissner et Schiller, 2001	95
Family Sphenoderiidae Chatelain, 2013	95
Genus Sphenoderia Schlumberger, 1845	95
Genus Trachelocorythion Bonnet, 1979	96
Family Trinematidae (Hoogenraad et de Groot, 1940)	0.6
Adl et al., 2012	96
Genus Corythion Taranek, 1881	97
Genus <i>Playfairina</i> Thomas, 1961	97
Genus <i>Trinema</i> Dujardin, 1841	98

Genus Puytoracia Bonnet, 1970	. 98
Family Cyphoderiidae de Saedeleer, 1934	. 98
Genus Campascus Leidy, 1879	. 100
Genus Corythionella Golemansky, 1970	. 100
Genus Cyphoderia Schlumberger, 1845	. 101
Genus Messemvriella Golemansky, 1973	. 101
Genus Pseudocorythion Valkanov, 1970	. 101
Genus Schaudinnula Awerintzew, 1907	. 103
Family Paulinellidae de Saedeleer, 1934, emend. Adl et al., 2012	. 103
Genus Micropyxidiella Tarnawski et Lara, 2015	. 104
Genus Ovulinata Anderson et al., 1997	. 104
Genus Paulinella Lauterborn, 1895	. 105
INCERTAE SEDIS Euglyphida	. 105
Genus Ampullataria van Oye, 1956	. 105
Genus Deharvengia Bonnet, 1979, emend.	
Bobrov et al., 2012	. 106
Genus Euglyphidion Bonnet, 1960	. 106
Genus Heteroglypha Thomas et Gauthier-Lièvre, 1959	. 107
Genus Matsakision Bonnet, 1967	. 107
Genus Pareuglypha Penard, 1902	. 107
Genus Pileolus Coûteaux et Chardez, 1981	. 107
Genus Tracheleuglypha Deflandre, 1928	. 108
Chapter 4. Order Amphitremida Poche, 1913, emend. Gomaa,	
Mitchell et Lara, 2013	. 109
Family Amphitremidae Poche, 1913	. 109
Genus Amphitrema Archer, 1869	. 109

Genus Archerella Loeblich et Tappan, 1961	109
References	111

Introduction

Testate amoebae (also referred to as testaceans) are free-living amoeboid protists in which the cytoplasm is enclosed within discrete shell or test and which produce pseudopodia for movement and feeding. Testate amoebae have worldwide distribution and form diverse and abundant communities in freshwater (Beyens, Meisterfeld, 2002; Mazei, Tsyganov, 2006) and terrestrial habitats, including soils, mosses, standing waters and sewage treatment works (Schönborn, 1962a, 1962b, 1966b, 1983, 1989; Coûteaux, 1976; Geltzer et al., 1985; Foissner, 1987; Charman et al., 2000; Finlay et al., 2000). They are generally considered rare in marine systems, but new taxa, predominantly interstitial, are continued to be described (Golemansky, 1978, 1990, 1994, 2000; Golemansky, Todorov, 1999; Scott et al., 2001; Nicholls, 2009a). Testate amoebae prey on a wide range of organisms including bacteria, fungi, algae, other protozoa, micrometazoa and may probably consume dead organic matter (Heal, 1963; Gilbert et al., 2000; Wilkinson, Mitchell, 2010). Some species of testate amoebae contain endosymbiotic algae and are probably mixotrophic; the genus Paulinella includes phototrophic species (Gomaa et al., 2014). The total number of testate amoebae species can be estimated as 2000 (Meisterfeld, 2000a, 2000b).

Studies on testate amoebae started in the beginning of the 19th century, when the first species were described (Leclerc, 1816; Ehrenberg, 1838; Dujardin, 1841). By the end of the 19th and beginning of the 20th century a considerable amount of material was published on the morphology and systematics of testate amoebae from different parts of the world (Wallich, 1864; Leidy, 1879; Penard, 1890, 1899, 1902, 1903; Cash, Hopkinson, 1905, 1909, Awerintzew, 1906; Wailes, 1912; Cash et al., 1915, 1919). Later fossil testate amoebae were discovered in lake sediments (Lagerheim, 1902) and peat deposits (Harnisch, 1924, 1925, 1927; Steinecke, 1927) that lay down the ground for further use of testate amoebae in palaeoecological studies. At the same time, accumulation of data on testate amoebae allowed to build a detailed systematic description of some genera based on morphology (Deflandre, 1928a, 1929, 1936), and to developing a macro taxonomy of testate amoebae (Saedeleer, 1934; Hoogenraad, de Groot 1940a; Jung, 1942; Deflandre, 1953). In the middle of the 20th century several monographs

and identification guides covering all aspects of the biology and ecology of testate amoebae were published (Bartoš, 1954; Grospietsch, 1958; Harnisch, 1958; Schönborn, 1966c; Chardez, 1967a). By the 1990s, most of the genera of testate amoebae were revised basing on shell morphology: Arcella (Deflandre, 1928; Décloître, 1976), Centropyxis (Deflandre, 1929; Décloître 1978, 1979), Cryptodifflugia (Grospietsch, 1964; Schönborn, 1965a; Page, 1966), Cucurbitella (Gauthier-Lièvre, Thomas, 1960), Cyclopyxis (Deflandre, 1929; Décloître, 1977a), Cyphoderia (Chardez, 1991), Difflugia (Štěpánek, 1952; Gauthier-Lièvre, Thomas, 1958; Chardez, 1961, 1967c; Ogden, 1979, 1980a, 1980b, 1983, 1984; Ogden, Hedley, 1980; Ogden, Meisterfeld, 1989; Ogden, Fairman, Živković, 1983), Euglypha Ogden. (Décloître, 1979: 1962b). Hyalosphenia (Grospietsch, 1965), Lesquereusia (Thomas, Gauthier-Lièvre, 1959b), Nebela (Deflandre, 1936; Gauthier-Lièvre, 1953; Jung, 1942; Décloître, 1977b), Plagiopyxis (Thomas, 1958a), Quadrulella (Chardez, 1967b), Paraquadrula (Décloître, 1962a; Schönborn, 1965b), Trinema (Chardez, 1960a). Numerous studies investigated distribution of testate amoebae around the globe, with greater attention to the temperate regions of Europe and North America though, and their role in various types of ecosystems (Heal, 1961; Schönborn, 1992).

Research on testate amoebae has increased substantially since 1990s due to their increasing use as bioindicators for paleoecological studies (Mitchell et al., 2008). Multivariate classifications of testate amoeba assemblages (Tolonen et al., 1992, 1994; Mitchell et al., 1999) and quantitative relationships between community structure and specific environmental variables have been explored using univariate and multivariate statistics (Bobrov et al., 1999; Booth, 2001; Lamentowicz, Mitchell, 2005; Charman et al., 2007), which resulted in development of quantitative reconstructions of peatland surface moisture basing on fossil testate amoebae assemblages. Also, it has been shown that testate amoebae play important roles in the cycling of elements in terrestrial ecosystems (Aoki et al., 2007; Schröter et al., 2003). They are also increasingly used in ecotoxicology, forensic sciences, biomonitoring and in many other applied aspects (Nguyen-Viet et al., 2007; Payne, 2011, 2013; Fournier et al., 2012; Szelecz et al., 2014). Testate amoebae are a good model for taxonomical, evolutionary and ecological evolutionary studies because of their diversity, ubiquity, the presence of a shell which is taxonomically diagnostic, and their long (but discontinuous and still very poorly studied) fossil record (Porter, Knoll, 2000; Schmidt et al., 2004; 2006).

Molecular phylogenetic studies have considerable advanced our understanding of the relationships among testate amoebae and established their phylogenetic position in the tree of eukaryotes (Wylezich et al., 2002; Nikolaev et al., 2005; Gomaa et al., 2012; 2017; Kosakyan et al., 2012, 2013, 2016, 2016a). However, most of the especially useful taxonomical monographs for identification of testate amoebae followed morphology-based approach mostly (Ogden, Hedley, 1980; Ellison, Ogden, 1987; Corbet, 1973; Charman et al., 2000; Mazei, Tsyganov, 2006; Mazei, Warren, 2012, 2014, 2015). The most recent synthesis on testate amoebae taxonomy is a genus-level work released shortly before any molecular data on testate amoebae were available (Meisterfeld, 2002a, 2002b).

The main purpose of this guide was to combine all modern data on genus level systematics of testate amoebae and to provide identification keys to genera with illustrations of the most typical taxa. In this Guide, we compiled descriptions for 104 genera of testate amoebae. This covers most of the existing genera of testate amoebae with new taxonomic alterations since the last revision done by R. Meisterfeld (2002a, 2002b). The Guide does not cover most of marine species and filose testate amoebae with flexible shells (Thecofilosea).

High taxonomic levels and phylogeny accepted in this work are based on revisions provided by Adl et al. (2012) and Ruggiero et al. (2015). Low taxonomic levels (i.e. genera) accepted in this work are based on the revision of Kosakyan et al. (2016). Families within orders and genera within families are listed alphabetically. Pictures and species names are taken from Mazei, Tsyganov (2006).

Acknowledgements

Financial assistance was provided by the Russian Science Foundation (grant N_{2} 14-14-00891 – taxonomic analysis, e.g. construction of keys, arrangement of taxa descriptions and figure plates) and by the Ministry of Education and Science of the Russian Federation (project N_{2} 1315 – logistical support in publishing).

Chapter 1 Systematics, biology and ecology of testate amoebae

Systematics. The traditional morphology-based classification grouped testate amoebae together with other amoeboid protists in a macrotaxon named Sarcodina (Levine et al., 1980). However, recent molecular research has demonstrated that testate amoebae are polyphyletic and belong in three evolutionary distinct major groups not directly related to each other (table).

Table

Systematics of the higher taxonomy of testate amoebae based on hierarchical classification from superkingdom to order proposed by Ruggiero et al. (2015)

Superkingdom Eukaryota

Kingdom Protozoa Owen, 1858, emend. Cavalier-Smith, 1998
Subkingdom Sarcomastigota Cavalier-Smith, 1983, emend. Cavalier-Smith, 2013
Phylum Amoebozoa Lühe, 1913, emend. Cavalier-Smith, 1998
Subphylum Lobosa Carpenter, 1861, emend. Cavalier-Smith, 2009
Class Tubulinea Smirnov et al., 2005 [= Lobosea Carpenter, 1861]
Order Arcellinida Kent, 1880 [= Testacealobosia de Saedeleer, 1934]

Kingdom Chromista Cavalier-Smith, 2010

Subkingdom Harosa Cavalier-Smith, 2010 [= "Supergroup SAR"] Infrakingdom Halvaria Cavalier-Smith, 2010 Superphylum Heterokonta Cavalier-Smith, 1981 [= "Supergroup Stramenopiles"] Phylum **Bigyra** Cavalier-Smith et Chao, 2006 Class Labyrinthulea/Labyrinthulomycetes Dick, 2001 Order Amphitremida Poche, 1913, emend. Gomaa, Mitchell et Lara, 2013 Infrakingdom Rhizaria Cavalier-Smith, 2002 Phylum Cercozoa Cavalier-Smith, 1998, emend. Adl et al., 2005 Subphylum Monadofilosa Cavalier-Smith, 2003 Class Imbricatea Cavalier-Smith, 2011, emend. Cavalier-Smith, 2003 Subclass Placonuda Cavalier-Smith et Chao, 2012 Order Eulyphida Copeland, 1956, emend. Cavalier-Smith, 1997 Class Thecofilosea Cavalier-Smith, 2003, emend. Cavalier-Smith, 2011 Subclass Eothecia Cavalier-Smith et Chao, 2012 Order Cryomonadida Cavalier-Smith, 1993 Subclass Tectosia Cavalier-Smith et Chao, 2012 Order Tectofilosida Cavalier-Smith, 2003

The largest group of testate amoebae named Arcellinida comprises testaceans with lobose pseudopodia and belongs in protozoan phylum Amoebozoa (Nikolaev et al., 2005). Most of the remaining testate amoebae with filose pseudopodia are now placed in the phylum Cercozoa (Wylezich et al., 2002) which belongs in the "SAR" clade (Burki et al., 2007), now formally subkingdom Harosa of the kingdom Chromista (Cavalier-Smith, 2010). Some of testate amoebae with anastamosing networks of reticulopodia and two symmetrical apertures (i.e. genera *Amphitrema* and *Archerella*) are now classified in the phylum Bigyra of the subkingdom Harosa (Gomaa et al., 2013). Despite being a polyphyletic group, testate amoebae form a reasonably uniform morphological and ecological group (Wilkinson, Mitchell, 2010) which can be studied using common methods. Treating testate amoebae as a single group in ecological studies can be compared to the studies on vegetation ecology, which often include lichens.

Cell. In its simplest division, the main elements of the living amoeba are the cell itself and the shell in which it lives. The cytoplasm usually fills the chamber in smaller testate amoebae such as *Euglypha* and *Trinema*, whereas in larger species it only partially fills the chamber and thin cytoplasmatic strands (epipodia) attach it the shell wall. The most important cell structures for identification of testate amoebae are nucleus and pseudopodium type. According to the classification of Raikov (1982) two principal types of nuclei are found in testate amoebae: 1) vesicular nuclei with one, often central nucleolus, sometimes with a few additional, very small nucleoli; 2) ovular nuclei with several or many small nucleoli.

Testate amoebae can have three types of pseudopodia: lobopodia, filopodia or reticulopodia. Lobopodia are projections more or less broad, with cytoplasmic streaming; they may have a clear hyaline region at the front; additional fine projections may extend from this hyaline region. This type of pseudopodia is typical for the order Arcellinida. Filopodia are fine, pointed hyaline projections, containing no granuloplasm or microtubules, sometimes branching, but never anastomosing. This type of pseudopodia is typical for the order Euglyphida. Reticulopodia represent a network or of fine anastomosing pseudopodia without granules.

Testate amoebae mostly reproduce by asexual binary fission. The role of sexual reproduction in the life cycle of testate amoebae remains basically unknown, however the available data point at the wide distribution and an important role of sexual reproduction (Lahr et al., 2011). Under unfavourable environmental conditions testate amoebae produce cysts (Thomas, 1962).

Shell. Testate amoeba shells normally consist of one chamber with a single aperture. However, some taxa are characterised by two-chamber shells or by the presence of two apertures. Shells are composed of organic material which is often encrusted with exogenous mineral particles or covered by self-secreted silica plates. The principal function of the shell is the protection of amoebae from negative environmental influence, mainly from predation and desiccation. The aperture connects amoebae with the external environment so that pseudopodia can extend through the aperture for locomotion and feeding. The aperture determines the maximal size of food particles that can be ingested by the cell, participates in holding and positioning of shells during cell division and protects the cell from desiccation and predator invasions.

Shells of testate amoebae can be radially symmetrical or bilaterally symmetrical. Radially symmetrical shells can in turn be round in cross section (with several, i.e. more than two, cutting plains parallel to the axis of symmetry) or compressed in cross section (only two cutting plains parallel to the axis of symmetry). The axis of symmetry extends from the center of the oral pole which contains the aperture (mouth) to the centre of the opposite, or aboral, end (i.e. radial heteropolar symmetry). Such shells do not have left or right sides. Testate amoebae with two apertures (*Amphitrema, Archerella*) have radial homopolar symmetry. Apertures are normally located perpendicular (at the right angle) to the axis of symmetry.

When the length of the axis of symmetry is relatively short as compared to the diameter of the shell, shell is hemispheric or disk-like. In this case, the apertural side can sometimes be referred to as a ventral side, whereas the aboral region is referred to as dorsal side. These shells can be normally seen in preparations in the frontal view, which corresponds to a frontal plain (plains perpendicular to the axis of symmetry and dividing the shell into dorsal and ventral portions). Most of the radially symmetrical shells with a shortened axis of symmetry are circle in cross section, whereas elliptical cross section is typical only for *Ellipsopyxella* and *Ellipsopyxis*. The aperture can be just a simple circular opening on the ventral side (*Phryganella*) or be located at the end of the invaginated (rarely turned out) tube (*Acella, Antracella, Pyxidicula, Cyclopyxis*). Sometimes, aperture can be lobate, denticulate or reinforced by apertural

apparatus such as thick teeth (Lamtopyxidae) or a perforated diaphragm (Distomatopyxidae).

In cases, when the length of the axis of the symmetry is comparable to the shell diameter or longer, shells can be spherical (close to or perfectly circular), ovoid (oval-shaped with convex sides), elongate (length more than 1.5 times greater than the breadth), pyriform (pearshaped, wider posterior end than the anterior) or bottle-shaped with a well defined neck. Neck is an extension from a shell which terminates by an aperture. Aperture is generally situated at right angles to the axis of symmetry of the shell (terminal aperture). The aboral region is referred to as fundus. The shells are normally seen in preparations lying of their side (lateral view). In compressed shells, broad and narrow lateral views can be distinguished. Aperture is generally circle, but it can also be denticulate or lobed. Sometimes, aperture can be located in a small chamber (Cucurbitellidae, *Lagenodifflugia, Zivkovicia, Pontigulasia* etc.)

In bilaterally symmetrical (also called plane symmetry) shells, only one cutting plane (called the sagittal plane) divides a shell into roughly mirror image halves (with respect to external appearance only). There is no axis of symmetry. Often the two halves can meaningfully be referred to as the right and left halves. Bilaterally symmetrical shells are characterized by ventral and dorsal sides. The aperture can be located at an oblique angle to the axis of the symmetry (subterminal), at the end of a curved neck (*Lesqueresia, Cyphoderia*) or on ventral side of the shell (Plagiopyxidae). The aperture can be a simple circle opening or be located at the end of an invaginated tube, but sometimes it evolves to a slit-like structure (cryptostome) often covered by the anterior (dorsal) lip (*Plagiopyxis*) or a visor (*Planhoogenraadia*). This type of symmetry is generally characteristics for organisms which live on the surface.

Depending on the composition of the shell four shell types can be distinguished: proteinaceous, agglutinated, siliceous and calcareous.

Proteinaceous shells are completely organic and bear no or very scarce covering materials. There are two main types of proteinaceous shells: areolate and non-areolate. Areolate shells are constructed of regularly arranged hollow building units to from an areolate surface. These shells can be rigid (*Arcella* and *Pyxidicula*) or flexible to semi-rigid (Microchlamyiidae). Non-areolate shells are mostly composed of rigid sheet of fibrous material (*Hyalosphenia, Archerella* etc.). In rare case the shell is more or less flexible and represents a membrane that encloses the cytoplasm (Microcoryciidae).

Agglutinated shells consist of a cement matrix of often perforated building units or sheet-like cement in which foreign material (xenosomes) is incorporated. As xenosomes testate amoebae can use quartz grains, whole diatoms or pieces of diatoms and chrysomonad cysts from the environment (*Difflugia, Centropyxis, Lagenodifflugia, Awerintzewia* etc.) or siliceous scales their prey testate amoebae, usually members of the order Euglyphida (*Nebela*). Normally, xenosomes are randomly arranged on the shell surface; however they can have specific pattern of organization usually around the aperture (*Cyclopyxis*).

Siliceous shells are built of a cement matrix which is coved by endogenously synthesized siliceous scales or plates (idiosomes). Normally, the plates are regularly arranged on the surface of the shell. The plates differ in shape, size and arrangement and are genus- and species-specific. Normally, there are no more than four types of plates or spines can be produced by one species. Most of the species with siliceous shells are found among filose amoeba (Euglyphidae, Sphenoderiidae, Trinematidae, Cyphoderiidae etc.). Among lobose amoebae, this type of shells is typical for *Lesquereusia* and *Quadrulella*. Some testate amoebae can use both idiosomes and xenosomes for shell construction, e.g. *Heleopera*.

Calcareous shells are characterized by the presence of calcareous elements, either plates or shell layers. These shells are characteristic for the genus *Cryptodiffludia* and the family Paraquadrulidae. The former has a thick layer of calcium phosphate deposited within an organic template, whereas the later has rectangular calcite plates bound by an internal sheet of cement.

Ecology. A great number of studies have demonstrated the important role of substrate moisture in regulation of abundance, species diversity and assemblage composition of testate amoebae in soils (Geltzer et al., 1985; Foissner, 1987) and in *Sphagnum* mosses (Tolonen et al., 1992, 1994; Bobrov et al., 1999; Charman et al., 2000; Booth, 2002; Lamentowicz, Mitchell, 2005). The importance of substrate moisture in regulation of testate amoeba assemblages is related to the fact that water basically provides the environment for testate amoeba activity. That is why thicker water film can physically accommodate a greater number of testate amoebae. Low substrate moisture demands special adaptation from species to survive so that dry biotopes are normally inhabited by smaller number of predominantly xerophilous species. Testate amoebae

adapted to water deficit by reduced shell size to reach smaller pores and inhabit thinner water films or by invagination of the pseudostome to protect cells from desiccation (Bonnet, 1975).

Testate amoebae have been shown to be sensitive to a number of other environmental parameters which affect testate amoebae either directly or indirectly: pH (Heal, 1961; Booth, 2002; Lamentowicz, Mitchell, 2005); nutrient availability (Aescht, Foissner, 1992; Mitchell, 2004; Krashevska et al., 2010); concentrations of metal ions (Mitchell et al., 2004; Carlson et al., 2010); vegetation (Carlson et al., 2010; Sullivan, Booth, 2011); food source type (Krashevska et al., 2008); concentration, composition of mineral of building particles for shells and light (Heal, 1962).

Chapter 2 Order Arcellinida Kent, 1880 [= Testacealobosia de Saedeleer, 1934]

Testate, inside an organic or mineral extracellular shell of either self-secreted elements (calcareous, siliceous, or chitinoid) or recycled mineral particles bound together, with a single main opening.

Key to the families of the order and genera *incertae sedis*

1. Shell is flexible or semi-rigid	2
1'. Shell is rigid	
2. Shell is finely areolate	Family Microchlamyidae
2'. Shell is not areolate	Family Microcoryciidae
3. Shell is radially symmetrical; cro	ss section is circular or
compressed	4
3'. Shell is bilaterally symmetrical of	or with a bent neck
4. Ventral face is flat, aperture is loo	cated in the center
of ventral surface, invaginated or not	5
4'. Aperture is terminal	
5. Shell is completely organic	
5'. Shell is covered by mineral parti	cles
6. Shell is areolate	7
6'. Shell is not areolate	
Genus Cryptodifflugi	a (Family Cryptodifflugiidae)
7. Aperture is smaller than half of the	ne shell diameter
	Family Arcellidae
7'. Aperture is almost as large as the	e shell diameter
	Genus Pyxidicula
8. Shell is circular in cross section	
8'. Shell is elliptic in cross-section.	9
9. Aperture is circular	Genus Ellipsopyxella
9'. Aperture is elliptical	Genus Ellipsopyxis
10. Aperture is circular, ovular, trian	ngular, polygonal, lobed or
crescent-shaped	

10'. Apertural apparatus with an internal opening at the end of an invaginated tubus and an external aperture with either teeth-like 11. Aperture normally is less than a half of the shell diameter 11'. Aperture normally is more than a half of the shell diameter..... 12. External aperture with two crescent-shaped openings 12'. External aperture bordered by large teeth..... 13. Shell is compressed or polygonal in cross section 13'. Shell is circular or slightly compressed in cross section......18 14'. Shell is polygonal in cross section17 15. Aperture is surrounded by a thin organic lip (rim, collar or fringle)16 15'. Aperture is without organic lipGenus Difflugia 16. Aperture is slit-like with acute notches at edges 16'. Aperture is circular or oval, without acute notches at edges..... 17. Shell is hexagonal in cross section, completely organic 17'. Shell is pentagonal, covered by mineral particles..... 18. Shell is completely organic Genus Leptochlamys 18'. Shell is covered by mineral particles, idiosomes of prey species 19. Aperture is surrounded by a distinct lobed or denticulate collar or by a complex, consisting of a central pore and irregularly elongate oval 19'. Aperture is circular, oval, lobed or denticulate, sometimes with an organic lip, no collar, no distinctions between internal and external 20. Aperture complex with central opening and numerous elongate oval pores radiating around central pore Genus Suiadifflugia

21. Collar is lobed	22
21'. Collar is relatively smooth or denticulate	24
22. Collar forms a cavity (frontal camera) which has an inner	
aperture (circular or lobed) on the level with the main body wall	
	dae
22'. Collar does not form a cavity, no inner aperture	23
23. Collar is lobed, with a thick organic rim or a necklace made	;
of small idiosomes	dae
23'. Collar is formed by 3–5 separate lobes	
	ella
24. Collar is straight, chitionoid or made of quartz grains	25
24'. Collar is recurved posteriorly, covered with siliceous	
exogenous plates	illa
25. Collar is chitionoidGenus Geamphor	ella
25'. Collar is short, made of quartz grains	ıgia
26. Shell composed mainly of mineral grains or diatoms.	0
often opaque gravish	27
26'. Shell completely organic or composed of circular, oval.	
nail-shaped or rectangular plates, collected or endogenously	
formed, sometimes with admixture of mineral grains	
or diatoms	36
27. Shell is with 3-4 lateral bulges near aperture	
	ebia
27'. No lateral bulges	28
28. Aperture is denticulate	29
28'. Aperture is not denticulate	30
29. The peristome is surrounded by distinctive diaphragm	
Genus Media	olus
29'. No diaphragm around the peristome	
	ella
30. Shell partitioned in two parts by a diaphragm between	
neck an main body, often visible as constriction	31
30'. No such diaphragm	33
31. Internal diaphragm with one opening	
	ıgia
31'. Internal diaphragm with two openings	32
32. Diaphragm composed of small mineral grains, two circular	
openings	icia

32'. The openings formed by a mainly organic bridge attached mineral particles which connects both broad sides	with few
	s Pontigulasia
33. Shell mainly composed of angular mineral particl	les
of diatoms; aperture is relatively large, circular, oval or lob	ed
Ge	enus <i>Difflugia</i>
33'. Aperture is relatively small, shell deep violet	
or brownish with smooth surface	
34. Aperture at the broader end of the brownish	
shell with smooth chitionoid surface Genus Pseud	lawerintzewia
34'. Aperture at the narrow end of the shell	
35. Aperture is oval, shell deep violetGenus	Awerintzewia
35'. Aperture is circular, relatively small	
apertureGe	nus <i>Schwabia</i>
36. Shell is completely organic or covered with	
rare particles of foreign material	
Genus Cryptodifflugia (Family Crypt	odifflugiidae)
36'. Shell is completely agglutinated	
37. Aperture with an organic lip	
37'. Aperture without any organic lip	
38. Organic lip is smooth	
Genus Longinebela, Genus Padaungiella (Family Hya	lospheniidae)
38'. Aperture with organic, denticulate lip, appears lo	bed
	Pseudonebela
39. Shell is covered by square plates	40
39'. Shape of the plates covering shell is different	41
40. Aperture is slit-like	
Genus Paraquadrula (Family Para	quadrulidae)
40'. Aperture is slightly truncated	licroquadrula
41. Shell is composed of collected idiosomes of smal	
euglyphids	Schoenbornia
41'. Aperture is surrounded by siliceous plates,	<i>,</i>
giving it a rough outline	nus Argynnia
42. Aperture is circular, elongated or irregular	
42'. Aperture is slit-like Family I	Plagiopyxidae
43. Shell is covered by mineral particles	44
43 [°] . Shell is completely organic	1.661
Genus Wailesella (Family Crypt	odifflugiidae)
44. Aperture is located on a ventral side	45

eck46	44'. Aperture is located at the end of a b
l side is flat	45. Aperture is relatively large, circle; v
amily Centropyxidae	
	45'. Aperture is relatively small, irregula
Genus Oopyxis	ventral side is convex
es sometimes with	46. Shell is covered by rod-like siliceous
. Genus <i>Lesquereusia</i>	admixture of mineral particles
47	46'. Shell is covered by square scales
iceous	47. Marine species, scales are presumab
Genus Pomoriella	
bly calcite	47'. Tropical soil species, scales are pres
ily Paraquadrulidae)	Genus Lamtoquadrula (

Suborder Sphaerothecina Kosakyan et al., 2016

Shell is circular in lateral view, with radial symmetry; spherical or slightly flattened in cross section, aperture is circular or with lobes. Type family: Netzeliidae Kosakyan et al., 2016.

Family Arcellidae Ehrenberg, 1843

Shell is rigid, radially symmetrical, round, polygonal or dentate in the front view; spherical, hemispherical, flattened or disk-like in lateral view. Shell is completely organic, composed of box-like building units arranged in a single layer and cemented together, resulting in an areolar surface. Shell is transparent, colorless, yellow or brownish. Aperture is located on the ventral side, invaginated (rarely evaginated), circular, polygonal or lobed, often surrounded by small organic rim or a circle of pores.

Key to the genera of the family

Genus Arcella Ehrenberg, 1832

Shell as the family description. Most species are binucleate, but several species have more nuclei, e.g. *A. megastoma* may have up to 200. These nuclei are always vesicular. Ecology: freshwater, mosses and soils. Type species: *Arcella vulgaris* Ehrenberg, 1832 (fig. 1–7).





a – A. apicata lateral view (after Schaudinn, 1898); b, f – A. arenaria lateral (f) and aperture (b) view (after Deflandre, 1928a); c, g – A. arenaria compressa lateral (g) and aperture (c) view (after Décloître, 1976); d – A. arenaria irregularis aperture view (after Décloître, 1972); h, i – A. arenaria sphagnicola lateral (h) view and cysts (i) (after Deflandre, 1928a); e – A. arenaria sphagnicola undulata (after Décloître, 1976); j – Arcella artocrea (after Leidy, 1879); k, n, m – A. artocrea pseudocatinus lateral (k), aperture (n) view and cyts (m) (after Leidy, 1879); l, o – A. bathystoma lateral (l) and aperture (o) view (after Deflandre, 1928a); p, q – A. brasiliensis lateral (p) and aperture (q) view (after Cunha, 1913); r – A. catinus cyst (after Deflandre, 1928a)



Fig. 2. Genus Arcella:

a–d – A. conica lateral (a, c) and dorsal (b, d) view (after Deflandre, 1928a); e, f – A. costata lateral (e) and aperture (f) view (after Playfair, 1917); g, h – A. costata angulosa lateral (g) and aperture (h) view (after Penard, 1902); i–k – A. dentata lateral (i), aperture (k) and aperture-lateral (j) view (after Deflandre, 1928a); l–n – A. dentata trapezica lateral (m, n) and aperture (l) view (after Deflandre, 1928a); o, p – A. dentata cashiana lateral (o) and aperture (p) view (after Deflandre, 1928a)



Fig. 3. Genus Arcella:

a, b – A. discoides lateral view (a – after Deflandre, 1928a; b – after Leidy, 1879); c – A. discoides difficilis lateral view (after Deflandre, 1928a); d – A. discoides foveosa lateral view (after Playfair, 1917); e – A. discoides pseudovulgaris lateral view (after Deflandre, 1928a); f – A. discoides pseudovulgaris arcuata lateral view (after Deflandre, 1928a); g – A. discoides pseudovulgaris tubulata lateral view (after Deflandre, 1928a); i – A. discoides pseudovulgaris undulata lateral view (after Deflandre, 1928a); i – A. discoides scutelliformis lateral view (after Deflandre, 1928a); j – A. discoides scutelliformis lateral view (after Deflandre, 1928a); j – A. discoides scutelliformis lateral view (after Deflandre, 1928a); j, k – A. elliptica lateral (k) and aperture (j) view (after Kufferath, 1932); l, m – A. excavata lateral (m) and aperture (l) view (after Todorov, Golemansky, 2003); n – A. gibbosa lateral view (after Deflandre, 1928a); g – A. gibbosa lateral view (after Deflandre, 1928a); g – A. gibbosa lateral view (after Deflandre, 1928a); g – A. gibbosa lateral view (after Deflandre, 1928a); n – A. gibbosa lateral view (after Deflandre, 1928a); g – A. gibbosa lateral view (after Deflandre, 1928a); g – A. gibbosa lateral view (after Deflandre, 1928a); g – A. gibbosa lateral view (after Deflandre, 1928a); g – A. gibbosa tuberosa lateral view (after Deflandre, 1928a); g – A. gibbosa tuberosa lateral view (after Deflandre, 1928a); g – A. gibbosa tuberosa lateral view (after Deflandre, 1976)



Fig. 4. Genus Arcella:

a, b – A. grospietchi lateral (b) and aperture (a) view (after Štěpànek, 1963); c – A. hemisphaerica lateral view (after Deflandre, 1928a); d – A. hemisphaerica angulata lateral view (after Schönborn, 1962a); e – A. emisphaerica depressa lateral view (after Playfair, 1917); f – A. intermedia laevis lateral view (after Deflandre, 1928a); g – A. intermedia lateral view (after Deflandre, 1928a); h – A. hemisphaerica playfairiana lateral view (Deflandre, 1928a); i – A. hemisphaerica tuberculata lateral view (after Štěpànek, 1963); j – A. hemisphaerica undulata lateral view (after Deflandre, 1928a); k – A. hemisphaerica undulata lateral view (after Deflandre, 1928a); k – A. hemisphaerica undulata lateral view (after Deflandre, 1976); l, m – A. infraterricola lateral (m) and aperture (l) view (after Chardez, 1971); n – A. irregularis lateral view (after Motti, 1961); o – A. jeanneli lateral view (after Virieux, 1916)



Fig. 5. Genus Arcella:

a – A. lichenophila aperture view (after Décloître, 1976); b, c – A. lobostoma lateral (c) and aperture (b) view (after Deflandre, 1928a); d, e – A. maggii lateral (e) and aperture (d) view (after Décloître, 1976); f, i – A. marginata lateral (i) and dorsal (f) view (after Daday, 1905); g, j – A. megastoma lateral (j) and aperture (g) view (after Penard, 1902); h, k – A. megastoma arcuata lateral (k) and aperture (h) view (after Deflandre, 1928a); l, m – A. mitrata lateral (l) and aperture (m) view (after Leidy, 1879); n – A. mitrata pyriformis lateral view (after Deflandre, 1928a)



Fig. 6. Genus Arcella:

a, b – A. multilobata lateral (b) and aperture (a) view (after Golemansky, 1964); c – A. muscicola aperture view (after Décloître, 1976); d, e – A. nordestina lateral (d) and aperture (e) view (after Décloître, 1976); f, g – A. ovaliformis lateral (f) and aperture (g) view (after Chardez, Beyens, 1987); h, i – A. oyei lateral (h) and aperture (i) view (after Štepànek, 1963); j – A. papyracea lateral view (after Playfair, 1914); k, l – A. pentastoma lateral (l) and aperture (k) view (after Deflandre, 1928a); m, p – A. polypora aperture (m) and lateral (p) view (after Deflandre, 1928a); n, q – A. polypora curvata lateral (q) and aperture (n) view (after Décloître, 1976); r – A. polypora undulata lateral view (after Decloire, 1976)



Fig. 7. Genus Arcella:

a, b – A. pygmaea aperture (a) and lateral (b) view (after Bartoš, 1963); c–e – A. rota aperture (c), lateral (d) and aperture-lateral (e) view (after Daday, 1905); f – A. rotundata lateral view (after Playfair, 1917); g – A. rotundata alta lateral view (after Playfair, 1917); h, i – A. rotundata stenostoma lateral (i) and aperture (h) view (after Deflandre, 1928a); j, k – A. rotundata stenostoma undulata lateral (k) and aperture (j) view (after Deflandre, 1928a); l, m – A. tuberosus aperture (l) and lateral (m) view (after Décloître, 1976); n – A. vulgaris lateral view (after Deflandre, 1928a); o, p – A. vulgaris crenulata lateral (p) and aperture (o) view (after Deflandre, 1928a)

Genus Antarcella Deflandre, 1928, emend. Deflandre, 1953

Shell is circular in frontal view, hemispherical in lateral view, aperture is circular, invaginated. In contrast to *Arcella, Antarcella* has one ovular nucleus. Contractive vacuole 10 to 15 μ m in diameter. To distinguish *Antarcella* species from *Arcella* spp., staining of the nucleus is recommended. Ecology: *Sphagnum* mosses. Two species. Type species: *Antarcella atava* (Collin, 1914) (fig. 8).



Fig. 8. *Antarcella atava*: a, c – lateral view; b – apertural view (after Penard, 1917)

Family Netzeliidae Kosakyan et al., 2016

Shell is ovoid, circular in cross-section, aperture is lobed, with a thick organic rim or a necklace made of small idiosomes. Shell is covered by idiosomes, although small sand grains, diatoms or undigested algal cell walls can be used as supplementary building material. These xenosomes are always smoothed and modified by the deposition of silica (Anderson, 1987). The idiosomes often have a nail-like shape. All particles are held in position by perforated cement units and are arranged in a single layer. Outline usually regular, but some species often have protuberances to give a mulberry-like appearance. Recently some spherical *Difflugia* (*D. oviformis, D. tuberculata, D. wailesi, D. tricuspis, D. geospherica,* etc.) that had been found able to produce endogeneous siliceous elements are expected to be removed from *Difflugia*, which is generally incapable to build shell without xenosomes (Ogden, 1979; Gomaa et al., 2017). Type genus: *Netzelia* Ogden, 1979.

Genus Netzelia Ogden, 1979

Corresponds to the description of the family. Ecology: freshwater, *Sphagnum* mosses. Type species: *Netzelia oviformis* (Cash, 1909) (fig. 9).







Fig. 9. Genus Netzelia:

a – N. compressa lateral view (after Dehtiar, 1994); b – N. tuberculata lateral view (after Ogden, 1980b); c – N. wailesi lateral view (after Ogden, Meisterfield, 1989)

Family Cucurbitellidae Gomaa et al., 2017

Shell is ovoid, circular in cross-section, with distinct apertural collar, dark grey or opaque in color, with regular outline, covered by small to medium mineral grains. Organic cement seldom visible as surface structure. Aperture is terminal, with 3 to 12 lobes, composed of small mineral grains. Collar forms a cavity (frontal camera) which has an inner aperture (circular or lobed) on the level with the main body wall. Nuclear is vesicular, cytoplasm of some species contains zoochlorellae. Type genus: *Cucurbitella* Penard, 1902.

Genus Cucurbitella Penard, 1902

Corresponds to the description of the family. Ecology: freshwater. Type species: *Cucurbitella mespiliformis* Penard, 1902 (fig. 10).

Family Centropyxidae Jung, 1942

Bilaterally symmetrical shell, with an eccentric elliptic or circular aperture located on the ventral side; shell with xenosomes.

Key to the genera of the family



Fig. 10. Genus Cucurbitella:

a, b – C. crateriformis aperture (a) and lateral (b) view (after Gauthier-Lièvre, Thomas, 1960); c, d – C. dentata aperture (c) and lateral (d) view (after Gauthier-Lièvre, Thomas, 1960); e, f – C. lunaris aperture (e) and lateral (f) view (after Gauthier-Lièvre, Thomas, 1960); g, h – C. madagascariensis aperture (h) and lateral (g) view (after Gauthier-Lièvre, Thomas, 1960); i, j – C. megastoma aperture (i) and lateral (j) view (after Gauthier-Lièvre, Thomas, 1960); k, 1 – C. mespiliformis aperture (l) and lateral (k) view (after after Gauthier-Lièvre, Thomas, 1960); m, n – C. modesta lateral (m) and aperture (n) view (after Gauthier-Lièvre, Thomas, 1960); o, p – C. obturata aperture (o) and lateral (p) view (after Gauthier-Lièvre, Thomas, 1960); q – C. vlasinensis lateral view (after Ogden, Meisterfield, 1989)

Genus Armipyxis Dekhtiar, 2009

Shell is ovoid, flattened in the lateral view. Aperture is wide, invaginated and slightly eccentric. Anterior walls of the aperture attach to the dorsal side with several appendages creating a complex system of inner partitioning. Shell may or may not be ornamented with spines of varying number and length. Type species: *Armipyxis disoid*es Dekhtiar, 2009 (basionym *Centropyxis discoides* Penard, 1902). The genus also includes *A. mirabilis* (Bartoš, 1940) Dekhtiar, 2009 (basionym *Centropyxis mirabilis* Bartoš, 1940) and *A. gasparella* (Chardez et Beyens, 1988) Dekhtiar, 2009 (basionym *Centropyxis gasparella* Chardez et Beyens, 1988) (fig. 11, 13, 15).

Genus Centropyxis Stein, 1857

Shell is circular, irregular circular of ovoid in apertural (or dorsal) view, flattened in the area of the aperture in lateral view. Aperture is located on ventral side, invaginated, eccentric, circular to ovate. The shell is colorless to brown and may or may not be covered by agglutinating material, which varies from mineral grains to organic debris, particularly diatom frustules. Shell may or may not be ornamented with spines of varying number and length. Ecology: freshwater, moss. Type species: *Centropyxis aculeata* (Ehrenberg, 1838) Stein, 1857 (fig. 12–16).

Genus Conicocassis Nasser et Patterson, 2015

Two-component shell comprised of an ovoid to subspherical main body and a relatively very large conical to funnel-like and asymmetrically positioned flange extending out from a small circular aperture. The lower shell body of *Conicocassis* is ovoid or subspherical to spherical with a wall composed of polymorphous mineral particles within an organic matrix. The colorless to brown main shell body is topped by a cone-like flange, which extends out from a small circular aperture. The cone-like flange is mostly embossed with broken, or intact, diatoms frustules and quartz particles, and appears to be nearly as large as the main shell body if observed from the apertural view. In some cases, the flange may be characterized by coloration distinct from the main shell body. The attachment area of the apertural flange to the main shell body produces a pronounced constriction in the shell wall that is particularly diagnostic (Nasser, Patterson, 2015). Ecology: freshwater, wet moss. Type species: *Conicocassis pontigulasiformis* (Beyens et Chardez, 1986).



Fig. 11. Genera Centropyxis and Armipyxis:

a, b – C. chardezi lateral (a) and aperture (b) view (after Štěpànek, 1963); c, d – C. chardeziella lateral (c) and aperture (d) view (after Laminger, 1973); e – C. compressa aperture view (after Laminger, 1973); f–h – C. constricta aperture (f, h) and lateral (g) view (after Deflandre, 1929); i–k – C. cordobensis aperture (i), dorsal (j) and lateral (k) view (after Vucetich, 1976); 1 – C. decloittella aperture view (after Laminger, 1973); m, n – C. declivistoma lateral (m) and aperture (n) view (after Chardez, 1990); o–q – A. discoides aperture (o, p) and lateral (q) view (p, q – after Penard, 1890; o – after Deflandre, 1929); r – A. discoides solari aperture view (after Štěpànek, 1963)



Fig. 12. Genus Centropyxis:

a, b – C. aculeata aperture (a) and lateral (b) view (after Leidy, 1879); c – C. aculeata dentistoma aperture view (after Chardez, 1970); d – C. aculeata grandis aperture view (after Chardez, 1970); e – C. aculeata lata aperture view (after Chardez, 1970); f – C. aculeata minima aperture view (after Oye, 1958); g – C. aculeata oblonga aperture view (after Deflandre, 1929); h – C. aculeata tropica aperture view (after Chardez, 1970); i–k – C. adami dorsal (i), aperture (j) and lateral (k) view (after Laminger, 1971); l, m – C. aerophila aperture (l) and lateral (m) view (after Deflandre, 1929); n, o – C. aerophila sphagnicola aperture (n) and lateral (o) view (after Deflandre, 1929); r – C. cassis aperture (p) and lateral (q) view (after Deflandre, 1929); r – C. cassis spinifera aperture view (after Deflandre, 1929)



Fig. 13. Genera Centropyxis and Armipyxis:

a-e - C. ecornis apertural (a, b, d, e) and lateral (c) view (a-c after Leidy, 1879; d, e – after Deflandre, 1929); f, g – C. elongata aperture (f) and lateral (g) view (after Lüftenegger et al., 1988); h-j - A. gasparella aperture (h), and fundus (j) view (after Chardez al., lateral (i) et 1988); k, 1 - C. gibba aperture (k) and lateral (l) view (after Deflandre, 1929); m, n – C. grelli aperture (m) and lateral (n) view (after Laminger, 1973); o, p – C. hemisphaerica aperture (o) and lateral (p) view (after Wailes, 1913); q, r -C. hirsuta aperture (q) and lateral (r) view (q – after Deflandre, 1929; r – after Bartoš, 1954)





a, b – C. *invaginata* lateral (a) and aperture (b) view (after Schönborn, 1966a); c, d – C. *janetscheki* aperture (c) and lateral (d) view (after Laminger, 1971); e, f – C. *kurakchaensis* aperture (e) and lateral (f) view (after Snegovaya, Alekperov, 2005); g, h – C. *laevigata* aperture (g) and lateral (h) view (after Schönborn et al., 1983); i, j – C. *latior* aperture (i) and lateral (j) view (after Bartoš, 1963); k, 1 – C. *loffleri* aperture (k) and lateral (l) view (after Laminger, 1972); m–r – C. *marsupiformis* aperture (m, o, q) and lateral (n, p, r) view (after Leidy, 1879); s, t – C. *marsupiformis obesa* aperture view (after Leidy, 1879)



Fig. 15. Genera Centropyxis and Armipyxis:

a, b – C. minuta aperture (a) and lateral (b) view (after Deflandre, 1929); c, d – A. mirabilis lateral (c) and aperture (d) view (after Bartoš, 1940); e, f – C. notonyx lateral (e) and aperture (f) view (after Jung, 1942); g–j – C. orbicularis aperture (g, h) and lateral (h, j) view (g, h – after Lüftenegger et al., 1988; i, j – after Deflandre, 1929); k, 1 – C. percolabiensis aperture (k) and ventral-lateral (l) view (after Dekhtiar, 1994); m, n – C. percolabiensis inermis aperture (m) and ventral-lateral (n) view (after Dekhtiar, 1994); o–r – C. platystoma aperture (o, q) and lateral (p, r) view (o, p – after Leidy, 1879; q, r – after Deflandre, 1929); s–t – C. platystoma armata aperture (s) and lateral (t) view (after Deflandre, 1929); u – C. pytiformis aperture view (after Oye, 1958)


Fig. 16. Genus Centropyxis:

a–c – *C. recurvata* aperture (a), dorsal (b) and lateral (c) view (after Vucetich, 1976); d, e – *C. sacciformis* lateral (d) and aperture (e) view (after Décloître, 1954); f, g – *C. spinosa* aperture (f) and lateral (g) view (after Deflandre, 1929); h, i – *C. sylvatica* aperture (h) and lateral (i) view (after Lüftenegger et al., 1988); j, k – *C villiersi* aperture (j) and lateral (k) view (after Décloître, 1954)

Genus Proplagiopyxis Schönborn, 1964

Shell is ovoid in frontal view, hemispheric with a slight slope to the aperture in lateral view, brown, xenosomes lacking or rare. Aperture is eccentric, circular, no invagination. Ecology: soil. Monospecific. Type species: *Proplagiopyxis nuda* Schönborn, 1964 (fig. 17).



Fig. 17. *Proplagiopyxis nuda*: a – lateral view; b – aperture view (after Schönborn, 1964)

Family Cryptodifflugiidae Jung, 1942

Shell is hyaline, surface organic or with attached mineral particles. Aperture is terminal or eccentric.

Key to the genera of the family

1. Shell is radially symmetrical, aperture is termin	al
Gen	us Cryptodifflugia
1'. Shell is bilaterally symmetrical	
2. Aperture is situated on a well-developed or poo	orly
expressed neck inclined ventrallyGe	enus Meisterfeldia

2'. Aperture is a simple perforation in the shell.... Genus Wailesella

Genus Cryptodifflugia Penard, 1890

Shell is oval, egg-shaped, pyriform with a short neck, circular or oval cross-section, with adhering foreign particles or smooth surface, colorless, yellow or brown, composed of an outer proteinaceous material usually lined, aperture terminal, circular or oval. Shell walls of some species with two distinct layers: outer surface is think organic, inner layer thick from calcified material (Hedley et al., 1977). Genus *Difflugiella* (Cash, 1904) is now considered to by a synonym of *Criptodifflugia* (Page, 1966). Ecology: freshwater, moss, soil. Type species: *Cryptodifflugia oviformis* Penard, 1890 (fig. 18–24).



Fig. 18. Genus Cryptodifflugia:

a – *C. angulata* lateral view (after Playfair, 1917); b–d – *C. angusta* lateral view (b, c), apertural view (d) (after Schönborn, 1965a); e–f – *C. angustatostoma* narrow side lateral view (e), broad side lateral view (f) (after Beyens, Chardez, 1982); g – *C. apiculata* lateral view, living specimen with cytoplasm (after Cash, 1904); h–i – *C. bassini* lateral view (h), apertural view (i) (after Bobrov, 2001). Scale bar 20 μ m



Fig. 19. Genus Cryptodifflugia:

a–c – *C. brevicolla* broad side lateral view (a), narrow side lateral view (b), living specimen with cytoplasm (c) (after Golemansky, 1979); d–e – *C. collum* apertural view (d), lateral view of living specimen with cytoplasm (e) (after Chardez, 1971). Scale bar 20 μ m



Fig. 20. Genus Cryptodifflugia:

a-f – C. compressa broad side lateral view (a, d), narrow side lateral view (b, e), apertural view (c, f) (after Penard, 1902); g-h – C. compressa angustioris narrow side lateral view (g), broad side lateral view (h) (after Tarnogradsky, 1959); i–k – C. compressa australis broad side lateral view (i), narrow side lateral view (j), apertural view (k) (after Palyfair, 1918); 1 – C. compressa ovata lateral view (after Palyfair, 1918). Scale bar 20 μ m



Fig. 21. Genus Cryptodifflugia:

a – *C. crenulata* lateral view (after Palyfair, 1918); b – *C. crenulata glabra* lateral view (after Palyfair, 1918); c – *C. crenulata globosa* lateral view (after Palyfair, 1918); d–f – *C. horrida* lateral view of empty shell (d), lateral view of living cell (e), apertural view of living cell (f) (after Schönborn, 1965a). Scale bar 20 μ m





a–c – *C. lanceolata* broad side lateral view (a), narrow side lateral view (b), apertural view (c) (after Golemansky, 1970a); d – *C. leachi* lateral view of living cell (after Nicholls, 2006a); e–f – *C. minuta* lateral view (after Palyfair, 1918); g–k *C. oviformis* lateral view of encysted specimen (g), lateral view (i), apertural view (h, j), lateral view of encysted specimen (k) (g–h – after Page, 1966, i–k – after Penard, 1890). Scale bar 20 μ m



Fig. 23. Genus Cryptodifflugia:

a–b – *C. oviformis fusca* lateral view of different specimens (after Penard, 1890); c–e – *C. paludosa* narrow side lateral view (c), apertural view (d), broad side lateral view (e) (after Golemansky, 1981); f–g – *C. patinata* lateral view (f), apertural view (g) (after Schönborn, 1965a); h – *C. psammophila* lateral view (after Golemansky, 1970a); i – *C. pusilla* lateral view (after Palyfair, 1917); j – *C. pusilla* var. *conica* lateral view (after Palyfair, 1917). Scale bar 20 μ m



Fig. 24. Genus Cryptodifflugia:

a – *C. sacculus* lateral view (after Penard, 1902); b–c – *C. sacculus* sakotschawi lateral view (b), apertural view (c) (after Tarnogradsky, 1959); d–e – *C. splendida* lateral view of different specimens (after Schönborn, 1965a); f – *C. valida* lateral view (after Penard, 1902); g–h – *C. voigti* lateral view (g), apertural view (h) (after Schmidt, 1926); i – *C. vulgaris* lateral view (after Francé, 1913). Scale bar 20 μ m

Genus Meisterfeldia Bobrov, 2016

Shell is ovoid, more or less laterally compressed; colorless, yellow or brown, composed of proteinaceous material without mineral particles. Aperture circular, subterminal, placed on ventrally and obliquely cut apertural end, or it is situated on a well developed or poorly expressed neck inclined ventrally; sometimes aperture border shows a slight swelling. Ecology: wet mosses, *Sphagnum* mosses, litter, soil. Type species: *Meisterfeldia chibisovi* Bobrov, 2016 (fig. 25).



Fig. 25. Genus Meisterfieldia:

a, b – *M. chibisovi* lateral (a) and ventral (b) view (after Bobrov, 2016); c, d – *M. wegeneri* lateral (c) and ventral (d) view (after Bobrov, 2016); e, f – *M. polygonia* lateral (e) and ventral (f) view (after Bobrov, 2016); g, h – *M. vanhoornei* lateral (g) and vebtral (h) view (after Beyens et al., 1986)

Genus Wailesella Deflandre, 1928

Shell is ovate, chitinoid, in lateral view truncated in the area of aperture; brown or yellow-brown. Aperture is subterminal, circular. Ecology: dry mosses, litter, soil. Monospecific. Type species: *Wailesella eboracensis* Wailes, 1911 (fig. 26).



Fig. 26. *Wailesella eboracensis*: a – apertural view; b – lateral view (after Deflandre, 1928b)

Family Distomatopyxidae Bonnet, 1970

Shell is circular or ovoid in frontal view, hemispheric in lateral view. Aperture is central, elliptical, invaginated and lying at the bottom of a tubular vestibule. The vestibule is partly covered by a diaphragm which is fixed by two bridges forming two crescent-shaped openings at opposite sites. Shell is covered with xenosomes.

Genus Distomatopyxis Bonnet, 1964

Shell morphology with characters of the family; covered by mineral particles in an organic cement, smooth surface. Ecology: soils. Type species: *Distomatopyxis couillardi* Bonnet, 1964 (fig. 27).



Fig. 27. *Distomatopyxis couillardi*, lateral view (after Bonnet, 1964)

Family Hyalospheniidae Schultze, 1877, emend. Kosakyan et Lara, 2012

Shell is rigid, colourless or yellowish-brown, flask-vase shaped, oval or pyriform (can be circular or elongated), generally compressed, rarely circular in cross section (e.g. *Longinebela golemanski, Padaungiella nebeloides*). The shell is either entirely self-secreted composed of an organic matrix, or with addition of self-secreted siliceous plates or recycled shell plates of euglyphids, *Quadrulella* or other similar material such as diatom frustules incorporated in the shell. Aperture is terminal and is bordered by a more or less thin organic lip (rim, collar or fringe). Currently, family includes thirteen genera.

Key to the genera of the family

3. Shell is entirely composed of square plates4

4. Hollow (not flat short keel as in *Quadrulella alata*) lateral keel is present (note: the origin of square plates is questionable).....

4'. Hollow lateral keel is absent (square plates are self secreted)...... Genus *Quadrulella*

6'. Shell with similar shape (or more pyriform), lacking pointed protuberances in the internal side of the neck Genus *Porosia*

8'. No constriction at the base of the neck Genus Padaungiella

9. Shell with pair of lateral conical expansions (horns, that can be external, or internal connected with each other with complete or partial horseshoe-like keel) protruding on either side...... Genus *Cornutheca*

10'. Shell is elongated pyriform or oval-elongated, lateral sides tapering toward the aperture (generally the length of the shell

is > 140	um, with	exception of	Gibbocarina	gracilis
----------	----------	--------------	-------------	----------

$(L = 90-130 \ \mu m)$	
11. Shell with lateral keel	
11'. Lateral keel is absent	Genus Longinebela
12. Lateral keel is flat	Genus <i>Planocarina</i>
12'. Lateral keel is hollow	Genus <i>Gibbocarina</i>

Genus Alocodera Jung, 1942

Shell is pyriform, compressed in cross section, with a welldeveloped neck separated from the posterior part of the shell by two lateral indentations. Two lateral pores are situated in the indentations. Shell very transparent, yellowish, smooth, covered by small xenosomes. Aperture is arched in the narrow lateral view, surrounded by a thickened rim. Ecology: mosses. Monospecific. Type species: *Alocodera cockayni* (Penard, 1902) (fig. 28).



Fig. 28. *Alocodera cockayni*: broad lateral view (after Deflandre, 1936)

Genus Apodera Loeblich et Tappan, 1961

Shell is subspherical or ellipsoidal, compressed in cross section, with a distinct neck separated from the rest of the shell by constriction. The neck tapers from the body towards the aperture. The whole shell is covered by is circular and oval plates of scales, distributed more or less regularly. Aperture is terminal, oval, slightly arched, surrounded by an organic rim. Ecology: mosses, organic soils. Type species: *Apodera vas* Certes, 1889 (fig. 29).



Fig. 29. *Apodera vas*: a, b – lateral view (a – after Certes, 1889; b – after Penard, 1911)

Genus Certesella Loeblich et Tappan, 1961

Shell is pyriform elongated or flask-shaped, compressed in cross section, neck with parallel sides, organic apertural rim. The development of the neck varies among species from slightly to well differentiated. The main characteristics of the genus are the presence of two lateral depressions with large central pores connected by two tubes located at approximately 2/3 of the distance between the fundus of the shell and the aperture, and the presence of internal teeth on the neck giving punctuated impression. Shell composed of collected euglyphid idiosomes in an unstructured cement, transparent. Type species: *Certesella martiali* (Certes, 1889) (fig. 30).

Genus Cornutheca Kosakyan et al., 2016

Shell is elongated-pyriform, with a distinct neck, lateral margins tapering towards the aperture. Two lateral horns, pointing towards the posterior part of the shell, either free or connected to the main part of the shell by a lateral keel surrounding the posterior part of the shell. Shell hyaline or slightly yellowish, composed of circular to elongated shell plates probably recycled from euglyphid testate amoeba prey. Members of this genus differ from other hyalospheniid genera in the presence of two lateral horns. Type species: *Cornutheca ansata* (Leidy, 1879) (fig. 31).



Fig. 30. Genus Certesella:

a, b – C. certesi lateral view (a – after Certes, 1889; b – after Deflandre, 1936); c–f – C. martiali broad (c, e, f) and narrow (d) lateral view (c – after Certes, 1889; d, e – after Penard, 1911; f – after Deflandre, 1936); g, h – C. murrayi broad (h) and narrow (g) lateral view (after Wailes, 1913)



Fig. 31. Genus Cornutheca:

a – C. ansata lateral view (after Leidy, 1879); b – C. equicalceus lateral view (after Leidy, 1879); c, d – C. saccifera broad (d) and narrow (c) lateral view (after Wailes, 1913)

Genus Gibbocarina Kosakyan et al., 2016

Shell is elongated-pyriform, with the lateral sides tapering towards the aperture, with a hollow tuberous keel surrounding the entire posterior end of the shell. Shell hyaline or slightly yellowish, composed of circular to elongated shell plates, probably recycled from euglyphid testate amoeba prey. Members of this genus differ from those in the genus *Nebela* by the strongly elongated shape of the shell, the presence of a hollow keel and in most cases a larger size. From the members *of Longinebela*, they differ mainly by the presence of a keel. *Gibbocarina* may be confused with *Planocarina*, although in the latter the keel is flat, while in *Gibbocarina* it is hollow (the difference is very obvious when seen in profile). Type species: *Gibbocarina galeata* (Penard, 1890) (fig. 32).



Fig. 32. Genus *Gibbocarina*: a, b – *G. galeata* broad (a) and narrow (b) lateral view (after Deflandre, 1936); c, d – *G. gracilis* broad (d) and narrow (c) lateral view (after Deflandre, 1936)

Genus Hyalosphenia Stein, 1859

Shell is rounded, ovoid or elongated elliptical or flask shaped, laterally compressed, aperture variable from linear to strongly curved, with or without thickened lip. Shell hyaline or slightly yellowish, with a smooth organic surface (exception: *Hyalosphenia punctata* which has a punctuated surface). Some species contain symbiotic zoochlorellae in cytoplasm. Ecology: freshwater, moss, soils. Type species: *Hyalosphenia ligata* (Tatem, 1870) (fig. 33–34).



Fig. 33. Genus Hyalosphenia:

a, b – *H. angulata* broad (a) and narrow (b) lateral view (after Schouteden, 1905); c, d – *H. cuneata* broad (d) and narrow (c) lateral view (after Penard, 1902); e, f – *H. elegans* broad (f) and narrow (e) lateral view (Penard, 1902); g, h – *H. elegans cylindricollis* broad (h) and narrow (g) lateral view (after Chardez, 1962); i, j – *H. gigantea* broad (i) and narrow (j) lateral view (after Graaf, 1952); k, 1 – *H. inconspicua* broad (l) and narrow (k) lateral view (after West, 1903); m, n – *H. insecta* broad (m) and narrow (n) lateral view (after Cash, Hopkinson, 1909); o – *H. jirovici* lateral view (after Cash, Hopkinson, 1909); r – *H. mraconia* lateral view (after Godeanu, 1972); s, t – *H. ovalis* broad (s) and narrow (t) lateral view (after Cash et al., 1919)



Fig. 34. Genus Hyalosphenia:

a–c – *H. papilio* broad (a, c) and narrow (b) lateral view (a, b – after Leidy, 1879; c – after Penard, 1902); d – *H. papilio stenostoma* lateral view (after Deflandre, 1931); e – *H. punctata* lateral view (after Penard, 1902); f, g – *H. penardi* broad (g) and narrow (f) lateral view (after Lauterborn, 1908); h, i – *H. platystoma* broad (h) and narrow (i) lateral view (after Cash, Hopkinson, 1909); j – *H. schoutedeni* lateral view (after Oye, 1926); k – *H. schoutedeni* rotundata lateral view (after Oye, 1958); 1–*H. subflava* lateral view (after Bonnet, Thomas, 1960)

Genus Mrabella Kosakyan et al., 2016

Shell is elongated-pyriform, laterally compressed, with the lateral sides gradually tapering toward the aperture. Lateral margins distinctly compressed giving the impression of a thick and wide hollow keel. Because of this, the shell is elongated elliptical with the pointed end in the profile. Shell colourless, composed of quadrangular shell plates similar to those in *Quadrulella* species. Type species: *Mrabella subcarinata* (Gauthier-Lièvre, 1957) (fig. 35).



Fig. 35. Genus Mrabella:

a, b – M. plicata narrow (a) and broad (b) lateral view (after Hoogenraad, de Groot, 1940b); c, d – M. subcarinata narrow (c) and broad (d) lateral view (after Gauthier-Lièvre, 1957)

Genus Nebela (Leidy, 1874) Kosakyan et al., 2016

Shell is rounded, ovoid-pyriform, or wide-pyriform, rarely with a keel (partial or complete) or other lateral expansions, with or without wavy lateral margins. Aperture is ranging from linear to strongly curved, bordered by a more or less thin organic lip (rim, collar or fringe). Shell is hyaline or slightly yellowish, reinforced with circular to elongated shell plates apparently recycled mostly from euglyphid testate amoeba prey, sometimes also including fragments of diatom frustules or other small mineral elements. Type species: *Nebela collaris* (Ehrenberg, 1848) Leidy, 1879 (fig. 36).



Fig. 36. Genus Nebela:

a, b – *N. carinatella* broad (b) and narrow (a) lateral view (after Beyens, Chardez, 1982); c, d – *N. collaris* broad (d) and narrow (c) lateral view (after Penard, 1902); e, f – *N. flabellum* broad lateral view (e – after Cash, Hopkinson, 1909; f – after Deflandre, 1936); g–i – *N. militaris* broad (h, i) and narrow (g) lateral view (after Penard, 1890); j–l – *N. penardiana* broad (j, l) and narrow (k) lateral view (after Deflandre, 1936); m – *N. tincta* lateral view (after Penard, 1890)

Genus Longinebela Kosakyan et al., 2016

Shell is elongated-pyriform, with a distinct neck, lateral margins (that can be straight or wavy) tapering towards the aperture. Shell is hyaline or slightly yellowish, composed of circular to elongated shell plates probably recycled from euglyphid testate amoeba prey. Type species: *Longinebela tubulosa* (Penard, 1902) (fig. 37).



Fig. 37. Genus *Longinebela*: a – *L. spesiosa* lateral view (after Deflandre, 1936); b, c – *L. tubulosa* broad (b) and narrow (c) lateral view (after Penard, 1902)

Genus Padaungiella Lara et Todorov, 2012

Shell is bottle-shaped, compressed, with a distinct elongated. Shell is covered by idiosomes of testate amoeba prey, various forms, randomly located. Aperture is elliptic or oval, surrounded by organic rim or straight cut. Ecology: freshwater, mosses. Type species: *Padaungiella lageni-formis* (Penard, 1890) (fig. 38).



Fig. 38. Genus Padaungiella:

a ,b – *P. lageniformis* lateral view (a – after Penard, 1902; b – after Deflandre, 1936); c–e – *P. tubulata* broad (c), narrow (d) lateral and aperture (e) view (after Lüftenegger, Fiossner, 1991); f – *P. wailesi* lateral view (after Deflandre, 1936); g – *P. wetekampi* lateral view (after Jung, 1942)

Genus Planocarina Kosakyan et al., 2016

Shell is elongated-pyriform, with a distinct neck, lateral margins tapering towards the aperture. A flat keel surrounds the entire posterior part of the shell. Shell is hyaline or slightly yellowish, composed of circular to elongated shell plates probably recycled from euglyphid testate amoeba prey. Type species: *Planocarina carinata* (Archer, 1867) (fig. 39).



Fig. 39. Genus Planocarina:

a-c - P. carinata broad (a, c) and narrow (b) lateral view (a, b – after Cash, Hopkinson, 1909; c – after Deflandre, 1936); d, e – P. marginata broad (d) and narrow (e) lateral view (after Deflandre, 1936); f, g – P. maxima broad (g) and narrow (f) lateral view (after Awerintzew, 1907); h, i – P. spumosa broad (i) and narrow (h) lateral view (after Awerintzew, 1907)

Genus Porosia (Jung, 1942) Bobrov et Kosakyan, 2015

Shell is pyriform, with rounded posterior end, laterally compressed. In front view two distinct lateral depressions with two large invaginated pores are situated on each side, which are connected by internal tubes as in genus *Certesella*. In profile, small lateral pores can be observed, just anterior to the large pores. The lateral keel can (or cannot) be present surrounding 1/3 of posterior lateral margin (keel is important distinctive character between *Porosia* species: *Porosia bigibbosa* – lacking of lateral keel, *Porosia paracarinata* – presence of lateral keel). Shell is composed of euglyphid shell plates embedded in unstructured cement. Aperture is curved, surrounded with organic lip. Habitat: *Sphagnum* mosses, litter, soil, rare genus. This genus is closely related to *Certesella* but lacks the punctuated neck. Type species: *Porosia bigibbosa* Jung, 1942 (fig. 40).



Fig. 40. *Porosia bigibossa*: a-c - broad (a, c) and narrow (b) lateral view (a - after Penard, 1890; b, c - after Wailes, Penard, 1911)

Genus Quadrulella (Cockerell, 1909) Kosakyan et al., 2016

Shell is pyriform, or elongated pyriform, with the lateral sides tapering towards the aperture, with or without distinct neck. Always laterally compressed, elliptical in cross section. Aperture ranging from linear to strongly curved, with or without thickened organic lip. Shell is hyaline, composed of self-secreted square plates. *Quadrulella* differs from *Nebela* and *Hyalosphenia* by the use of square, siliceous self-secreted plates in the construction of the shell. It may be confused with *Mrabella*, which has similar shell shape and square plates. The most distinctive characteristic is the presence of a lateral pronounced hollow keel in the genus *Mrabella* (although *Q. alata* also exhibits a lateral keel, we consider it to be flat and not hollow, even if this detail is not mentioned in the original description of the species), and the fact that square plates of *Quadrulella* are self-secreted, while the origin of square plates of *Mrabella* is unclear. Type species: *Quadrulella symmetrica* (Wallich, 1864) Cockerell, 1909 (fig. 41).

Family Lamtopyxidae Bonnet, 1974

Shell is circular in frontal view; ventral surface is flat, dorsal surface is hemispheric. External aperture is bordered by large teeth, internal opening at the end of deeply invaginated tube, elliptic. Shell is covered with xenosomes.

Genus Lamtopyxis Bonnet, 1974

External opening with three to five teeth. The base of apertural tube is reinforced by a more or less quadratic organic frame; the internal opening is bordered by a collar. Shell is composed of flat mineral particles which are held together by unstructured organic cement giving the shell a smooth surface. Ecology: tropical forest soils. Type species: *Lamtopyxis callistoma* Bonnet, 1974 (fig. 42).



Fig. 41. Genus Quadrulella:

a – Q. acuminata lateral view (after Oye, 1958); b, c – Q. alata narrow (b) and broad (c) lateral view (after Gauthier-Lièvre, 1957); d, e – Q. camerounensis narrow (d) and broad (e) lateral view (after Gauthier-Lièvre, 1957); f – Q. debonti lateral view (after Gauthier-Lièvre, 1957); g, h – Q. elegans narrow (g) and broad (h) lateral view (after Gauthier-Lièvre, 1953); i – Q. elongata lateral view (after Chardez, 1967b); j – Q. lageniformis lateral view (after Oye, 1949); k – Q. quadrigera lateral view (after Deflandre, 1936); l, m – Q. scutellata lateral view (after Deflandre, 1936); n, o – Q. symmetrica lateral view (after Deflandre, 1936); p, q – Q. tropica broad (p) and narrow (q) lateral view (after Wailes, 1912); r, s – Q. tubulata narrow (r) and broad (s) lateral view (after Gauthier-Lièvre, 1953)



Fig. 42. *Lamtopyxis callistoma*: a – aperture view; b – lateral view (after Bonnet, 1974)

Family Microchlamyiidae Ogden, 1985, emend. Kudryavtsev et Hausmann, 2007

Shell is proteinaceous, flexible or rigid, finely areolate. Cytoplasm either enclosed in a separate membrane sac with single aperture, attached to the main shell, or the peripheral part of the shell membranous with a single aperture, in which case there is no separate membrane sac between the cell body and shell.

Key to the genera of the family

1. Cytoplasm enclosed in a separate membrane sac with an aperture, attached to the main shell Genus *Microchlamys*

Genus Microchlamys Cockerell, 1911

Flexible organic shells, finely areolated, chitinoid, foldede at the ventral side; transparent, yellow to brown in color. Cell enclosed within a membranous sac, which is fused to the shell at intervals, lost in empty shells. This membrane closes the shell from the ventral side and forms an aperture for the lobopodia. Ecology: freshwater, mosses, soil. Type species: *Microchlamys patella* (Claparéde et Lachmann, 1859) (fig. 43).

Genus Spumochlamys Kudryavtsev et Hausmann, 2007

Flexible organic shell with a spongious dorsal part, which becomes thinner towards the margin and ends in a delicate membrane that partially covers the cytoplasm on the ventral side, leaving an aperture through which some cytoplasm can extend. Ecology: brackish waters. Type species: *Spumochlamys iliensis* Kudryavtsev et Hausmann, 2007.



Fig. 43. *Microchlamys patella*: a – dorsal view; b – lateral view (after Penard, 1902)

Family Microcoryciidae de Saedeleer, 1934

Protenaceous, flexible or semi-rigid shells, not areolate; aperture ventral, distinct, variable.

Key to the genera of the family

1. External layer of shell is gelatinous; usually one nucleus; plasm	a
violet; aperture invaginatedGenus Amphizonel	lla
1'. Shell is not gelatinous	. 2
2. Aperture is slit-like	. 3
2'. Aperture of variable shape or circular, sometimes wide open	•••
· · · · · · · · · · · · · · · · · · ·	.4
3. Shell is pyriform or discoid, plasma violet Genus Zonomy.	xa
3'. Shell is laterally compressed, ovoid in oral view	
	na
4. Dorsal face with mineral particle or attached debris	. 5
4'. Dorsal face without foreign materials, shell is smooth	•••
Genus Penardochlam	ys
5. Shell is hemispheric, bilayered; flexible, wide, ventral aperture.	
	ys
5'. Dorsal side is more or less rigid, ventral like a flexible skirt	
	ea
•	

Genus Amphizonella Greeff, 1866

Shell is more or less round, bilayered pellicle, out layer gelatinous $8-12 \mu m$ with fine denticles; inner layer is thin, chitinoid, undulates with internal movements, sac-like. Aperture is invaginated, variable. Pseudopods cyllindroid, finely granular, rounded ends. Movement

sluggish. Endoplasm clear, violet; yellow granules in purple vesicles. One ovular nucleus. Ecology: mosses on trees. Monospecific. Type species: *Amphizonella violacea* Greef, 1866 (fig. 44).



Fig. 44. *Amphizonella violacea*: a – lateral view (after Penard, 1902); b – dorsal view (after Penard, 1906)

Genus Diplochlamys Greeff, 1888

Shell is small, round, grayish-yellow, bilayered; inner hyaline sac enclosing the cell with flexible aperture, outer layer consists of loosely arranged debris. Ecology: mosses, soils. Type species: *Diplochlamus leidyi* Greef, 1888 (fig. 45).



Fig. 45. *Diplochlamys fragilis*: a – lateral view; b – apertural view (after Penard, 1909)

Genus Microcorycia Cockerell, 1911

Shell is flexible and comprised of two parts: upper and lower. The upper shell part is dome-shaped, sometimes with concentric ridges or small horns at the dorsal side; thick and less flexible walls than the lower shell part; maybe covered with small debris or mineral particles. The lower shell part consists of a thin, transparent skin, which shows no structure in the light microscope, can be folded up and is open downwards for extruding pseudopodia. The skin only loosely covers the cell body and can be contracted like a sack. There is a zone of gradual transition from the upper shell part to the lower shell part. Ecology: mosses, soils, freshwater. Type species: *Microcorycia flava* (Greef, 1866) Penard, 1902 (fig. 46).



Fig. 46. Microcorycia flava:

a – lateral view, lower part of the shell open; b – lateral view, lower part of the shell closed; c – ventral view, lower part of the shell closed (after Penard, 1902)

Genus Parmulina Penard, 1902

Shell is ovoid in aperture view, laterally compressed, hemispheric in lateral view; clear, flexible, chitionoid, opaque with attached debris. Aperture is slit-like. Pseudopods rare, bluntly lobate. Endoplasm granular. One vesicular nucleus. Ecology: mosses on trees. Type species: *Parmulina cyathus* Penard, 1902 (fig. 47).



Fig. 47. *Parmulina cyathus*: a – lateral view; b – dorso-lateral view; c – dorsal view (after Penard, 1902)

Genus Penardochlamys Deflandre, 1953

Shell is round, wavy outline, chitionoid, flexible, punctuate. Aperture is circular or wavy, rarely visible, slightly invaginated. Pseudopods cylindroid, finely granular, round ends. Endoplasm granular. Two vesicular nuclei; one or two contractive vacuoles. Ecology: freshwater. Monospecific. Type species: *Penardochlamys arcelloides* (Penard, 1904) (fig. 48).

Genus Zonomyxa Nüsslin, 1882

Shell is large, resting form is discoid, locomotive form is pyriform; chitinoid pellicle, without mucilaginous envelope, flexible, follows the movements of the cell. Surface with small temporal perforations trough which plasma threads emerge. Pseudopods single, clear, conical, from slit-like aperture. Endoplasm granular, violet tinted. Ecology: *Sphagnum* or aquatic vegetation. Monospecific. Type species: *Zonomyxa violacea* Greef, 1866 (fig. 49).



Fig. 48. *Penardochlamys arcelloides*: lateral view (after Penard, 1909)



Fig. 49. Zonomyxa violacea (after Penard, 1906)

Family Plagiopyxidae Bonnet et Thomas, 1960

Shell is bilaterally symmetrical; aperture is on ventral side, eccentric, cryptostome. Shell is covered by xenosomes.

Key to the genera of the family

1. Anterior lip of aperture is a distinct overhanging hood
1'. Aperture is more or less slit-like, no overhanging hood
2. Anterior lip of aperture with pores Genus Bullinularia
2'. No such pores
3. Anterior (dorsal) lip of aperture curves below ventral face
3'. Posterior (ventral) lip of aperture curves below anterior
(dorsal) lip, or aperture is an open slit
4. Aperture is an open slit Genus Protoplagiopyxis
4'. Aperture is covered by the anterior lip

ted to the dorsal side of the shell	5. Posterior (venteral) lip conne
rds anterior lip	from inside and has an extension tow
Genus Paracentropyxis	
lip Genus Plagiopyxis	5'. No extension of the posterio
Genus Hoogenraadia	6. Ventral face is rounded
Genus Planhoogenraadia	6'. Ventral face is flattened

Genus Bullinularia Deflandre, 1953

Shell is ovoid or circular in front view, hemispherical with flattened ventral surface in lateral view; covered by small mineral particles in sheet-like organic cement, smooth, dark brown and opaque. Aperture is eccentric, invaginated, completely or partially hidden by the dorsal apertural lip (cryptostome). A key character is the pores on the dorsal lip, on the apex and, depending on the species, on the ventral side. Ecology: soils, mosses. Type species: *Bullinularia indica* (Penard, 1907) (fig. 50).



Fig. 50. Genus Bullinularia:

a, b – B. *indica* aperture (a) and lateral (b) view (after Geltzer et al., 1995); c, d – B. *minor* aperture view (after Hoogenrad et al., 1948)

Genus Geoplagiopyxis Chardez, 1961

Shell is ovoid in frontal view, hemispheric in lateral view; ventral surface slightly vaulted; dorsal face curving below the convex ventral face. Aperture is not easily visible as an irregular slit. Shell composed of amorphous plates without larger mineral particles. Ecology: soil. Type species: *Geoplagiopyxis declivis* Chardez, 1961 (fig. 51).



Fig. 51. *Geoplagiopyxis declivus*: a – apertural view; b – lateral view (after Chardez, 1960)

Genus Hoogenraadia Gauthier-Lièvre et Thomas, 1958

Shell is ovoid-globular, overhanging anterior visor with two lateral groves, incurved ventral lip; covered by xenosomes. Ventral face circular. Aperture is more or less circular, partly covered by visor. Ecology: tropical bogs, swamps, soils. Type specie: *Hoogenraadia africana* Gauthier-Lièvre et Thomas, 1958 (fig. 52).

Note: Chardez (1963) has erected a similar perhaps synonymous genus *Gillardella*, which has a much smaller visor.

Genus Paracentropyxis Bonnet, 1960

Shell is circular or slightly elliptical in frontal view; at the dorsal face, a groove separates the reminder of the shell from a small visor in from of tapered crescent. Ventral surface is concave; at median level the visor is rounded, forming a buccal cavity, ventral face and the side walls of this vestibulum are connected to the dorsal part of the shell; with small internal opening as in *Centropyxis sylvatica*. The slit-like aperture is hidden by the dorsal lip and an extension of the ventral shell wall (cryptostomia). Shell transparent and composed of exogenous mineral particles, which are embedded in a hyaline cement. Ecology: tropical soils. Monospecific. Type species: *Paracentropyxis mimetica* Bonnet, 1960 (fig. 53).

Note: Dekhtiar (2009) placed the genus *Paracentropyxis* in the family Centropyxidae with species *Paracentropyxis sylvatica* (Deflandre, 1929) Dekhtiar, 2009 (basionym *Centropyxis sylvatica* Deflandre, 1929) and *Paracentropyxis matthesi* (Rauenbusch, 1987) Dekhtiar, 2009 (basionym *Centropyxis matthesi* Rauenbusch, 1987).



Fig. 52. *Hoogenradia africana:* a – lateral view; b – apertural view (Gauthier-Lièvre, Thomas, 1958)



Fig. 53. *Paracentropyxis mimetica*: a – apertural view; b – lateral view (after Bonnet, 1960a)

Genus Plagiopyxis Penard, 1910

Shell is circular of oval in frontal view, hemispheric in lateral view. Aperture is elongate slit, perpendicular to long axis of the shell; dorsal aperture lip somewhat incurved usually hiding the opening. Ecology: most of the species are edaphobionts (soil-inhabited), some freshwater. Type species: *Plagiopyxis declivis* Bonnet et Thomas, 1955 (fig. 54).



Fig. 54. Genus Plagiopyxis:

a, b – P. callida aperture (a) and lateral (b) view (after Geltzer et al., 1995); c, d – P. declivis aperture (c) and lateral (d) view (after Geltzer et al., 1995); e, f – P. penardi aperture (e) and lateral (f) view (after Geltzer et al., 1995); g – P. labiata aperture view (after Thomas, 1958a)

Genus Planhoogenraadia Bonnet, 1977

Shell is ovoid in frontal view, hemispheric with flat ventral surface and a visor-like extension separated by two groves from the mina body of the shell in lateral view. Aperture is semicircular or elongated oval, located back into the shell, partly covered by the visor-like extension of the dorsal surface. Ecology: soils. Type species: *Planhoogenraadia acuta* Bonnet, 1977 (fig. 55).

Genus Protoplagiopyxis Bonnet, 1962

Shell is ovoid in frontal view, hemispheric with anterior flattening in lateral view; covered by extraneous mineral particles. Ventral surface is flattened very clearly and slightly or not inclined. Aperture is a straight or crescent-shaped slit of "plagiostome" type with tendency to "cryptostome" (aperture located inside the shell), no vestibule, but apertural region is well separated by fairly abrupt dorso-ventral depression in the anterior part. Ecology: soils. Type species: *Protoplagiopyxis delamarei* Bonnet, 1962 (fig. 56).



Fig. 55. *Planhoogenradia acuta*: a – apertural view; b – lateral view (Bonnet, 1977)

Fig. 56. *Protoplagiopyxis delamarei*: a – lateral view; b – apertural view (Bonnet, 1962)

Family Paraquadrulidae Deflandre, 1953

Shells are composed of endogenously formed rectangular calcite plates embedded in a sheet-like organic cement. The presence of calcite still should be verified.

Key to the genera of the family

Genus Paraquadrula
a bended neck
Genus Lamtoquadrula

Genus Lamtoquadrula Bonnet, 1974

Shell is retort-shaped with bending neck in lateral view; circular cross-section; covered by endogenously secreted square or rectangular plates placed in regular rows. Aperture is circular with a thin transparent organic lip, only visible with phase or interference contrasts. Ecology: soils. Monospecific. Type species: *Lamtoquadrula deflandrei* Bonnet, 1974 (fig. 57).

Genus Paraquadrula Deflandre, 1932

Shell is transparent, broad oval, slightly compressed laterally; covered by endogenously secreted square or rectangular plates placed in regular rows. Aperture is terminal, slit-like or irregular. Ecology: freshwater, moss and soil. Type species: *Paraquadrula irregularis* Archer, 1877 (fig. 58).



Fig. 57. *Protoplagiopyxis delamarei*: a – lateral view; b – apertural view (Bonnet, 1962)



Fig. 58. *Lamtoquadrula deflandrei*: a – lateral view; b – aperture view (Bonnet, 1975)

Family Phryganellidae Jung, 1942

Axially symmetrical, agglutinated shells; pseudopodia slightly conical pointed, sometimes branched, may sometimes anastomose. The status of this taxon is unclear, due to insufficient information is available.

Genus Phryganella Penard, 1902

Shell is circular in frontal view, hemispherical or higher in lateral view; covered with mineral particles of various size embedded in an organic matrix, which in older specimens can become dark frown due to manganese and iron deposition. Aperture is centrostome, circle, large often about two thirds of the shell diameter, not or only slightly invaginated. Around the aperture and the ventral face, covering particles are small are small giving a regular and smooth outline while larger grains are incorporated at the arboreal pole. Ecology: moss, soils. Type species: *Phryganella nidulus* Penard, 1902 (fig. 59).



Fig. 59. Genus Phryganella:

a, b – *P. acropodia* lateral (b) and aperture (a) view (after Chardez, 1961a); c – *P. microps* lateral view (after Valkanov, 1963); d – *P nidulus* lateral view (after Bartoš, 1954); e – *P. paradoxa* (after Bartoš, 1954)

Family Trigonopyxidae Loeblich et Tappan, 1964

Shell is radially symmetrical, circular in outline; composed of sheet-like proteinaceoius matrix with agglutinated particles; aperture central.

Key to the genera of the family

2. Aperture is invaginated	
2'. Aperture is not invaginated	Genus Geopyxella
3. Aperture is circular	Genus Cyclopyxis
3'. Aperture is crescent-shaped	Genus Cornuapyxis

Genus Cornuapyxis Coûteaux et Chardez, 1981

Shell is circular in frontal view, hemispherical in lateral view. Dorsal face is covered with small mineral grains, ventral surface is smooth. Aperture is crescent-shaped, slightly invaginated. Ecology: tropical mosses and soils. Type species: *Cornuapyxis lunaristoma* Coûteaux et Chardez, 1981 (fig. 60).



Fig. 60. *Cornuapyxis lunaristoma*: aperture view (after Coûteaux, Chardez, 1981)

Genus Cyclopyxis Deflandre, 1929

Shell is circular in frontal view, hemispherical in lateral view. Dorsal face is often rough, covered by large sand grains, ventral face is smooth. Aperture is invaginated, in most species circular, few species with irregular or lobed aperture; margin never with thick organic rim but often with small mineral particles. Ecology: freshwater, mosses, soils. Type species: *Cyclopyxis arcelloides* (Penard, 1902) (fig. 61, 62).

Genus Geopyxella Bonnet et Thomas, 1955

Shell is circular in frontal view, subglobular or hemispherical in lateral view. Dorsal face is covered with thin siliceous particles, ventral surface is smooth. Aperture is not invaginated, circular. Ecology: soil. Type species: *Geopyxella sylvicola* Bonnet et Thomas, 1955 (fig. 63).



Fig. 61. Genus Cyclopyxis:

a, b – C. amplecta lateral (a) and aperture (b) view (after Schönborn, 1966a); c, d – C. aplanata aperture (c) and lateral (d) view (after Penard, 1911); e, f – C. aplanata microstoma lateral (e) and aperture (f) view (after Schönborn, 1966a); g, h – C. arcelloides aperture (g) and lateral (h) view (after Penard, 1902); i – C. bacillifera aperture view (after Chardez, 1966); j, k – C. crucistoma aperture (j) and lateral (k) view (after Bartoš, 1963); l, m – C. eurystoma aperture (l) and lateral (m) view (after Deflandre, 1929); n, o – C. grospietschi aperture (n) and lateral (o) view (after Schönborn, 1962a)



Fig. 62. Genus Cyclopyxis:

a–d – *C. kahli* lateral (a, d), aperture (b) and dorsal (c) view (a, b – after Deflandre, 1929; c, d – after Foissner, Korganova, 1995); e, f – *C. penardi* aperture (e) and lateral (f) view (after Penard, 1911); g, h – *C. plana* aperture (g) and lateral (h) view (after Bartoš, 1963); i, j – *C. plana microstoma* lateral (j) and aperture (i) view (after Schönborn, 1966a); k–m – *C. stellata* lateral (k), aperture (l) view and aperture structure (m) (after Wailes, 1927); n, o – *C. tronconica* aperture (n) and lateral (o) view (after Godeanu, 1972)



Fig. 63. *Geopyxella aquatica*: a – lateral view; b – aperture view (Schönborn, 1965c)

Genus Trigonopyxis Penard, 1912

Shell is circular in frontal view, hemispherical or higher in lateral view; yellowish or brownish. Dorsal face is covered by xenosomes, ventral face is smooth. Aperture is invaginated, frequently triangular but more often irregular, always surrounded by a ring of organic cement, with a second membranous wall. Ecology: soil, litter, moss. Type species: *Trigonopyxis arcula* (Leidy, 1879) (fig. 64).



Fig. 64. Genus Trigonopyxis:

a, b – *T. arcula* aperture (a) and lateral (b) view (after Bobrov et al., 1995); c, d – *T. microstoma* aperture (c) and lateral (d) view (after Hoogenraad, de Groot, 1948); e, f – *T. minuta* aperture (e) and lateral (f) view (after Schönborn, Peschke, 1988)

INCERTAE SEDIS Arcellinida

Genus Argynnia Vucetich, 1974

Shell is ovoid, slightly compressed in cross section; covered with various euglyphid plates mixed with diatom fragments and mineral particles in porous cement, grayish. Aperture is terminal, circular, surrounded by siliceous plates giving a rough outline. Ecology: freshwater, moss, soil. Type species: *Argynnia schwabei* Jung, 1942 (figs 65, 66).





a – A. bipes lateral view (after Wailes, Penard, 1911); b – A. ertli lateral view (after Laminger, 1973); c – A. columbiana lateral view (after Wailes, 1925); d– f – A. caudata broad (d, f) and narrow (e) lateral view (after Leidy, 1879); g–j – A. dentistoma broad (g, j), narrow (h) and aperture (i) view (g – after Deflandre, 1936; h–j – after Penard, 1890); k – A. dentistoma laevis lateral view (after Deflandre, 1936); l – A. dentistoma hesperica lateral view (after Wailes, 1913); m – A. dentistoma lacustris lateral view (Wailes, 1912); n – A. retorta lateral view (after Chardez, 1958); o, p – A. spicata narrow (o) and broad (p) lateral view (after Jung, 1942)



Fig. 66. Genus Argynnia:

a – A. gertrudeana lateral view (after Jung, 1942); b, c – A. vitraea broad (c) and narrow (b) lateral view (after Penard, 1899); d, e – A. vitraea minor broad lateral view (d) and shell cover details (e) (after Deflandre, 1936)

Genus Awerintzewia Schouteden, 1906

Shell is ovoid, circular or slightly compressed in cross section; covered with irregular, embedded siliceous particles (no idiosomes of other testate amoebae). Aperture is oval, with internally thickened border. Ecology: freshwater, *Sphagnum*. Monospecific. Type species: *Awerintzewia cyclostoma* (Penard, 1902) (fig. 67).



Fig. 67. *Awertintzewia cyclostoma*: lateral view (after Geltzer et. al., 1995)

Genus Difflugia Leclerc, 1815

Shell is agglutinated, with a terminal aperture that is circular, oval, lobed or toothed (but never slit-like), sometimes with a collar or necklace (sensu Ogden, Meisterfeld, 1989; Mazei, Warren, 2012, 2014, 2015) but never with an internal diaphragm. The shell is composed of mineral particles or diatom frustules, collectively called xenosomes that are assembled on structured or sheet-like organic cement. All species of *Difflugia* acquire their xenosomes from their environment. Many select and arrange these xenosomes according to their size and shape in order to
construct a shell with a morphology that is unique to that particular species. The nucleus is usually ovular, but in some species it is vesicular. Several freshwater species have green endosymbionts (Meisterfeld, Mitchell, 2008). Type species: *Difflugia proteiformis* Lamarc, 1816 (fig. 68–70).



Fig. 68. Genus Difflugia:

a, b – D. acuminata lateral view (a – after Ogden, 1979; b – after Gauthier-Lièvre, Thomas, 1958); c – D. acuminata umbilicata lateral view (after Chardez, 1961); d – D. acutissima lateral view (after Gauthier-Lièvre, Thomas, 1958); e – D. acutissimella lateral view (after Chardez, 1985); f – D. bacillariarum lateral view (after Gauthier-Lièvre, Thomas, 1958); g – D. bacillifera lateral view (after Bartoš, 1954); h, i – D. compressa broad (h) and narrow (i) lateral view (after Schönborn, 1965a); j, k – D. compressa africana narrow (j) and broad (k) lateral view (after Gauthier-Lièvre, Thomas, 1958); n – D. elegans lateral view (after Gauthier-Lièvre, Thomas, 1958); n – D. elegans teres lateral view (after Gauthier-Lièvre, Thomas, 1958); p – D. elegans parva lateral view (after Gauthier-Lièvre, Thomas, 1958); p – D. elegans parva lateral view (after Jax, 1985); q – D. elegans lepida lateral view (after Schönborn, 1966a)



Fig. 69. Genus Difflugia:

a – D. gigantea lateral view (after Chardez, 1967c); b – D. gigantea acuminata lateral view (after Chardez, Gaspar, 1984); c, d – D. globulosa apertural (c) and lateral (d) view (after Gauthier-Lièvre, Thomas, 1958); e – D. globularis lateral view (after Beyens, Chardez, 1984); f – D. lebes lateral view (after Bartoš, 1954); g, h – D. lebes sphaerica aperture (g) and lateral view (h) (after Gauthier-Lièvre, Thomas, 1958); i – D. lebes masurica lateral view (after Schönborn, 1965a); j, k – D. leidyi lateral view (after Wailes, 1912); l, m – D. linearis lateral view (after Gauthier-Lièvre, Thomas, 1958); n, o – D. lingual narrow (n) and broad (o) lateral view (after Gauthier-Lièvre, Thomas, 1958); p, q – D. lingula regularis narrow (p) and broad (q) lateral view (after Gauthier-Lièvre, Thomas, 1958)



Fig. 70. Genus Difflugia:

a, b – *D. nodosa* narrow (a) and broad (b) lateral view (after Gautier-Lièvre, Thomas, 1958); c – *Difflugia oblonga* lateral view (after Chardez, 1967c); d, e – *D. oblonga angusticollis* lateral view (d – after Chardez, 1967c; e – after Štěpánek, 1952); f – *D. oblonga caudata* lateral view (after Chardez, 1967c); g – *D. oblonga cornuta* lateral view (after Chardez, 1967c); h – *D. oblonga incondita* lateral view (after Gauthier-Lièvre, Thomas, 1958); i – *D. oblonga schizocaulis* lateral view (after Chardez, 1967c); j – *D. oblonga stepaneki* lateral view (after Chardez, 1967c); k, 1 – *D. parva* lateral view (k – after Gauthier-Lièvre, Thomas, 1958; 1 – after Chardez, 1967c); m, n – *D. paulii* в плане (after Gauthier-Lièvre, Thomas, 1958;); o, p – *D. penardi* lateral view (after Bartoš, 1954)

Genus Geamphorella Bonnet, 1959

Shell is ovoid, circular in cross section, covered with amorphous silica elements, tinted clear yellow or grayish white. Aperture is terminal, with a chitionoid collar, slightly widened at the circular aperture. Ecology: calcareous soils. Monospecific. Type species: *Geamphorella lucida* Bonnet, 1959 (fig. 71).

Note: Décloître (1964) described *Pseudogeamphorella* with a compressed shell. However, the description is too vague.



Fig. 71. *Geamphorella lucida*: lateral view (after Bonnet, 1959)

Genus Ellipsopyxella Bonnet, 1975

Shell is elliptic in frontal view, hemispherical in lateral view. Dorsal surface is covered with relatively flat mineral particles (crystalline quartz) giving a smooth surface, ventral surface thicker than the rest of the shell. Aperture is central, circular, not or only weakly invaginated. Ecology: soils. Monospecific. Type species: *Ellipsopyxella regularis* Bonnet, 1975 (fig. 72).

Genus Ellipsopyxis Bonnet, 1965

Shell is elliptic in frontal view, semi-elliptic in lateral view on to the large axis; yellowish or clear maroon. Dorsal and lateral surfaces incrusted with small mineral particles, ventral surface is smooth due to organic cement. Aperture is central, elliptical, without or only slight Invagination. Ecology: mainly rich organic tropical soils. Type species: *Ellipsopyxis pauliani* Bonnet, 1965 (fig. 73).



Fig. 72. *Ellipsopyxella regularis*: a – lateral view; b – aperture view (after Bonnet, 1975a)



Fig. 73. *Ellipsopyxis pauliani*: a – aperture view; b – lateral view (after Bonnet, 1965)

Genus Heleopera Leidy, 1879

Shell is ovoid, laterally compressed, composed of collected euglyphid body plates, mineral particles or diatoms. These elements are often coated and reinforced with siliceous material. Besides colorless and yellow, several red or purple species are common. Aperture is terminal, lenticular or slit-like, with thin organic rim, with acute notches at edges. Some species have endosymbiotic zoochlorellae. Ecology: freshwater, moss, soil. Type species: *Heleopera sphagni* (Leidy, 1874) (fig. 74).



Fig. 74. Genus Heleopera:

a – *H. lata* broad lateral view (after Cash et al., 1909); b, c – *H. petricola* narrow (b) and broad (c) lateral view (after Geltzer et al., 1995); d – *H. rosea* broad lateral view; e, f – *H. sphagni* broad (e) and narrow (f) lateral view (after Geltzer et al., 1995); g, h – *H. sylvatica* broad (g) and narrow (h) lateral view (after Lüftenegger, Foissner, 1991)

Genus Jungia Loeblich et Tappan, 1961

Shell is globular to ovate, cross section is circular, covered with irregular polygonal or elongate plates. Aperture is terminal, circular, surrounded by sand grains forming a short collar. Ecology: mosses. Type species: *Jungia sudanensis* van Oye, 1949 (fig. 75).

Genus Lagenodifflugia Medioli et Scott, 1983

Shell pyriform, with a constriction of the neck, cross section is circular or slightly compressed, composed of mineral particles bounded by a structured organic cement network. In the region of the constriction, the shell is divided in two parts by a diaphragm-like part of the shell wall with a single central opening. Aperture is terminal, circular. Ecology: freshwater, *Sphagnum*. Type species: *Lagenodifflugia vas* (Leidy, 1874) Medioli et Scott, 1983 (fig. 76).



Fig. 75. *Jungia sudanensis*: a – narrow lateral view; b – broad lateral view (after Oye, 1951)



Fig. 76. *Lagenodifflugia montana*: lateral view (after Ogden, Živković, 1983). Scale bar 50 μm

Genus Leptochlamys West, 1901

Shell is ovoid or flask-shaped, cross section is circular; completely organic, non-alveolar, no exogenous material. Aperture is terminal, circular, with short collar. Ecology: freshwater. Two species. Type species: *Leptochlamys ampullacea* West, 1901. Also includes *Leptochlamys galippei* Schmidt et al., 2010 described from amber (fig. 77).



Fig. 77. *Leptochlamys ampullacea*: lateral view (after West, 1901)

Genus Lesquereusia Schlumberger, 1845

Shell with asymmetrical neck, more or less attached to the body; composed of endogenous siliceous rods or in some species with collected mineral particles in structured mesh-like cement. Fundus is rounded. Aperture is located at the end of the neck, circular in cross section. Ecology: freshwater. Type species: *Lesquereusia spiralis* (Ehrenberg, 1840) Bütschli, 1888 (fig. 78).



Fig. 78. Genus Lesquereusia:

a – L. combinata lateral view (after Bartoš, 1954); b–d – L. epistonium narrow (b), broad (c, d) lateral view (b, c – after Bartoš, 1954; d – after Ogden, 1984); e – L. inequalis lateral view (after Cash, Hopkinson, 1909); f – L. longicollis lateral view (after Dekhtiar, 1994); g – L. longicollis depressa lateral view (after Dekhtiar, 1994); h – L. modesta lateral view (after Bartoš, 1954); i, j – L. spiralis broad (i) and narrow (j) lateral view (after Bartoš, 1954)

Genus Maghrebia Gauthier-Lièvre et Thomas, 1960

Shell is cylindroid, slightly compressed in cross section, round base; constricted at short neck with four ridges like water jar with handles; covered with embedded siliceous particles. Aperture is terminal, circular. Habitat: tropical freshwater. Monospecific. Type species: *Maghrebia spatulata* Gautier-Lièvre et Thomas, 1958 (fig. 79).



Fig. 79. *Maghrebia spatulata*: a – aperture view; b – lateral view (after Gauthier-Lièvre, Thomas, 1958)

Genus Mediolus Patterson, 2014

Shell is ovoid to subspherical to spheroid; circular in cross-section, about the vertical aperture through the fundus; shell wall comprised of agglutinating particles variously composed of xenogenous mineral grains and/or organic material derived from the ambient environment; agglutinating particles attached together with an organic cement; circular aperture characterized by thin collar of secreted cement with variable number of inward-oriented angular crenulations also composed of cement; delicate spines may be present, spines long and narrow, hollow, and composed of very fine agglutinating particles. Ecology: freshwater. Type species: *Mediolus corona* (Wallich, 1864) (fig. 80).



Fig. 80. *Mediolus corona*: a – aperture view; b – lateral view (after Patterson, 2014). Scale bar 100 μm

Genus Microquadrula Golemansky, 1968

Shell is ovoid, circular in cross-section, covered with square or rectangular siliceous plates. Aperture is terminal, circular, truncated. Ecology: moss. Monospecific. Type species: *Microquadrula musciphila* Golemansky, 1968 (fig. 81).



Fig. 81. *Microquadrula musciphila*: lateral view (after Golemansky, 1968)

Genus Oopyxis Jung, 1942

Shell is ovoid, round or oval in cross section, ventral face is never flat; brown. Shell is covered with silica plates, irregular shape. Aperture is subterminal, slightly invaginated, irregular shape, relatively narrow, may be surrounded by large mineral particles. Ecology: freshwater. Type species: *Oopyxis cophostoma* Jung, 1924 (fig. 82).



Fig. 82. Genus Oopyxis:

a, b – O. cophostoma aperture (a) and lateral (b) view (after Jung, 1942); c, d – O. cyclostoma aperture (c) and lateral (d) view (after Schönborn, 1965a); e, f – O. danubialis aperture (e) and lateral (f) view (after Godeanu, 1972); g – O. islandica aperture view (after Décloître, 1966)

Genus Pentagonia Gauthier-Lièvre et Thomas, 1960

Shell is pyriform; in cross-section polygonal with three to five lateral swellings sometimes terminating in a horn; fundus with horn; covered with mineral particles. Aperture is terminal circular, with a short collar. Ecology: freshwater. Monospecific. Type species: *Pentagonia maroccana* Gautier-Lièvre et Thomas, 1958 (fig. 83).

Genus Physochilla Jung, 1942

Shell is pyriform, slightly compressed; covered with siliceous exogenous plates, grey in color. Aperture is terminal, more or less circular, with collar recurved posteriorly. Ecology: mosses. Type species: *Physochilla griseola* Wailes et Penard, 1911 (fig. 84).



Fig. 83. *Pentagonia maroccana*: a – lateral view; b – aperture view (after Gauthier-Lièvre, Thomas, 1958)



Fig. 84. Genus Physochilla:

a – *P. corniculata* lateral view (after Jung, 1942); b – *P. cratera* lateral view (after Wailes, 1912); c – *P. gauthier-lievri* lateral view (after Štěpanėk, 1963); d, e – *P. griseola* aperture (d) and lateral (e) view (after Penard, 1911); f, g – *P. tenella* narrow (f) and broad (g) lateral view (after Penard, 1893)

Genus Pomoriella Golemansky, 1970

Shell is flask-shaped, with a bent neck; covered with siliceous, nonoverlapping, intrinsic plates. Aperture is terminal, circular. Ecology: interstitial. Type species: *Pomoriella valkanovi* Golemansky, 1970 (fig. 85).



Fig. 85. Pomoriella valkanovi:

a – lateral view; b – aperture view; c – lateral view with pseudopodia (after Golemansky, 1970b)

Genus Pontigulasia Rhumbler, 1896

Shell is pyriform, circular or slightly compressed in cross-section, sometimes with a constriction between the main body and the neck. In the region of the constriction the shell is divided internally into two parts by a narrow mainly organic bridge with few attached mineral particles, which connects both broad sides. Shell is composed mainly of agglutinate mineral particles with some diatom frustules. Aperture is terminal, circular. Ecology: freshwater, mosses. Type species: *Pontigulasia rhumbleri* Hopkinson, 1919 (fig. 86).



Fig. 86. Genus Pontigulasia:

a, b – *P. breviottis* lateral view (after Snegovaya, Alekperov, 2005); c, d – *P. compressoidea* narrow (c) and broad (d) lateral view (after Chardez, 1958); e – *P. incisa* lateral view (after Bartoš, 1954); f, g – *P. rhumbleri* lateral (f) and aperture (g) view (after Cash et al., 1919); h – *P. sarrazinensis* lateral view (after Chardez, Gaspar, 1984); i – *P. spiralis* lateral view (after Bartoš, 1954)

Genus Protocucurbitella Gauthier-Lièvre et Thomas, 1960

Shell is agglutinated, ovoid, cross section is round. Aperture is circle, denticulate, surrounded by a diaphragm around peristome. Ecology: freshwater. Type species: *Protocucurbitella coroniformis* Gautier-Lièvre et Thomas, 1960.

Note: The taxonomic status of the genus is unclear and widely discussed (Dekhtiar, 1993; Snegovaya, Alekperov, 2010; Patterson, 2014).

Genus Pseudocucurbitella Gauthier-Lièvre et Thomas, 1960

Shell is agglutinated, ovoid, cross section is round. Aperture is circular, surrounded with 3–5 separate lobes forming a short collar. Ecology: freshwater. Type species: *Pseudocucurbitella subangelica* Gauthier-Lièvre & Thomas, 1960.

Note: The taxonomic status of the genus is unclear. Probably it has affinities to lobed *Difflugia* as well as to the genus *Cucurbitella*.

Genus Pseudawerintzewia Bonnet, 1959

Shell is oviform, circular in cross section; mainly composed of small siliceous plates, pasted together with abundant brown organic cement and arranged so that the shell surface appears smooth. Aperture is terminal in fattened center of the broader end, small, elliptical or circular, thickened lip, no collar. Ecology: soils. Type species: *Pseudoawerint-zewia calcicola* Bonnet, 1959 (fig. 87).

Genus Pseudonebela Gauthier-Lièvre, 1953

Shell is flask-like, round in cross section; composed of organic secretion of an apparently smooth cement and agglutination of extraneous particles including diatoms and, more commonly, many rounded plates, and with many small irregular platelets juxtaposed in between the larger rounded plates. Aperture is terminal, surrounded by an organic lip with 3–11 denticulations that give the opening a lobed appearance. Ecology: tropical freshwater. Monospecific. Type species: *Pseudonebela africana* Gautier-Lièvre, 1953 (fig. 88).



Fig. 87. *Pseudoawerintzewia calciola*: a – lateral view; b – aperture view (after Bonnet, 1959)



Fig. 88. *Pseudonebela africana*: lateral view (after Gauthier-Lièvre, 1953)

Genus Pyxidicula Ehrenberg, 1838

Shell is round in frontal view, hemispheric in lateral view; shell wall areolate, composed of organic, hollow building units, which in older, brown specimens become filled with inorganic material like manganese. Aperture is almost as wide as the diameter of the shell, apertural rim is usually recurved outside. One vesicular nucleus. Ecology: freshwater, soils. Type species: *Pyxidicula operculata* (Agardh, 1827) Ehrenberg, 1834 (fig. 89).



Fig. 89. Genus Pyxidicula:

a, b – *P. cymbalum* lateral (b) and dorsal view (a) (after Bartoš, 1954); c – *P. gibbosa* lateral view (after Schönborn, 1966a); d – *P. operculata* lateral view (after Bartoš, 1954); e, f – *P. ornata* (after Bartoš, 1954); g – *P. patens* lateral view (after Bartoš, 1954)

Genus Schoenbornia Décloître, 1964

Shell is ovoid, circular in cross section, hyaline, transparent; composed of collected idiosomes of small euglyphids, angular quartz and amorphous siliceous elements. Schönborn et al. (1987) suppose that the latter are produced by amoebae, but this remains unproved. Aperture is terminal, circular. Ecology: soils. Type species: *Schoenbornia humicola* Schönborn, 1964 (fig. 90).

Note: *Heleoporella* Coûteaux, 1978 is probably a synonym of this genus.

Genus Schwabia Jung, 1942

Shell is ovoid, circular in cross section, chitinous with small small mineral particles which produce a smooth surface; opaque, grayish, dirtyellow or brown in color. Aperture is terminal, circular. Ecology: freshwater, soils. Type species: *Schwabia regularis* Jung, 1942 (fig. 91).





Fig. 90. *Schoenbornia vescicula*: a – lateral view; b – aperture view (after Schönborn, 1964) Fig. 91. *Schwabia regularis*: a – aperture-lateral view; b – lateral view (after Jung, 1942a)

Genus Sexangularia Awerintzew, 1906

Shell is angular in frontal view; polygonal, mostly hexagonal, in cross section, completely organic. Aperture is terminal, circular. Nucleus is vesicular. Ecology: freshwater, *Sphagnum*. Type species: *Sexangularia minutissima* (Penard, 1904) Deflandre, 1931 (fig. 92).



Fig. 92. *Sexangularia minutissima*: a – aperture view; b – lateral view (after Deflandre, 1953)

Genus Suiadifflugia Green, 1975

Shell is sub-spherical, cross section is circular, covered by xenosomes. Aperture is terminal, complex, consisting of a central pore and numerous elongate oval pores radiating around the central pore like the petals of a flower. Ecology: freshwater plants and sediments. Monospecific. Type species: *Suiadifflugia multipora* Green, 1975.

Genus Zivkovicia Ogden, 1987

Shell is pyriform, mostly with a distinct constriction of the neck which sometimes can be obscured by larger mineral particles, circular or compressed in cross section, built of agglutinated mineral particles bound by a structured organic cement. The neck is separated from the main body of the shell by an internal mineral diagram with two circular openings. Aperture is terminal, circular. Ecology: freshwater. Type species: *Zivkovicia compressa* (Carter, 1864) (fig. 93).



Fig. 93. Genus Zivkovicia:

a, b – Z. compressa broad (a) and narrow lateral view (b) (after Cash, Hopkinson, 1909); c, d – Z. flexa narrow (c) and broad (d) lateral view (after Cash, Hopkinson, 1909); e – Z. spectabilis lateral view (after Bartoš, 1954)

Chapter 3 Order Euglyphida Copeland, 1956, emend. Cavalier-Smith, 1997

Shell is made of organic material; most taxa with secreted silica scales held together by an organic cement; tubular mitochondrial cristae.

Key to the families of the order and genera incertae sedis

1. Shell is radially symmetrical; cross section is circular
or elliptic2
1'. Shell is bilaterally symmetrical12
2. Shell is organic, without scales; if scales present,
they are long, with length perpendicular to aperture
2'. Shell is covered by scales
3. Shell with a with a straight cylindrical neck
3'. Shell without neck, if present very short4
4. Fundus with scaly spine of variable length Genus Pareuglypha
4'. Fundus is without scaly spine5
5. Scales are overlapping, regularly distributed
5'. Scales are non-overleaping, randomly distributed11
6. Aperture is surrounded by specialized scales7
6'. No specialized type of scales around aperture9
7. Scales around aperture are denticulate
7'. Scales around aperture are small, circular or oval,
without teeth Family Sphenoderiidae
8. Shell surface is tuberculate due to large elongate idiosomes
surrounded by smaller idiosomes
8'. Shell surface is flat Family Euglyphidae
9. Aperture is circular or elongate10
9'. Aperture is slit-like or slightly arched, located at right angle
to the broad diameter, with clear organic rimGenus Heteroglypha
10. Shell is compressed in cross section, covered by small oval
overlapping scales Family Assulinidae
10'. Shell cross-section is circular, covered by large, circular
overlapping idiosomes Genus Tracheleuglypha

Suborder Euglyphina Kosakyan et al., 2016

Comprises the last common ancestor of families Assulinidae, Euglyphidae, Sphenoderiidae and Trinematidae and all its descendants.

Family Assulinidae Lara et al., 2007

Acrostome shell composed of elliptic plates disposed in a regular, alternate pattern; shell strongly compressed; no specialized type of scales around aperture. Includes three genera: *Assulina, Placocista, Valkanovia*.

Key to the genera of the family

1. Aperture is slit-like, biconvex, shell often with	
spines	. Genus <i>Placocista</i>
1'. Aperture is oval or circular, never with spines	
2. Aperture is crenulate	Genus Assulina
2'. Aperture is smooth	Genus <i>Valkanovia</i>

Genus Assulina Leidy, 1879

Shell is ovoid or broad oval, flattened, biconvex in cross section, covered by overlapping elliptical idiosomes, clear to brown. Aperture is narrow oval, more or less crenulate, formed by organic margin. Ecology: mosses, soils. Type species: *Assulina seminulum* (Ehrenberg, 1848) Leidy, 1879 (fig. 94,a–k).

Genus Placocista Leidy, 1879

Shell is ovoid, flattened, with an acute lateral border; edge may bear many short, often paired, lanceolate spines, cemented between scales. Idiosomes are broad- or long-elliptic, sometimes circular, margins overlap. Aperture is large, narrowly biconvex, with pointed edges, in lateral view incised and surrounded by a thin rim of organic cement. Cytoplasm sometimes contains zoochlorellae. Habitat: mosses. Type species: *Placocista spinosa* (Carter, 1865) (fig. 94,1–y).



Fig. 94. Genera Assulina and Placocista:

a–c – A. collaris broad (a), narrow (b) lateral and aperture (c) view (after Schönborn, Peschke, 1988); d–g – A. muscorum broad (d, g), narrow (e) lateral and aperture (f) view (d–f – after Schönborn, Peschke, 1988; g – after Lüftenegger, Foissner, 1991); h, i – A. quadratum broad lateral view (after Oye, 1958); j, k – A. seminulum broad (j) and narrow (k) lateral view (after Lüftenegger et al., 1988); l, m – P. glabra narrow (l) and broad (m) lateral view (after Jung, 1936); n – P. glabra minima broad lateral view (after Décloître, 1955); o – P. jurassica broad lateral view (after Penard, 1905); p, q – P. lapponum narrow (p) and broad (q) lateral view (after Penard, 1917); r, s – P. lens broad lateral (r) and aperture (s) view (after Rauenbush, 1987); t, u – P. spinosa in narrow (v) and broad (w) lateral view (after Chardez, 1966); v, w – P. spinosa in narrow (v) and broad (w) lateral view (after Chardez, 1966); x, y – P. ventricosa broad (x) and narrow (y) lateral view (after Chardez, 1966)

Genus Valkanovia Tappan, 1966

Valkanovia is closely related to *Assulina* and differs from it by the smooth aperture edge. The shell is generally colorless. The status of the genus is uncertain as there are transitional forms between the genera. Ecology: soils. Type species: *Valkanovia delicatula* (Valkanov, 1962) Tappan, 1966 (fig. 95).



Fig. 95. *Valkanovia delicatula*: broad lateral view (after Valkanov, 1962)

Family Euglyphidae Wallich, 1864, emend. Lara et al., 2007

Acrostome shell composed of elliptic, sub-rectangular, scutiform, or almost circular body plates which are disposed in a regular, alternate pattern. Aperture is surrounded by denticulate plates (Lara et al., 2007). Includes two genera: *Euglypha, Scutiglypha*.

Key to the genera of the family

Genus Euglypha Dujardin, 1841

Shell is radially symmetric, ovoid or long-ovoid, circular or elliptic in cross-section; covered by circular or ovoid endogenous siliceous plates which overlap and usually arranged in longitudinal rows. Sometimes, body scales carry needles or spines. Aperture is terminal (acrostome), circular or ovoid, always surrounded by denticulate plates of various form (oval, circular, rhombic, triangular). Shell is colorless, transparent, rare brownish or yellowish. Ecology: freshwaters, mosses, soils. Type species: *Euglypha tuberculata* Dujardin, 1841(fig. 96–98).



Fig. 96. Genus *Euglypha*:

a-c – *E. acanthophora* lateral view (a), aperture part (b), idiosomes with spine (c) (after Wailes, 1912); d – *E. acanthophora brevispina* lateral view (after Penard, 1902); e – *E. acanthophora cirrata* fundus base with spines (after Wailes, 1912); f – *E. acanthophora cylindracea* lateral view (after Playfair, 1917); g – *E. acanthophora deflandrei* lateral view (after Deflandre, 1956); h – *E. acanthophora equeis* lateral view (after Décloître, 1956); i – *E. acanthophora flexuosa* lateral view (after Penard, 1902); j – *E. acanthophora flexuosa* lateral view (after Décloître, 1949); k – *E. andonta* lateral view (after Geltzer et al., 1995); l, m – *E. brachiata* lateral view (l) and aperture idiosomes (m) (after Leidy, 1879); n – *E. brachiata librata* lateral view (after Wailes, 1912); o – *E. bryophila* lateral view (after Wailes, 1912); p–r – *E. capsiosa* lateral view (p), aperture idiosomes broad (r) and narrow (q) lateral view (after Coûteaux et al., 1979); s – *E. ciliata* lateral view (after Wailes, 1912); t – *E. ciliata glabra* lateral view (after Wailes, 1912); u – *E. ciliata heterospina* lateral view (after Wailes, 1912); u – *E. ciliata heterospina* lateral view (after Wailes, 1912); u – *E. ciliata heterospina* lateral view (after Wailes, 1912); u – *E. ciliata* heterospina



Fig. 97. Genus Euglypha:

a – E. compressa broad lateral view (after Wailes, 1912); b – E. crenulata elongata (after Thomas, 1958b); c–e – E. cristata lateral view (e), aperture idiosomes broad (d) and narrow (c) lateral view (after Coûteaux et al., 1979); f, g – E. cristata acicularis fundus base (after Wailes, 1912); h – E. cristata decora lateral view (after Jung, 1942); i – E. cristata lanceolata lateral view (after Playfair, 1917); j – E. cristata major lateral view (after Wailes, 1912); k – E. denticulata (after Geltzer et al., 1995); l–n – E. filifera lateral view (n) and aperture idiosomes broad (m) and narrow (l) lateral view (after Coûteaux et al., 1979); o – E. filifera cylindracea lateral view (after Playfair, 1917); p – E. filifera magna lateral view (after Oye, 1958); q – E. filifera pyriformis lateral view (after Wailes, 1913); r – E. filifera spinosa lateral view (after Wailes, 1912); s, t – E. gauthieri lateral view (s) and aperture idiosomes (t) (after Thomas, 1958); u – E. hutchinsoni (after Oye, 1932); v–x – E. hyalina lateral view (x), aperture idiosomes broad (w) and narrow (v) lateral view (after Coûteaux et al., 1979)



Fig. 98. Genus Euglypha:

a – E. laevis lateral view (after Geltzer et al., 1995); b – E. marginata lateral view (after Oye, 1958); c – E. mucronata lateral view (after Leidy, 1879); d – E. rotunda lateral view (after Cash et al., 1915); e – E. rotunda oblique lateral view (after Geltzer et al., 1995); f–h – E. simplex lateral view (h), aperture idiosomes broad (g) and narrow (f) lateral view (after Coûteaux et al., 1979); i–m – E. strigosa broad (k, m), narrow (l) lateral view, aperture idiosomes broad (j) and narrow (i) lateral view (i–j – after Coûteaux et al., 1979; l, m – after Geltzer et al., 1995); n – E. strigosa glabra lateral view (after Wailes, 1912); o – E. strigosa heterospina lateral view (after Wailes, 1912); p – E. strigosa muscorum lateral view (after Wailes, 1912); q – E. tuberculata lateral view (after Geltzer et al., 1995); r – E. van oyei lateral view (after Oye, 1958)

Genus Scutiglypha Foissner et Schiller, 2001

Body scales are scutiform or crenate. Ecology: freshwaters, mosses, soils. Type species: *Scutiglypha crenulata* Wailes, 1912 (basionym: *Euglypha crenulata* Wailes, 1912) (fig. 99).



Fig. 99. Genus Scutiglypha:

a–c – *S. aspera* lateral view (c), aperture idiosome (a) and parietal idiosome (b) (after Penard, 1902); d – *S. cabrolae* lateral view (after Smet, Gibson, 2009); e–g – *S. crenulata* lateral view (f), parietal idiosome with spine (g) and without spine (e) (after Wailes, 1912); h – *S. scutigera* lateral view (after Penard, 1912)

Family Sphenoderiidae Chatelain, 2013

Shell is covered with self-secreted circular to elliptical silica scales that can be of different sizes and shapes, but without indentations. Aperture is surrounded with small circular or oval scales. Because usually one side ("ventral") of the aperture is shorter the opening lies subterminal. Includes two genera: *Sphenoderia, Trachelocorythion*.

Key to the genera of the family

Genus Sphenoderia Schlumberger, 1845

Scales of one or more types on the main body the shell, aperture slit-like, mostly surrounded by a collar that comprises small scales that can be sometimes invaginated (*S. sphaerica*). Circular or oval cross section (e.g. *S. compressa, S. labiata*). Ecology: freshwater, moss, soils. Type species: *Sphenoderia lenta* Schlumberger, 1845 (fig. 100).



Fig. 100. Genus Sphenoderia:

a, b – *S. fissirostris* narrow (a) and broad (b) lateral view (after Geltzer et al., 1995); c – *S. lenta* broad lateral view (after Bartoš, 1954); d – *S. macrolepsis* broad lateral view (after Bartoš, 1954)

Genus Trachelocorythion Bonnet, 1979

Main body of the shell is covered by scales of regular size and shape; flattened cross section. The upper lip of the aperture larger than the lower resulting in a slightly subterminal opening, no collar. Ecology: dry soils to forest litter and *Sphagnum* mosses. Monospecific. Type species: *Trachelocorythion pulchellum* (Penard, 1890) Bonnet, 1979 (fig. 101).



Fig. 101. *Trachelocorythion pulchellum*: ventral view (after Meisterfeld, 2002b)

Family Trinematidae (Hoogenraad et de Groot, 1940) Adl et al., 2012

Shell with bilateral symmetry; scales oval or circular, sometimes of both types; specialized tooth-shaped scales around the aperture; aperture invaginated in some taxa. Includes four genera: *Corythion, Playfairina, Puytoracia, Trinema*.

Key to the genera of the family

1. Aperture is invaginated (lateral view)	
1'. Aperture is not invaginated	Genus <i>Playfairina</i>
2. Body plates are usually of two sizes,	the smaller plates filling the
gaps between the larger ones	2

usually of the same size	2'. Body plates are elongate ova
Genus Corythion	
Genus Trinema	3. Body scales are circular
Genus Puytoracia	3'. Body scales are elliptical

Genus Corythion Tarànek, 1881

In ventral view, the shell is ovoid; in lateral view the shell is compressed; cross-section lenticular lateral margins often slightly acute. Shell is covered by on type of body plates: circular, rectangular, imbricated, irregularly distributed but at lateral margins in rows. Aperture is invaginated, oval, semicircular or circular, surrounded by plates with one central tooth. Some species have short organic or siliceous spines. Type species: *Corythion dubium* Tarànek, 1881(fig. 102,a–i).

Genus Playfairina Thomas, 1961

Shell is circular in cross section, tapering towards the aperture; fundus rounded or pointed. Shell is covered large and small idiosomes, arrangement in most cases as in *Trinema*. Aperture is circular, not invaginated, surrounded by one row of denticulated marginal plates. Pseudopoida are unknown, but doe to the homologies in shell morphology *Playfairina* is placed here. Two species. Ecology: mosses, soils. Type species: *Playfairina caudata* (Playfair, 1917) Thomas, 1961 (fig. 102,j).



Fig. 102. Genera Corythion and Playfairina:

a–c – *C. asperulum* aperture (a, b) and lateral view (c) (a – after Lüftenegger, Foissner, 1991; b, c – after Schönborn, Peschke, 1998); d–g – *C. dubium* aperture (d, g) and lateral view (e, f) (d, e – after Schönborn, Peschke, 1998; f, g – after Lüftenegger et al., 1988); h, i – *C. orbicularis* aperture (h) and lateral view (i) (after Iudina, 1996); j – *P. cauduta* aperture view (after Thomas, 1961)

Genus Trinema Dujardin, 1841

In ventral view, shell is ovoid or broad oval, sometimes with almost parallel sides. In lateral view, the dorsal side is arched or flattened towards the aperture, ventral side is either flat or concave. Shell is covered by body-scales of two types: large circular plates and smaller circular or broad elliptical scales in between, overlapping. Aperture is sub-terminal or ventral, circular or oval, invaginated, surrounded by rows of toothed apertural plates. Ecology: freshwaters, mosses, soils. Type species: *Trinema enchelys* (Ehrenberg, 1838) Leidy, 1878 (fig. 103, 104,a–i).

Genus Puytoracia Bonnet, 1970

The genus is closely related to shell shape and size to genus *Trinema* and is distinguished by the presence of elliptical scales, covering the shell. Aperture is invaginated (Santibáñeza et al., 2011). Ecology: mosses, soils. Type species: *Puytoracia bergeri* Bonnet, 1970 (fig. 104,j–l).

Family Cyphoderiidae de Saedeleer, 1934

Scales are circular, oval or kidney-shaped, juxtaposed or imbricated; apertural end of the shell is angled, sometimes aperture is surrounded by collar. Includes six genera: *Campascus, Corythionella, Cyphoderia, Messemvriella, Pseudocorythion, Schaudinnula*.

Key to the genera of the family

1. Aperture is surrounded by a collar or a f	unnel3
1'. Aperture is without a collar or a funnel	
2. Cross-section of the shell is circular	Genus Cyphoderia
2'. Cross-section is triangular	Genus <i>Schaudinnula</i>
3. Apertural collar is made of organic matr	ix of the shell4
3'. Apertural collar is hyaline, fragile and rapidly	
disappears in empty shell	Genus Campascus
4. Fundus is rounded	5
4'. Fundus is pointed	.Genus Pseudocorythion
5. Shell is dorso-ventrally compressed	Genus Corythionella
5'. Shell is circular in cross section	Genus Messemvriella



Fig. 103. Genus Trinema:

a, b – *T. chardezi* aperture (a) and lateral view (b) (after Décloître, 1981); c – *T. ciliata* aperture view (after Štěpànek, 1963); d–g – *T. complanatum* aperture (d, f) and lateral view (e, g) (d, e – after Lüftenegger et al., 1988; f, g – after Geltzer et al., 1995); h, i – *T. enchelys* aperture (h) and lateral view (i) (after Lüftenegger et al., 1988); j, k – *T. enchelys biconvex* aperture (j) and lateral view (k) (after Awerintzew, 1907); l – *T. galeata* (after Penard, 1890); m, n – *T. grandis* aperture (m) and lateral view (n) (after Chardez, 1960); o – *Trinema intermedia* lateral view (Décloître, 1965); p, q – *T. leidyi* aperture (p) and lateral view (q) (after Décloître, 1981); r, s – *T. lineare* aperture (r) and lateral view (s) (after Lüftenegger et al., 1988); t, u – *T. lineare truncatum* aperture (t) and lateral view (u) (after Chardez, 1964)



Fig. 104. Genera Trinema and Puytoracia:

a, b – *T. lincostoma* aperture (a) and lateral view (b) (after Décloître, 1962c); c, d – *T. navicularis* aperture (c) and lateral view (d) (after Décloître, 1973); e–h – *T. penardi* aperture (e, g) and lateral view (f, h) (d, e – after Lüftenegger et al., 1988; g, h – after Thomas, Chardez, 1958); i, j – *P. bergeri* aperture (i) and lateral view (j); k, 1 – *P. bonneti* aperture (k) and lateral view (l) (after Nicholls, 2006b)

Genus Campascus Leidy, 1879

Shell is broadly ovoid; in cross-section circular, ovoid or triangular. Neck is cylindrical, curved. Fundus is rounded or with up to three projections. Aperture is circular, surrounded by a characteristic hyaline collar. Composition of the shell is variable; some species have circular, other amorphous, siliceous scales sometimes mixed with mineral particles, usually not regularly arranged. Ecology: freshwater and marine. Type species: *Campascus cornutus* Leidy, 1879 (fig. 105).

Genus Corythionella Golemansky, 1970

Shell is ovoid, dorso-ventrally compressed; fundus is rounded. Aperture is circular, located on the ventral side, surrounded by large circular collar. Shell is covered with elongate, siliceous idiosomes, irregularly arranged as in *Corythion*, the plates of the collar are smaller (Nicholls, 2009b). Ecology: marine and freshwater interstitial. Type species: *Corythionella pontica* Golemansky, 1970 (fig. 106).



Fig. 105. Campascus cornutus: ventral view (after Leidy, 1879)



Fig. 106. *Corythionella pontica*: a – lateral view; b – apertural view; c – shell plates (after Golemansky, 1970b)

Genus Cyphoderia Schlumberger, 1845

Shell is retort-shaped laterally; ovoid in ventral and dorsal views, usually circular in cross-section, sometimes slightly compressed. Fundus is round or tapered to mammillate tip. Aperture is circular without a collar. Shell is covered by small circular or triangular plates, either adjoining or overlapping on organic matrix depending on species. Endoplasm contains refractive grains. Ecology: marine interstitial, freshwater and moss. Type species: *Cyphoderia ampulla* (Ehrenberg, 1840) Leidy, 1879 (fig. 107).

Genus Messemvriella Golemansky, 1973

Shell is retort-shaped laterally, ovoid in ventral and dorsal view, circular in cross section. Fundus is round. Shell is covered with regularly overlapping siliceous scales. Aperture is subterminal, circular, surrounded by a collar, which is less developed than in *Corythionella*. Ecology: marine interstitial. Type species: *Messemvriella filose* Golemansky, 1972 (fig. 108).

Genus Pseudocorythion Valkanov, 1970

Shell is ovoid in ventral view, tapering at both ends, slightly flattened dorso-ventrally. Fundus carries a small pointed spine of variable length. Shell is covered with small or oval overlapping plates which are randomly arranged. Aperture is located on the ventral side; surrounded by a large circular collar of the same size as the maximum breadth of the shell. Outer edge of the collar is with very thin organic rim. Ecology: marine interstitial. Type species: *Pseudocorythion acutum* (Wailes, 1927) (fig. 109).





a – C. ampulla lateral view (after Chardez, 1991); b – C. ampulla bicornis lateral view (after Chardez, 1991); c – C. ampulla crassa lateral view (after Chardez, 1991); d – C. ampulla papillata lateral view (after Chardez, 1991); e – C. ampulla thomasi lateral view (after Chardez, 1991); f – C. ampulla virtae lateral view (after Chardez, 1991); g – C. bonetti lateral view (after Štěpànek, 1967); h – C. calceolus lateral view (after Chardez, 1991); i – C. compressa lateral view (after Chardez, 1991); j – C. loevis lateral view (after Chardez, 1991); k – C. lunata lateral view (after Štěpànek, 1967); l – C. perlucidus lateral view (after Chardez, 1991); m – C. schonborni lateral view (after Laminger, 1973); n, o – C. trochus lateral view (after Chardez, 1991); p – C. trochus amphoralis lateral view (after Chardez, 1991); q – C. ventricosa lateral view (after Chardez, 1991); r – C. venustus lateral view (after Chardez, 1991)





Fig. 108. *Messemvriella filose*: a – lateral view; b – apertural view (after Golemansky, 1973)

Fig. 109. *Pseudocorythion acutum*: a – aperture view; b – lateral view (after Wailes, 1927)

Genus Schaudinnula Awerintzew, 1907

Shell is retort-shaped ventrally, neck is bent more or less, fundus is pointed; cross-section is triangular. Shell is covered with circular overlapping idiosomes forming hexagonal structure. Aperture is circular, without a hyaline collar. Ecology: freshwater. Type species: *Schaudinnula arcelloides* Awerintzew, 1907 (fig. 110).



Fig. 110. *Schaudinnula arcelloides*: a – lateral view; b, c – cover (after Schönborn, 1965a)

Family Paulinellidae de Saedeleer, 1934, emend. Adl et al., 2012

Pyriform, uncompressed shape; scales, when present, long, with length perpendicular to aperture. Includes three genera: *Micropyxidiella*, *Ovulinata*, *Paulinella*.

Key to the genera of the family

ous rectangular plates, scarcely	1. Shell is covered with sili
ential longitudinal rows	overlapping, arranged in circumf
Genus Paulinella	
thout self-secreted scales or mineral	1'.Shell is entirely organic
	particles
Genus Micropyxidiella	2. Fundus is pointed
	2'.Fundus is rounded

Genus Micropyxidiella Tarnawski et Lara, 2015

Testate amoebae with filamentous pseudopodia, entirely organic shell (without self-secreted scales or mineral particles). Shell is ovoid, transparent and comparable to the related species of *Ovulinata parva*. Shell with a pointed end, reminding of certain members of genus *Difflugia* such as *D. acuminata*, clearly visible under scanning electron microscopy, but not under light microscopy. Aperture is terminal, slitlike. Large round nucleus (about 20 % of shell length). Ecology: soil. Monospecific. Type species: *Micropyxidiella edaphonis* Tarnawski et Lara, 2015.

Genus Ovulinata Anderson et al., 1997

Shell is ovoid, small (15 μ m), organic, lacking scales or mineral particles, aperture is terminal, circular to oval, pseudopodia filose, hyaline (sometimes branched), long tapering to a point, arising directly from aperture or from the periphery of a web of hyaline cytoplasm emergent from the aperture. Mitochondria with tubular cristae. Ecology: marine interstitial. Type species: *Ovulinata parva* Anderson et al., 1997.

Note: After original description (Anderson et al., 1996, 1997) Howe et al. (2011) placed the genus to a new family Ovulinatidae. Diagnosis: filose amoebae with ovoid organic shell, lacking scales or mineral particles, aperture is circular to oval; pseudopodia hyaline, sometimes branched, long, tapering to a point, arising directly from aperture or fromweb of hyaline cytoplasm emergent from it. Differs from its sister family Paulinellidae by lacking silica scales or plastid-like enslaved cyanobacterium. Type genus: *Ovulinata* Anderson, Rogerson et Hannah, 1997. This new family is essential because the trees show that *Ovulinata* was previously wrongly classified in Pseudodifflugiidae (Thecofilosea) and must be transferred to Imbricatea and Euglyphida. It differs so radically from *Paulinella* that it cannot be included in the same family.

Genus Paulinella Lauterborn, 1895

Shell is ovoid, covered with siliceous rectangular plates with slightly rounded, scarcely overlapping ends, arranged in circumferential longitudinal rows; in apertural view the plates form a counter-clockwise spiral, with one pentagonal scale at the aboral pole. Aperture is terminal or suberminal, circular or oval, sometimes located on the end of a small neck. Cytoplasm usually contains two sausage-like endosymbionts of cyanobacterial origin. Ecology: marine, brackish water and freshwater plants. Type species: *Paulinella chromatophora* Lauterborn, 1895 (fig. 111).



Fig. 111. *Paulinella chromatophora*: lateral view (after Saedeleer, 1934)

INCERTAE SEDIS Euglyphida

Genus Ampullataria van Oye, 1956

Shell is ellipsoid, circular in cross-section, with a cylindrical neck, covered by oval overlapping scales which appear polygonal. Neck scales are irregular at aperture. Aperture is circular, terminal. Ecology: moss. Type species: *Ampullataria rotunda* van Oye, 1956 (fig. 112).



Fig. 112. Ampullataria rotunda: lateral view (after Oye, 1956)

Genus *Deharvengia* Bonnet, 1979, emend. Bobrov et al., 2012

Shell is transparent, oval or ovoid in the broad view, circular to narrow elliptical in the cross-section. Aperture elliptic or narrow elliptic, the ventral side is cut at about 6/7 of the length of the entire shell and is surrounded by a thin organic lip. On the dorsal side the shell forms a small cap over the aperture, sometimes with a narrow organic lip along the edge. Shell is covered by elongate-elliptic idiosomes. Transparent clavate spines may be present on each side of the shell (Bobrov et al., 2012). Ecology: freshwater, soils. Type species: *Deharvengia papuensis* Bonnet, 1979 (fig. 113).



Fig. 113. *Deharvengia papuensis*: a – apertural view; b, c – aperture (after Bonnet, 1979)

Genus Euglyphidion Bonnet, 1960

Shell is ovoid, circular in cross-section, sides taper. Aperture is terminal, circular or broadly ovoid. Shell is covered by siliceous two types of plates which do not overlap, distributed chaotically. Ecology: soils. Monospecific. Type species: *Euglyphidion enigmaticum* Bonnet, 1960 (fig. 114).



Fig. 114. *Euglyphidion enigmaticum*: lateral view (after Bonnet, 1960b)

Genus Heteroglypha Thomas et Gauthier-Lièvre, 1959

Shell is wedge-shaped, fundus is rounded, sides sub-parallel, anterior end only slighlty smaller than fundus. In lateral view, shell is tapered almost to point at apertural end. Aperture is terminal, slit-like or slightly arched, located at right angle to the broad diameter, with clear organic rim. Scales are elliptical, arranged as in *Euglypha*. Ecology: freshwater. Type species: *Heteroglypha delicatula* Thomas et Gauthier-Lièvre, 1959 (fig. 115).



Fig. 115. *Heteroglypha delicatula*: a – broad lateral view; b – narrow lateral view; c – apertural view (after Thomas, Gauthier-Lièvre, 1959a)

Genus Matsakision Bonnet, 1967

Shell is transparent, long ovoid, slightly compressed, perpendicularly truncated near the aperture area. Shell is covered by elliptic plates, not overlapping, although sometimes may be connected, located on light yellow hyaline cement. Aperture is terminal, oval, surrounded by small organic collar, hardly visible. Pseudopodia are not reported. Ecology: terrestrial mosses. Type species: *Matsakision cassagnaui* Bonnet, 1967.

Genus Pareuglypha Penard, 1902

Shell is ovoid, tapered toward aperture, yellowish; fundus with scaly spine of variable length. Shell is covered by small, circular or ovoid scales, slightly overlapping, randomly distributed. Aperture is terminal, circular. Ecology: freshwater. Type species: *Pareuglypha reticulata* Penard, 1902 (fig. 116).

Genus Pileolus Coûteaux et Chardez, 1981

Shell is acrostome, covered by two types of idiosomes. Large elongate idiosomes are thickened in the middle that gives tuberculated view to the shell. Small elongate idiosomes fill the gaps between the larger ones. Aperture is circular, with a collar turned up and outside, surrounded by small elongated plates with a single tooth. Monospecific. Ecology: soil. Type species: *Pileolus tuberosus* Coûteaux et Chardez, 1981(fig. 117).



Fig. 116. *Pareuglypha reticulata*: a – lateral view; b – cover; c – lateral view with filopodia (after Penard, 1902)

Genus Tracheleuglypha Deflandre, 1928

Shell is ovoid, cross-section is circular, covered by large, circular overlapping idiosomes; colourless, transparent. Aperture is terminal, circular, sometimes may be surrounded by a small denticulate collar made of the organic material of the shell. Nucleus is ovular. Ecology: freshwater, moss and soil. Type species: *Tracheleuglypha dentata* Deflandre, 1938 (fig. 118).



Fig. 117. *Pileolus tuberosus:* apertural view (after Coûteaux, Chardez, 1981)



Fig. 118. *Tracheeuglypha dentata*: a – lateral view; b – apertural view (after Lüftenegger, Foissner, 1991)
Chapter 4 Order Amphitremida Poche, 1913, emend. Gomaa, Mitchell et Lara, 2013

Pseudopods are filamentous, sometimes anastomosing and branching. Shell is rigid, with two apertures located in front of each other on the main axis of symmetry of the shell.

Family Amphitremidae Poche, 1913

Shell is elliptic or barrel like in the broad lateral view and compressed in the narrow lateral view; either organic or with attached exogenous material. Two apertures are located in front of each other on the main axis of symmetry of the shell. Cytoplasm contains symbiotic zoochlorellae.

Key to the genera of the family

Genus Amphitrema Archer, 1869

Shell is elliptic in the broad lateral view, compressed in the narrow lateral view, covered by exogenous particles (diatom frustules, flagellate cysts etc.). A patterned cement network can be seen sometimes by scanning electron microscopy. Two elliptic apertures, with or without short collar, are located on the opposite sides of the shell. Type species: *Amphitrema wringhtianum* Archer, 1869 (fig. 119).

Genus Archerella Loeblich et Tappan, 1961

Shell is elliptic in the broad lateral view and compressed in the narrow lateral view. Covering elements are absent. Two elliptic apertures are located at the opposite sides of the shell and surrounded by short, very thin collars. Shell is often brown due to iron accumulation. Type species: *Archerella flavum* Archer, 1877 (fig. 120).



Fig. 119. Genus Amphitrema:

a – A. congolense lateral view (after Oye, 1958); b – A. lemanense lateral view with pseudopodia (after Bartoš, 1954); c – A. paparoensis lateral view (after Oye, 1958); d – A. stenostoma lateral view (after Bartoš, 1954); e – A. Wringhtianum lateral view (after Bartoš, 1954)





a – *A. flavum* broad lateral view (after Schönborn, 1966a); b – *A. jollyi* broad lateral view (after Oye, 1958)

References

1. Adl S.M., Simpson A.G.B., Lane C.E., Lukeš J., Bass D., Bowser S.S., Brown M.W., Burki F., Dunthorn M., Hampl V., Heiss A., Hoppenrath M., Lara E., Gall L. Le, Lynn D.H., McManus H., Mitchell E.A.D., Mozley -Stanridge S.E., Parfrey L.W., Pawlowski J., Rueckert S., Shadwick L., Schoch C.L., *Smirnov A., Spiegel F.W., le Gall L., 2012. The revised classification of eukaryotes // J. Euk. Microbiol. – Vol. 59. – P. 429–514.

2. Aescht E., Foissner W., 1992. Effects of mineral and organic fertilizers on the microfauna in a high-altitude reafforestation trial // Biol. Fertil. Soils. – Vol. 13. – P. 17–24.

3. Anderson O.R., Rogerson A., Hannah F., 1996. A description of the testate amoeba *Ovulina parva* gen. nov., sp. nov. from coastal marine sediments // J. Mar. Biol. Assoc. UK. – Vol. 76. – P. 851–865.

4. Anderson O.R., Rogerson A., Hannah F., 1997. *Ovulinata* nom. nov., a replacement name for *Ovulina* Anderson, Rogerson and Hannah, 1996 (Protista: Filosea) // J. Mar. Biol. Assoc. UK. – Vol. 77. – P. 1259.

5. Aoki Y., Hoshino M., Matsubara T., 2007. Silica and testate amoebae in a soil under pine-oak forest // Geoderma. – Vol. 142. – P. 29–35.

6. Awerintzew S., 1906. Die Susswasserrhizopoden // Trudy Imperatorskago Sankt Peterburgskago Obshchestva Estestvoisptatelei. – Vol. 36. – P. 1–346.

7. Awerintzew S., 1907. Ueber einige neue Arten gehaüsetragender Rhizopoden des Susswassers // Arch. Protistenk. – Vol. 8. – P. 86–94.

8. Badewitz H., 2004. The genus *Microcorycia* Cockerell, 1911 (Testacealobosia, Rhizopoda, Protozoa). A critical monograph of the genus including a first description of a new species: *Microcorycia scutella* n. sp. // Lauterbornia. – Vol. 50 (1984). – P. 111–146.

9. Bartoš E., 1940. Studien über die moosbewohnenden Rhizopoden der Karpaten // Arch. Protistenk. – Vol. 94. – P. 93–160.

10. Bartoš E. 1954. Koreňonožce radu Testacea. – Bratislava : Vyd. Slov. Akad. Vied. – 189 p.

11. Bartoš E. 1963. Rhizopoden einiger moosproben aus Java // Acta Univ. Carolinae. Biologica. – Vol. 2. – P. 119–190.

12. Beyens L., Chardez D., 1982. *Cryptodifflugia angustastoma* et *Nebela carinatella*, nouveaux thécamoebiens des tiurbienns dans la Campine belge // Arch. Protistenk. – Vol. 126. – P. 169–172.

13. Beyens L., Chardez D., 1984. Testate amoeba Rhizopoda from Southwest Ireland // Arch. Protistenk. – Vol. 128. – P. 109–126.

14. Beyens L., Chardez D., de Bock P., 1986. Some new and rare testate amoebae from the Arctic // Acta Protozool. – Vol. 25. – P. 81–91.

15. Beyens L., Meisterfeld R., 2002. Protozoa: Testate Amoebae // In Smol J.P., Birks H.J.B., Last W.M. (eds). Tracking environmental change using lake sediments. Vol. 3: Terrestrial, Algal, and Siliceous Indicators. Dordrecht: Kluwer Academic Publishers. – P. 121–153.

16. Bobrov A.A., 2001. *Cryptodifflugia bassini* – new species of sphagnobiontic testate amoebaes (Protozoa, Testacea) // Zool. Zhurn. – Vol. 80. – P. 1010–1013.

17. Bobrov A.A., 2016. Description of a new testate amoebae genus *Meisterfeldia* with notes on the systematics of the suborder Phryganellina (Amebozoa; Tubulinea; Arcellinida) // Acta Protozool. – Vol. 55 (4).

18. Bobrov A.A., Charman D.J., Warner B.G., 1999. Ecology of testate amoebae (Protozoa: Rhizopoda) on peatlands in Western Russia with special attention to niche separation in closely related taxa // Protist. – Vol. 150. – P. 125–136.

19. Bobrov A.A., Shimano S., Mazei Yu., 2012. Two new species of testate amoebae from mountain forest soils of Japan and redescription of the genus *Deharvengia* Bonnet, 1979 // Acta Protozool. – Vol. 51. – P. 55–63.

20. Bobrov A.A., Yazvenko S.B., Warner B.G., 1995. Taxonomic and ecological implications of shell morphology of free testaceans (Protozoa: Rhizopoda) in Russia and Canada // Arch. Protistenk. – Vol. 145. – P. 119–126.

21. Bonnet L., 1959. Nuveaux Thécamoebiens du sol. 1. // Bull. Soc. Hist. Nat. Toulouse. – Vol. 94 (1–2). – P. 177–188.

22. Bonnet L., 1960a. Thécamoebiens du sol d'Angola // Publicaçes culturais da Companhia de Diamantes de Angola. – P. 78–86.

23. Bonnet, L. 1960b. Nouveaux Thécamoebiens du sol (III): Bulletin de la Société d'Histoire Naturelle de Toulouse. – Vol. 95. – P. 209–211.

24. Bonnet L., 1962. Biologie de l'Amerique Australe // Centre National de la Recherche Scientifique. – Vol. 1. – P. 43–47.

25. Bonnet L., 1964. Le peuplement thecamoebien des sols // Rev. Ecol. Biol. Sol. – Vol. 1. – P. 123–408.

26. Bonnet L., 1965. Nouveaux thecamoebiens du sol // Bulletin de la Societe d'Histoire Naturelle de Toulouse. – Vol. 100. – P. 330–332.

27. Bonnet L., 1974. Les Lamtopyxidae fam. nov. et la structure propilostome chez les Thecamoebiens (Rhizopoda, Testacea) // C. r. Séanc. Acad. Sci., Paris. – Vol. 278. – P. 2935–2937.

28. Bonnet L., 1975. *Lamtoquadrula* gen. nov. et la structure plagiostome chez les Thecamoebiens nebeliformes // Bulletin de la Societe d'Histoire Naturelle de Toulouse. – Vol. 110. – P. 297–299.

29. Bonnet L., 1975a. Nouveaux Thecamoebiens du sol (VII) // Bulletin de la Societe d'Histoire Naturelle de Toulouse. – Vol. 110. – P. 283–290.

30. Bonnet L., 1977. Nouveaux thecamoebiens du sol (IX) // Bull. Soc. Hist. Natur. Toulouse. – Vol. 113. – P. 152–156.

31. Bonnet L., 1979. Nouveaux Thecamoebiens du sol (X) // Bulletin de la Societe d'Histoire Naturelle de Toulouse. – Vol. 115. – P. 106–118.

32. Bonnet L., Thomas R., 1960. Thécamoebiens du sol // Hermann (ed.), Faune terrestre et d'eau douce des Pyrénées-Orientales. (Supplement to Vie et Milieu). – Vol. 5. – 113 p.

33. Booth R.K., 2001. Ecology of testate amoebae (Protozoa) in two lake superior coastal wetlands: Implications for paleoecology and environmental monitoring // Wetlands. – Vol. 21. - P.564-576.

34. Burki F., Shalchian-Tabrizi K., Minge M., Skjaeveland A., Nikolaev S., Jakobsen K.S., Pawlowski J., 2007. Phylogenomics reshuffles the eukaryotic supergroups // PLoS One. – Vol. 2 (8). – P. 790.

35. Carlson M.L., Flagstad L.A., Gillet F., Mitchell E.A.D., 2010. Community development along a proglacial chronosequence: are aboveground and below-ground community structure controlled more by biotic than abiotic factors // J. Ecol. – Vol. 98. – P. 1084–1095.

36. Cash J., 1904. On some new and little known British freshwater Rhizopoda // J. Linn. Soc. London, Zoology. – Vol. 29. – P. 218–225.

37. Cash J., Hopkinson J., 1905. The British freshwater Rhizopoda and Heliozoa. Vol. I. Rhizopoda. Part I: London: Ray Society publication. – Vol. 85. – 151 p.

38. Cash J., Hopkinson J., 1909. The British freshwater Rhizopoda and Heliozoa. Vol.II. Rhizopoda. Part II. London: Ray Society publication. – Vol. 89. – 166 p.

39. Cash J., Wailes G.H., Hopkinson J., 1915. The British Freshwater Rhizopoda and Helioza. Vol. III. Rhizopoda. Part III. London: Ray Society publication. – Vol. 98. – 156 p. 40. Cash J., Wailes G.H., Hopkinson J., 1919. The British freshwater Rhizopoda and Heliozoa. Vol. IV. London: Ray Society publication. – Vol. 103. – 130 p.

41. Cavalier-Smith T., 2010. Kingdoms Protozoa and Chromista and the eozoan root of the eukaryotic tree // Biology Letters. – Vol. 6. – P. 342–345.

42. Certes A., 1889. Protozoaires // Mission scientifique du Cap Horn 1882–1883. – Vol. 6. – 53 p.

43. Chardez D., 1958. Etude sur les thecamoebiens d'une petite piece d'eau // Hydrobiologia. – Vol. 10. – P. 292–304.

44. Chardez D., 1960. Note sur les Thecamoebiens d'Otrange (Hesbaye) et decouverte d'un genere nouveau // Ann. Soc. Royal. Zool. Belgique. – Vol. 91. – P. 39–43.

45. Chardez D., 1960. Sur quelques Thecamoebiens du genre *Trinema* Dujardin // Bull. Inst. Agron. Stat. Rech. Gembloux. – Vol. 28. – P. 266–271.

46. Chardez D., 1961a. Note sur *Phryganella acropodia* (Hertwig et Less) Hopkinson et sa variete *penardi* Declootre (Protozoa Rhizopoda) // Bull. Inst. Agron. Stat. Rech. Gembloux. – Vol. 29(2). – P. 122–124.

47. Chardez D., 1961. Sur *Difflugia acuminata* Ehrenberg (Rhizopoda, Testacea) // Bull. Inst. Agron. Stat. Rech. Gembloux. – Vol. 29 (3–4). – P. 301–308.

48. Chardez D., 1962. Deux vaeietes nouvelles de Thecamoebiens, Rhizopoda testacea // Bull. Inst. Agron. Stat. Rech. Gembloux. – Vol. 30 (3–4). – P. 260–262.

49. Chardez D., 1963. Thecamoebiens des Hautes Fagnes en Belgique // Revue des Hautes Fagnes. – Vol. 3. – P. 142–153.

50. Chardez D., 1964. Thecamoebiens (Rhizopodes testaces) // Symoens J.-J. (ed.): Exploration hydrobiologique du bassin du Lac Bangweolo et du Luapula. Resultats scientifiques, Thecamoebiens. Bruxelles: Cercle Hydrobiologique de Bruxelles. – Vol. 10 (2). – 53 p.

51. Chardez D., 1966. Note sur quelques Thecamoebiens // Bull. Inst. Agron. Stat. Rech. Gembloux, new series. – Vol. 1 (1). – P. 20–24.

52. Chardez D., 1967a. Histoire Naturelle des Protozoaires Thecamoebiens // Naturalistes Belges. – Vol. 48 (10). – P. 484–576.

53. Chardez D., 1967b. Monographie du genre *Quadrulella* Cockerell (Protozoa, Rhizopoda, Testacea) // Bull. Inst. Agron. Stat. Rech. Gembloux, new series. – Vol. 2 (2). – P. 230–247.

54. Chardez D., 1967c. *Difflugia oblonga* Ehrenberg et ses varietes // Bull. Inst. Agron. Stat. Rech. Gembloux, new series. – Vol. 2 (4). – P. 589–595.

55. Chardez D., 1970. Etude sur *Centropyxis aculeata* (Ehrenberg) Stein (Protozoa Rhizopoda Testacea) // Bull. Rech. Agron. Gembloux, new series. – Vol. 5 (1–2). – P. 76–86.

56. Chardez D., 1971. Etude sur les thecamoebiens des biotopes interstitiels psammons littoraux et zones marginales souterraines des eaux douces // Bull. Rech. Agron. Gembloux, new series. – Vol. 6 (3–4). – P. 257–268.

57. Chardez D., 1985. Observations sur la repartition des Thecamoebiens dans un lac // Acta Protozool. – Vol. 24. – P. 217–223.

58. Chardez D. 1990, Contribution a la connaissance des Thecamoebiens aquatiques du Tyrol allemand (Rhizopoda, Testacea) // Acta Protozool. – Vol. 29. – P. 153–156.

59. Chardez D. 1991, Le genre *Cyphoderia* Schlumberger, 1845 (Protozoa: Rhizopoda: Testacea) // Acta Protozool. – Vol. 30. – P. 49–53.

60. Chardez D., Beyens L., 1987. *Arcella ovaliformis* sp. nov., a new testate amoeba from Edgeøya, a High Artic Island // Arch. Protistenk. – Vol. 134. – P. 297–301.

61. Chardez D., Beyens L., de Bock P., 1988. *Centropyxis gasparella* sp. nov. and *Parmulina louisi* sp. nov., new testate amoebae from the Canadian High Arctic (Devon Island, NWT) // Arch. Protistenk. – Vol. 136. – P. 337–344.

62. Chardez D., Gaspar Ch., 1984. Nouveaux Thecamoebiens aquatiques du domaine des Epioux (Ardenne, Belgique) // Biologish Jaarb. Dodonaea. – Vol. 52. – P. 57–63.

63. Charman D.J., Blundell A., Members A., 2007. A new European testate amoebae transfer function for palaeohydrological reconstruction on ombrotrophic peatlands // J. Quat. Sci. – Vol. 22. – P. 209–221.

64. Charman D.J., Hendon D., Woodland W.A., Woodland A., 2000. The identification of testate amoebae (Protozoa: Rhizopoda) in peats. QRA Technical Guide No 9. Quaternary Research Association, London.

65. Corbet S.A., 1973. An illustrated introduction to the testate Rhizopods in *Sphagnum* with special reference to the area around Malham Tarn, Yorkshire // Field Studies. – Vol. 3. – P. 801–838.

66. Coûteaux M.M., 1976. Dynamisme de l'equilibre des Thecamoebiens dans quelques sols climaciques // Mem. Mus. Nat. Hist. Nat. Nouv. ser. Ser. A. Zool. – Vol. 96. – P. 1–183.

67. Coûteaux M.M., Chardez D., 1981. Thécamoebiens édaphiques et muscicoles de Guyane Française // Revue d'Écologie et de Biologie du Sol. – Vol. 18. – P. 193–208.

68. Coûteaux M.M., Munsch A., Ponge J.-F., 1979. Le genre *Euglypha*: essai de taxinomie numerique // Protistologica. – Vol. 15 (4). – P. 565–579.

69. Cunha A.M. da, 1913. Contribuico para o conhecimento da fauna de protozoarios do Brazil // Mem. Inst. Oswaldo Cruz. - Vol. 5 (2). - P. 101–122.

70. Daday J., 1905. Untersuchungen uber die Susswasser-Mikrofauna Paraguays // Zoologica. – Vol. 18 (44). – P. 1–374.

71. Décloître L., 1949. Materiaux pour une faune Rhizopodique d'A.O.F. // Bull. Inst. Franc. Afr. Noire. – Vol. 11 (3). – P. 281–301.

72. Décloître L., 1954. Contribution a l'etude du peuplement de la Mauritanie // Bull. Inst. Franc. Afr. Noire. – Vol. 16 (2). ser. A. – P. 398–413.

73. Décloître L., 1955. Speleologia Africana. Thecamoebiens de la grotte des Singes a Segea (Guinee) // Bull. Inst. Franc. Afr. Noire. – Vol. 17 (4). ser. A. – P. 989–1019.

74. Décloître L., 1956. Les Thecamoebiens de l'Eqe (Groenland) // Expeditions Polaires Francaises – Missions Paul-Emile Victor. – Vol. VIII. – 100 p. (Actualites Scientifiques et Industrialles, № 1242).

75. Décloître L. 1962a. Le Genre *Paraquadrula* (Thekamoebina) // Internationale Revue dergesamten Hydrobiologie und Hydrographie. – T. 47. № 2. – P. 321–330.

76. Décloître L., 1962b. Le genre *Euglypha* Dujardin // Arch. Protistenk. – Vol. 106. – P. 51–100.

77. Décloître L., 1962c. Thecamoebiens de la XII Expedition Antartique Francaise // Territoires des terres australes et antartiques francaises, publication N_{2} 259. – 47 p.

78. Décloître L. 1964. Materiaux pour une faune thecamoebienne du Maroc // Bulletin de la Societe des Sciences Naturelles et Physiques du Maroc. – Vol. 44. – P. 121–136.

79. Décloître L., 1965. Amoebida testacea (Rhizopoda) // Bertelsen E.et al. (eds.). The Zoology of Iceland. Copenhagen and Reykjavik: Ejnar Munksgaard. – Vol. 2 (1). – 58 p.

80. Décloître L., 1966. Compléments à The Zoology of Iceland. Vol. II. Part I. Amoebida Tesatcea (Rhizopoda) // Videnskabelige Meddelelser fra dansk naturhistorisk forening. – P. 129. – P. 67–71.

81. Décloître L., 1972. Thecamoebiens du Var (Suite) // Ann. Soc. Sci. Nat. Archeol. Toulon Var. – Vol. (24). – P. 164–171.

82. Décloître L., 1973. Thecamoebiens des Iles Galapagos // Ciencia y Naturaleza. – Vol. 19 (1). – P. 11–20.

83. Décloître L., 1976. Le genre *Arcella* Ehrenberg, complements a jour au 31 decembre 1974 de la monographie du genre parue en 1928 // Arch. Protistenk. – Vol. 118. – P. 291–309.

84. Décloître L., 1977a. Le genre *Cyclopyxis*. Complements a jour au 31 decembre 1974 de la monographie du genre parve en 1929 // Arch. Protistenk. – Vol. 119. – P. 31–53.

85. Décloître L., 19776. Le genre *Nebela*. Complements a jour au 31 december 1974 de la monographie du genre parve en 1936 // Arch. Protistenk. – Vol. 119. – P. 325–352.

86. Décloître L., 1978. Le genre *Centropyxis* I. Complements a jour au 31 decembre 1974 de la Monographie du genre parve en 1929 // Arch. Protistenk. – Vol. 120. – P. 63–85.

87. Décloître L., 1979. Le genre *Centropyxis* II. Complements a jour au 31 decembre 1974 de la Monographie du genre parue en 1929 // Arch. Protistenk. – Vol. 121. – P. 162–192.

88. Décloître L., 1981. Le genre *Trinema* Dujardin, 1841. Revision a jour au 31 XII. 1979 // Arch. Protistenk. – Vol. 124. – P. 193–218.

89. Deflandre G., 1928a. Le genre *Arcella* Ehrenberg. Morphologie-Biologie. Essai phylogenetique et systematiqe // Arch. Protistenk. – Vol. 64. – P. 152–287.

90. Deflandre G., 1928b. Deux genres nouveaux de Rhizopodes testaces. 1. *Wailesella* gen. nov. 2. *Tracheleuglypha* gen. nov. // Ann. Protistol. – Vol. 1. – P. 37–43.

91. Deflandre G., 1929. Le genre *Centropyxis* Stein // Arch. Protistenk. – Vol. 67. – P. 322–375.

92. Deflandre G., 1931. Thecamoebiens nouveaux ou peu connus, I // Ann. Protistol. – Vol. 3. – P. 81–95.

93. Deflandre G., 1936. Etude monographique sur le genre *Nebela* Leidy (Rhizopoda-Testacea) // Ann. Protistol. – Vol. 5. – P. 201–322.

94. Deflandre G., 1953. Ordres des Testaceolobosa (de Saedeleer, 1934), Testaceafilosa (de Saedeleer, 1934), Thalamia (Haeckel, 1862) ou Thecamoebiens (Auct.) (Rhizopoda Testacea). In: Grasse P.-P., (ed.), Traite de Zoologie, Paris: Masson and Co, – Vol. I, fasc. II. – P. 97–148.

95. Dekhtiar M.N., 1993. New species of the family Difflugiidae (Lobosea, Rhizopoda) with remarks on validity of the genus *Protocucurbitella* // Zool. Zhurn. – Vol. 72. – P. 5–15.

96. Dekhtiar M.N., 1994. The new and rare species of testate amebae of the families Arcellidae, Centropyxidae, Lesquereusiidae and Hyalospheniidae (Rhizopoda, Testacealobosia) // Zool. Zhurn. – Vol. 73. – P. 3–14.

97. Dekhtiar M.N., 2009. The peculiarities of shell morphology and taxonomic composition of the family Centropyxidae (Testa-cealobosia, Rhizopoda) // Vestnik zoologii. – Vol. 43 (4). – P. 297–303.

98. Dujardin F., 1841. Histoire naturelle des Zoophytes. Infusoires, comprenant la physiologie et la classification de ces animaux, et la mani μ re de les etudier a l'aide du microscope. Paris. – Vol. 1. – 684 p.

99. Edmondson C.H., Kingman A., 1914. Notes on japanese Protozoa // Annot. Zool. Jap. Tokyo. – Vol. 8. – P. 531–542.

100. Ehrenberg C.G., 1838. Die infusionthierchen als Vollkommene Organismen. Verlag. von Leopold Voss. Leipzig. – 547 p.

101. Ellison R.L., Ogden C.G., 1987. A guide to the study and identification of fossil testate amoebae in Quaternary lake sediments // Int. Rev. ges. Hydrobiol. – Vol. 72 (5). – P. 639–652.

102. Finlay B.J., Black H.I.J., Brown S., Clarke K.J., Esteban G.F., Hindle R.M., Olmo J.L., Rollett A., Vickerman K., 2000. Estimating the growth potential of the soil protozoan community // Protist. – Vol. 151. – P. 69–80.

103. Francé R.H., 1913. Das Edaphon, Untersuchungen zur Oekologie der bodenbewohnenden Mikroorganismen // Arb. Biologisch. Inst. München. – Vol. 2. – P. 1–99.

104. Foissner W., 1997. Protozoa as bioindicators in agroecosystems, with emphasis on farming practices, biocides, and biodiversity // Agriculture Ecosystems and Environment. – Vol. 62. – P. 93–103.

105. Foissner W., Korganova G.A. 1995. Redescription of three testate amoebae (Protozoa, Rhizopoda) from a Caucasian soil: *Centropyxis plagiostoma* Bonnet et Thomas, *Cyclopyxis kahli* (Deflandre) and *C. intermedia* Kufferath // Arch. Protistenk. – Vol. 146. – P. 13–28.

106. Fournier B., Malysheva E., Mazei Yu., Moretti M., Mitchell E.A.D., 2012. Toward the use of testate amoeba functional traits as indicator of floodplain restoration success // Eur. J. Soil Biol. – Vol. 49. – P. 85–91.

107. Gauthier-Lièvre L., 1953. Les genres *Nebela*, *Paraquadrula* et *Pseudonebela* (Rhizopodes testaces) en Afrique // Bull. Soc. Hist. Nat. Afr. Nord. – Vol. 44 (7–8). – P. 324–366.

108. Gauthier-Lièvre L., 1957. Addition aux *Nebela* d'Afrique // Bull. Soc. Hist. Nat. Afr. Nord. – Vol. 48. – P. 494–523.

109. Gauthier-Lièvre L., Thomas R., 1958. Les genres *Difflugia*, *Pentagonia*, *Maghrebia* et *Hoogenraadia* (Rhizopodes testaces) en Afrique // Arch. Protistenk. – Vol. 103 (1–2). – P. 241–370.

110. Gauthier-Lièvre L., Thomas R., 1960. Le genre *Cucurbitella* Penard // Arch. Protistenk. – Vol. 104 (4). – P. 569–602.

111. Gilbert D., Amblard C., Bourdier G., Francez A.J., Mitchell E.A.D., 2000. Le régime alimentaire des thécamoebiens // L'Année Biologique. – Vol. 39. – P. 57–68.

112. Geltzer Yu.G., Korganova G.A., Alekseev D.A., 1985. Soil testate amoebae and methods of their research. – Moscow : MSU Publ. – 79 p.

113. Geltzer Yu.G., Korganova G.A., Alekseev D.A., 1995. Identification guide for soil testate amoebae (a practical guide). – Moscow : MSU Publishing house. – P. 1–88.

114. Godeanu S., 1972. Especes nouvelles de thecamoebiens (Protozoa, Rhizopoda, Arcellinida) // Rev. Roum. Biol. (ser. Zool.). – Vol. 17. – P. 227–236.

115. Golemansky V.G., 1964. Thecamoebiens nouveaux et peu connus // Acta Protozool. – Vol. 2. – P. 197–200.

116. Golemansky V., 1968. Materiaux sur la faune thecamoe-bienne (Rhizopoda, Testacea) de Cuba // Acta Protozoologica. – Vol. 6. – P. 335–340.

117. Golemansky V., 1970a. Rhizopodes nouveaux du psammon littoral de la mer Noire // Protistologica. – Vol. 6. – P. 365–371.

118. Golemansky V., 1970b. Contribution a la connaissance des Thecamoebiens (Rhizopoda, Testacea) des eaux souterraines littorales du Golf de Gdansk (Pologne) // Bull. Inst. Zool. Acad. Bulg. Sci. (Sofia). – Vol. 31. – P. 77–87.

119. Golemansky V., 1973. *Messemvriella filosa* n. gen. n. sp.- une nouvelle thecamoebienne psammobionte (Rhizopoda, Testacea) des eaux souterraines littorales de la Mer Noire // Zoologische Anzeiger. – Vol. 190. – P. 302–304.

120. Golemansky V., 1978. Adaptations morphologiques des thecamoebiens psammobiontes du psammal supralittoral des mers // Acta Protozool. – Vol. 17 (1). – P. 141–152.

121. Golemansky V., 1979 Thécamoebiens psammobiontes du supralittoral vietnamien de la Mer Chinoise et description de *Cryptodifflugia brevicolla* sp. n. (Rhizopoda: Arcellinida) // Acta Protozool. – Vol. 18. – P. 285–292.

122. Golemansky V., 1981. Descrption de trois thecamoebiens (Protozoa: Rhizopodea) nouveaux des eaux souterraines littorales des mers // Acta Protozool. – Vol. 20. – P. 115–119.

123. Golemansky V., 1990. Interstitial testate amoebas (Rhizopoda: Testacea) from the Mediterraneen Basin // Stygologia. – Vol. 5. – P. 49–54. 124. Golemansky V., 1994. Thecamoebiens (Rhizopoda: Testacea) d'hydroppsammon littoral de trois lacs Macedoniens d'origine tectonique: Ohrid, Prespa et Doiran // Arch. Protistenk. – Vol. 144. – P. 309–313.

125. Golemansky V., 2000. Marine interstitial rhizopods (Rhizopoda: Arcellinida, Gromida and Foraminiferida) from the South-West Atlantic (Region of Rio de Janeiro) and Description of *Psammolagynis atlantica* gen. n., sp. n. // Acta Zool. Bulg. – Vol. 52. – P. 3–12.

126. Golemansky V., Todorov M., 1999. First report of the interstitial testate amoebae (Protozoa: Testacea) in the marine supralittoral of the Livingston Island (the Antarctic) // Bulg. Antarct. Res. – Life Sci. – Vol. 2. – P. 43–47.

127. Gomaa F., Kosakyan A., Heger T.J., Corsaro D., Mitchell E.A.D., Lara E., 2014. One alga to rule them all: Unrelated mixotrophic testate amoebae (Amoebozoa, Rhizaria and Stramenopiles) share the same symbiont (Trebouxiophyceae) // Protist. – Vol. 165. – P. 161–176.

128. Gomaa F., Mitchell E.A.D., Lara E., 2013. Amphitremida (Poche, 1913) is a new major, ubiquitous Labyrinthulomycete clade // PLoS ONE. – Vol. 8. – P. 53 046.

129. Gomaa F., Todorov M., Heger T.J., Mitchell E.A.D., Lara E., 2012. SSU rRNA phylogeny of Arcellinida (Amoebozoa) reveals that the largest Arcellinid genus, *Difflugia* Leclerc 1815, is not monophyletic // Protist. – Vol. 163. – P. 389–399.

130. Gomaa F., Lahr D., Li J., Todorov M, Lara E., 2017. Phylogenetic reconstruction based on SSU rDNA demonstrates that genus *Difflugia* (Arcellinida: Amoebozoa) is paraphyletic, consisting of two major unrelated groups characterized by test shape // Eur. J. Protistol. (in press).

131. Graaf F. de, 1952. A new Rhizopode of the genus *Hyalosphenia* Stein 1857 (Protozoa; Sarcodina; Amoebina) // Beaufortia. – Vol. 23.

132. Grospietsch T., 1958. Wechseltierchen (Rhizopoden). Stuttgart: Kosmos-Gesellschaft der Naturfreunde/Franckh'sche Verlagshandlung; Sammlung Einfuhrung in die Kleinlebewelt. – 82 p.

133. Grospietsch T., 1965. Monographische Studie der Gattung *Hyalosphenia* Stein (Rhizopoda Testacea) // Hydrobiologia. – Vol. 26. – P. 211–241.

134. Grospietsch T., 1964. Die Gattungen *Cryptodifflugia* und *Dif-flugiella* (Rhizopoda Testacea) // Zool. Anz. – Vol. 172 (4). – P. 243–257.

135. Harnish O., 1924. Studien zur Okologie der Moorfauna // Biologisches Zentralblatt. – Vol. 44. – P. 110–127.

136. Harnish O., 1925. Studien zur Okologie und Tiergeographie der Moore // Zoologisch Jahrbuch (Abteilung Systematik). – Vol. 51. – P. 1–166.

137. Harnish O., 1927. Einige Daten zur rezenten und fossilen testaceen Rhizopodenfauna der Sphagnen // Arch. Hydrobiol. – Vol. 18. – P. 345–360.

138. Harnisch O., 1958. Wurzelfussler, Rhizopoda // Tierwelt Mitteleuropas (eds. Brohmer P. et al.). – Vol. 1. – P. 1–75.

139. Heal O.W., 1961. The distribution of testate amoebae (Rhizopoda, Testacea) in some fens and bogs in northern England // J. Linn. Soc. Zoology. – Vol. 30. – P. 369–382.

140. Heal O.W., 1962. The abundance and micro-distribution of testsate amoeba (Rhizopoda: Testacea) in *Sphagnum* // Okios. – Vol. 13. – P. 35–47.

141. Heal O.W., 1963. Soil fungi as food for amoebae // Soil Organisms (eds. J. Doeksen, J. et van der Drift). – Amsterdam : North-Holland. – P. 289–297.

142. Hoogenraad H.R., de Groot A.A. 1940a. Zoetwaterrhizopoden en heliozo π // A.W.Sijthoff (ed.). Fauna von Nederland. Afl. – Vol. 9. – 303 p.

143. Hoogenraad H.R., de Groot A.A., 1948. Thecamoebous mossrhizopods from New Zealand // Hydrobiologia. – Vol. 1. – P. 28–43.

144. Howe A.T., Bass D., Scoble J.M., Lewis R., Vickerman K., Arndt H., Cavalier-Smith T., 2011. Novel cultured protists identify deepbranching environmental DNA clades of Cercozoa: new genera *Tremula*, *Micrometopion*, *Minimassisteria*, *Nudifila*, *Peregrinia* // Protist. – Vol. 162. – P. 332–372.

145. Iudina T.A., 1996. A comparative study of testaceans of the genus *Corythion* Taranek, 1882 (Sarcodina, Filosea) // Zool. Zh. – Vol. 75. – P. 609–612.

146. Jung W., 1936. Thekamoben eines Eggegebirgsmoores und zweier Moore im Hohen Venn // Ann. Protistol. – Vol. 5. – P. 83–123.

147. Jung W., 1942. Sudchilenische Thekamoben (Aus dem sudchilenischen Kustengebiet, Beitrag 10) // Arch. Protistenk. – Vol. 95 (3). – P. 253–356.

148. Jung W., 1942a. Illustrierte Thekamoben-Bestimmungsta-bellen. I. Die Systematik der Nebelinen // Arch. Protistenk. – Vol. 95 (3). – P. 357–390.

149. Kosakyan A., Gomaa F., Mitchell E.A.D., Heger T.J., Lara E., 2013. Using DNA-barcoding for sorting out protist species complexes: A case study of the *Nebela tincta-collaris-bohemica* group (Amoebozoa; Arcellinida, Hyalospheniidae) // Eur. J. Protistol. – Vol. 49. – P. 222–237.

150. Kosakyan A., Gomaa F., Lara E., Lahr D.J.G., 2016. Current and future perspectives on the systematics, taxonomy and nomenclature of testate amoebae // Eur. J. Protistol. – Vol. 55. Part B. – P. 105–117.

151. Kosakyan A., Heger T.J., Leander B.S., Todorov M., Mitchell E.A.D., Lara E., 2012. COI barcoding of Nebelid testate amoebae (Amoebozoa: Arcellinida): extensive cryptic diversity and redefinition of the Hyalospheniidae Schultze // Protist. – Vol. 163. – P. 415–434.

152. Kosakyan A., Lahr D.J.G., Mulot M., Meisterfeld R., Mitchell E.A.D., Lara E., 2016a. Phylogenetic reconstruction based on COI reshuffles the taxonomy of hyalosphenid shelled (testate) amoebae and reveals the convoluted evolution of shell plate shapes // Cladistics (in press).

153. Krashevska V., Maraun M., Ruess L., Scheu S., 2010. Carbon and nutrient limitation of soil microorganisms and microbial grazers in a tropical montane rain forest // Oikos. – Vol. 119. – P. 1020–1028.

154. Krashevska V., Bonkowski M., Maraun M., Ruess L., Kandeler E., Scheu S., 2008. Microorganisms as driving factors for the community structure of testate amoebae along an altitudinal transect in tropical mountain rain forests // Soil Biol. Biochem. – Vol. 40. – P. 2427–2433.

155. Kufferath H., 1932. Rhizopodes du Congo // Rev. Zool. Bot. Afr. – Vol. 23 (1). – P. 52–60.

156. Lagerheim G., 1902. Om *Quadrula subglobosa* Lagerheim // Geologiska Foreningen i Stockholm Forhandlingar. – Vol. 24. – P. 346–352.

157. Lahr D.J.G., Parfrey L.W., Mitchell E.A.D., Katz L., Lara E., 2011. The chastity of amoebae: re-evaluating evidence for sex in amoeboid organisms // Proc. Biol. Sci. Royal Soc. – Vol. 278. – P. 2081–2090.

158. Lamentowicz M., Mitchell E.A.D., 2005. The ecology of testate amoebae (Protists) in *Sphagnum* in North-western Poland in relation to peatland ecology // Microb. Ecol. – Vol. 50. – P. 48–63.

159. Laminger H., 1971. Sedimentbewohnende Schalenamoben (Rhizopoda Testacea) der Finstertaler Seen (Tirol) // Arch. Hydrobiol. – Vol. 69 (1). – P. 106–140.

160. Laminger H., 1972. Die profundale Testaceenfauna (Protozoa Rhizopoda) alterer und jungerer Bodensee-Sedimente // Arch. Hydrobiol. – Vol. 70 (1). – P. 108–129.

161. Laminger H., 1973. Die Testaceen (Protozoa, Rhizopoda) einiger Hochgebirgsgewasser von Mexiko, Costa Rica und Guatemala // Intern. Rev. Ges. Hydrobiol. – Vol. 58 (2). – P. 273–305.

162. Lauterborn R., 1908. Protozoen Studien. V. Zur Kenntniss einiger Rhizopoden und Infusorien aus dem Gebiete des Oberrheins // Zeitsch. Wissenschaft. Zool. – Vol. 90. – P. 645–669.

163. Leclerc L., 1816. Note sur la Difflugie, nouveau genre de Polype amorph // Mem. Mus. Hist. Nat. (Paris). – Vol. 2 (12). – P. 474–478.

164. Leidy J., 1879. Fresh-water Rhizopods of North America // Rep. US Geol. Surv. Terr. – Vol. 12. – 324 p.

165. Levine N.D., Corliss J.O., Cox F.E.G., Deroux G., Grain J., Honigberg B.M., Leedale G.F., Loeblich A.R., Lom J., Lynn D.H., Merinfeld D., Page F.C., Poljansky G., Sprague V., Vavra J., Wallace F.G., 1980. A newly revised classification of the Protozoa // J. Protozool. – Vol. 27. – P. 37–58.

166. Lüftenegger G., Petz W., Berger H., Foissner W., Adam H., 1988. Morphologic and biometric characterzization of twenty-four soil testate amoebae (Protozoa, Rhizopoda) // Arch. Protistenk. – Vol. 136. – P. 153–189.

167. Lüftenegger G., Foissner W., 1991. Morphology and biometry of twelwe soil testate amoebae (Protozoa, Rhizopoda) from Australia, Africa, and Austria // Bull. Brit. Mus. nat. Hist. (Zool.). – Vol. 57 (1). – P. 1–16.

168. Mazei Yu., Tsyganov A., 2006. Freshwater Testate Amoebae. – Moscow : KMK Sci Press. – 300 p.

169. Mazei Yu., Warren A., 2012. A survey of the testate amoeba genus *Difflugia* Leclerc, 1815 based on specimens in the E. Penard and C.G. Ogden collections of the Natural History Museum, London. Part 1: Species with shells that are pointed aborally and/or have aboral protuberances // Protistology. – Vol. 7 (3). – P. 121–171.

170. Mazei Yu., Warren A., 2014. A survey of the testate amoeba genus *Difflugia* Leclerc, 1815 based on specimens in the E. Penard and C.G. Ogden collections of the Natural History Museum, London. Part 2: Species with shells that are pyriform or elongate // Protistology. – Vol. 8 (4). – P. 133–171.

171. Mazei Yu., Warren A., 2015. A survey of the testate amoeba genus *Difflugia* Leclerc, 1815 based on specimens in the E. Penard and C.G. Ogden collections of the Natural History Museum, London. Part 3: Species with shells that are spherical or ovoid // Protistology. – Vol. 9 (1). – P. 3–49.

172. Meisterfeld R. 2002a. Order Arcellinida Kent, 1880 // Lee J. J., Leedale G. F., Bradbury P. (eds.). The Illustrated Guide to the Protozoa, Society of Protozoologists, Lawrence, Kansas, USA. – P. 827–860.

173. Meisterfeld R. 2002b. Testate amoebae with filopodia // Lee J. J., Leedale G. F., Bradbury P. (eds.). The Illustrated Guide to the Protozoa. Society of Protozoologists, Lawrence, Kansas, USA. – P. 1054–1084.

174. Meisterfeld R., Badewitz H.J., 2006. A redescription of *Amphizonella violacea* (Amoebozoa: Arcellinida) // Acta Protozoolo. – Vol. 45. – P. 167–173.

175. Mitchell E.A.D., 2004. Response of testate amoebae (Protozoa) to N and P fertilization in an arctic wet sedge tundra // Arct. Antarct. Alp. Res. – Vol. 36. - P. 77-82.

176. Mitchell E.A.D., Bragazza L., Gerdol R., 2004. Testate amoebae (Protista) communities in *Hylocomium splendens* (Hedw.) B.S.G. (Bryophyta): relationships with altitude and moss elemental chemistry // Protist. – Vol. 155. – P. 423–436.

177. Mitchell E.A.D., Buttler A.J., Warner B.G., Gobat J.M., 1999. Ecology of testate amoebae (Protozoa: Rhizopoda) in *Sphagnum* peatlands in the Jura mountains, Switzerland and France // Ecoscience. – Vol. 6. – P. 565–576.

178. Mitchell E.A.D., Charman D.J., Warner B.G., 2008. Testate amoebae analysis in ecological and paleoecological studies of wetlands: past, present and future // Biodivers. Conservat. – Vol. 17. – P. 2115–2137.

179. Motti F., 1961. Generos de Tecamebianos de la Republica Argentina y una especie nueva de Arcella // Physis – Rev. Soc. Argent. Cienc. Nat. – Vol. 19 (51).

180. Nasser N.A., Patterson R.T., 2015. *Conicocassis* a new genus of Arcellinina (testate lobose amoebae) // Palaeontologia Electronica. – Vol. 18. 3.46A. – P. 1–11.

181. Nicholls K.H., 2006a. *Cryptodifflugia leachi* n. sp., a minute new testate rhizopod species (Rhizopoda: Phryganellina) // Acta Protozool. – Vol. 45. – P. 295–299.

182. Nicholls K.H., 2006b. Form variation in *Puytoracia bergeri* Bonnet and a description of P. bonneti n. sp. (Rhizopoda, Filosea) // European Journal of Protistology. – Vol. 42. – P. 155–163.

183. Nicholls K.H., 2009a. Six new marine species of the genus *Paulinella* (Rhizopoda: Filosea, or Rhizaria: Cercozoa) // J. Mar. Biol. Ass. UK. – Vol. 89. – P. 1415–1425.

184. Nicholls K.H., 2009b. A multivariate statistical evaluation of the "acolla-complex" of *Corythionella* species, including a description of C. darwini n. sp. (Rhizopoda: Filosea or Rhizaria: Cercozoa) // Eur. J. Protistol. – Vol. 45. – P. 183–192.

185. Nikolaev S.I., Mitchell E.A.D., Petrov N.B., Berney C., Fahrni J., Pawlowski J., 2005. The testate lobose amoebae (Order Arcellinida Kent, 1880) finally find their home within Amoebozoa // Protist. – Vol. 156. – P. 191–202.

186. Nguyen-Viet H., Gilbert D., Mitchell E.A.D., Badot P.M., Bernard N., 2007. Effects of experimental lead pollution on the microbial communities associated with *Sphagnum fallax* (Bryophyta) // Microb. Ecol. – Vol. 54. – P. 232–241.

187. Ogden C.G., 1979. Comparative morphology of some pyriform species of *Difflugia* (Rhizopoda) // Arch. Protistenk. – Vol. 122. – P. 143–153.

188. Ogden C.G., 1980a. Shell structure in some pyriform species of *Difflugia* (Rhizopoda) // Arch. Protistenk. – Vol. 123 (4). – P. 455–470.

189. Ogden C.G., 1980b. Notes on some Difflugiidae from Norfolk (Rhizopodea, Protozoa) // Bull. Brit. Mus. (Nat. Hist.) Zool. Ser. – Vol. 39 (3). – P. 125–138.

190. Ogden C.G., 1983. Observations on the systematics of the genus *Difflugia* in Britain (Rhizopoda, Protozoa) // Bull. Brit. Mus. (Nat. Hist.) Zool. Ser. – Vol. 44 (1). – P. 1–73.

191. Ogden C.G., 1984. Notes on testate amoebae (Protozoa: Rhizopoda) from Lake Vlasina, Yugoslavia // Bull. Brit. Mus. (Nat. Hist.) Zool. Ser. – Vol. 47 (5). – P. 241–263.

192. Ogden C.G., Fairman S., 1979. Further observations on pyriform species of *Difflugia* (Rhizopodea) // Arch. Protistenk. – Vol. 122. – P. 372–381.

193. Ogden C.G., Hedley R.H., 1980. An atlas of freshwater testate amoebae. – London : Oxford Univ. Press. – 222 p.

194. Ogden C.G., Meisterfeld R., 1989. The taxonomy and systematics of some species of *Cucurbitella*, *Difflugia* and *Netzelia* (Protozoa: Rhizopoda), with an evaluation of diagnostic characters // Eur. J. Protistol. – Vol. 25. – P. 109–128.

195. Ogden C.G., Živković A., 1983. Morphological studies on some Difflugidae from Yugoslavia (Rhizopoda, Protozoa) // Bull. Brit. Mus. (Nat. Hist.) Zool. Ser. – Vol. 44 (6). – P. 341–375.

196. Olivier A., 1945. Decription d'une nouvelle espèce d'A*r*cella // Bull. Soc. Zool. France. – Vol. 69. – P. 4–5.

197. Oye P. van, 1926. Six Rhizopodes nouveau du Congo-Belge // Arch. de Zool. Exp. et Génér. – Vol. 65 (3). – P. 64–74. 198. Oye P. van, 1932. Rhizopoda from South Africa // Revue de Zoologie et de Botanique Africaines. – Vol. 31. – P. 54–73.

199. Oye P. van, 1949. Rhizopodes de Java // Bijdragen tot de Dierkunde. – Vol. 28. – P. 327–352.

200. Oye P. van, 1951. Au sujet des Rhizopodes du Grand-Duche de Luxembourg // Biologisch Jaarboek Antwerpen. – Vol. 18. – P. 82–121.

201. Oye P. van, 1956. On the thecamoeban fauna of New Zealand with description of four new species and biogeografical discussion // Hydrobiologia. – Vol. 8 (1–2). – P. 16–37.

202. Oye P. van, 1958. Etude sur les Rhizopodes des marais du Sud-ouest d'Uvira (Congo-belge) // Hydrobiologia. – Vol. 10. – P. 85–127.

203. Page F.C., 1966. *Cryptodifflugia operculata* n. sp. (Rhizopodea: Arcellinida, Cryptodifflugidae) and the status of the genus *Cryptodifflugia* // Trans. Amer. Microsc. Soc. – Vol. 85. – P. 506–515.

204. Patterson R.T., 2014. *Mediolus*, a new genus of Arcellacea (Testate Lobose Amoebae) // Palaeontologia Electronica. – Vol. 17. – P. 1–8.

205. Payne R., 2011. Can testate amoeba-based palaeohydrology be extended to fens? // J. Quat. Sci. – Vol. 26. – P. 15–27.

206. Payne R.J., 2013. Seven reasons why protists make useful bioindicators // Acta Protozool. – Vol. 52. – P. 105–113.

207. Penard E., 1890. Etudes sur les Rhizopodes d'eau douce // Mem. Soc. Phys. Hist. Nat. Geneve. – Vol. 31 (2). Part 1. – P. 1–230.

208. Penard E., 1893. *Pelomyxa palustris* et quelques autres organismes inferieurs // Arch. Sci. Phys. Nat., ser. 3 (Bibl. Univers.). – Vol. 29. – P. 161–180.

209. Penard E., 1899. Les Rhizopodes de faune profonde dans le lac Leman // Rev. Suisse Zool. – Vol. 7 (1). – P. 1–142.

210. Penard E., 1902. Faune Rhizopodique du Bassin de Léman. – Genéve : Kündig. – 714 p.

211. Penard E., 1903. Notice sur les Rhizopodes du Spitzberg // Arch. Protistenk. – Vol. 2. – P. 238–282.

212. Penard E., 1905. Notes sur quelques Sarcodines. 1e partie // Rev. Suisse Zool. – Vol. 13 (3). – P. 585–616.

213. Penard E., 1906. Notes sur quelques sarcodinés // Rev. Suisse Zool. – Vol. 18. – P. 109–141.

214. Penard E., 1909. Sur quelques rhizopodes des mouses // Arch. Protistenk. – Vol. 17. – P. 258–296.

215. Penard E., 1911. Rhizopodes d'eau douce // Shackleton, E.H. (ed.). British Antarctic Expedition 1907–9, under the command of Sir E.

H. Shackleton, C.V.O. Reports on the scientific investigations. London: William Heinemann. – Vol. 1, "Biology", Murray, J. (ed.), part 6. – P. 203–262.

216. Penard E., 1912. Notes sur quelques Sarcodines. 3me oartie // Revue Suisse de Zoologie. – Vol. 20. – P. 1–29.

217. Penard E., 1917. Observations sur quelques Protozoaries peu connus ou nouveaux // Ann. Soc. Zool. Suisse. – Vol. 25. – P. 1–50.

218. Playfair G.I., 1914. Contributions to a knowledge of the biology of the Richmond River // Proceedings of the Linnean Society of New South Wales. – Vol. 39. part 1 (N_{2} 153). – P. 93–151.

219. Playfair G.I., 1917. Rhizopods of Sydney and Lismore // Proc. Linn. Soc. New South Wales. – Vol. 42. – P. 633–675.

220. Porter S.M., Knoll A.H., 2000. Testate amoebae in the Neoproterozoic Era: evidence from vase-shaped microfossils in the Chuar Group, Grand Canyon // Paleobiology. – Vol. 26. – P. 360–385.

221. Raikov I.B., 1982. The Protozoan nucleus. Morphology and Evolution. – Springer-Verlag, Berlin.

222. Ruggiero M.A., Gordon D.P., Orrell T.M., Bailly N., Bourgoin T., Brusca R.C., Cavalier-Smith T., Guiry M.D., Kirk P.M., 2015. A higher level classification of all living organisms // PLoS ONE. – Vol. 10. – P. 1–60.

223. Saedeleer H. de, 1934. Beitrag zur Kenntnis der Rhizopoden: morphologische und systematische Untersuchungen und ein Klassifikationsversuch // Mem. Mus. Roy. Hist. Nat. Belgique. – Vol. 60. – P. 1–128.

224. Santibáñeza P.A., Kohshima S., Scheihinga R.A., Ricardo Silva R., Jaramillom J.I., Labarcap P.J., Gino Casassar R., 2011. First record of testate amoebae on glaciers and description of a new species *Puytoracia jenswendti* nov. sp. (Rhizaria, Euglyphida) // Acta Protozool. – Vol. 50. – P. 1–14.

225. Schaudinn F., 1898. Rhizopoda Ost-Afrikas // Deutsch-Ost Afrika. Bd. 4. Die Thierwelt Deutsch-Ostafriks und der Nachbargebeite. Wirbellose Thiere. Part 18. – P. 1–13.

226. Schmidt H., 1926. Untersuchungen an Rhizopoden aus Buchenhohlen // Verh. Naturk.Ver. Rheinlande. – Vol. 82. – P. 218–226.

227. Schmidt A.R., Ragazzi E., Coppellotti O., Roghi G., 2006. A microworld in Triassic amber // Nature. – Vol. 444. – P. 835.

228. Schmidt A.R., Schönborn W., Schafer U., 2004. Diverse fossil amoebae in German Mesozoic amber // Palaeontology. – Vol. 47. – P. 185–197.

229. Schönborn W., 1962a. Neue Testaceen aus dem Grossen Stechlinsee und dessen Umgebung // Limnologica. – Vol. 1 (1). – P. 83–91.

230. Schönborn W., 1962b. Die Okologie der Testaceen im oligotrophen See, dargestellt am Beispel des Grossen Stechlinsees // Limnologica. – Vol. 1. – P. 111–182.

231. Schönborn W., 1962c. Zur Okologie der sphagnikolen, bryokolen und terricolen Testaceen // Limnoloica. – Vol. 1. – P. 231–254.

232. Schönborn W., 1964. Bodenbewohnende Testaceen aus Deutschland. II. Untersuchungen in der Umgebung des Grossen Stechlinsees (Brandenburg) // Limnologica. – Vol. 2 (4). – P. 491–499.

233. Schönborn W., 1965a. Studien uber die Gattung *Difflugiella* Cash (Rhizopoda, Testacea) // Limnologica. – Vol. 3 (3). – P. 315–328.

234. Schönborn W., 1965b. Beobachtungen an der Zellteilung von *Paraquadrula* (Testacea) // Limnologica. – Vol. 3. – P. 235–236.

235. Schönborn W., 1965c. Die Sedimentbewohnenden Testaceen einiger Masurischer Seen // Acta Protozool. – Vol. 3. – P. 297–309.

236. Schönborn W., 1966a. Untersuchungen uber die Testaceen Schwedisch-Lapplands. Ein Beitrag zur Systematik und Okologie der beschalten Rhizopoden // Limnologica. – Vol. 4 (3). – P. 517–559.

237. Schönborn W., 1966b. Beitrag zur Okologie und Systematik der Testaceen Spitzbergens // Limnologica. – Vol. 4 (3). – P. 463–470.

238. Schönborn W., 1966c. Beschalte Amöben (Testacean). Wittenberg-Lutherstadt : Ziemsen. – 112 p.

239. Schönborn W., 1983. Modifikabitat und Evolutionstrends bei Protozoen // Biol. Rdsch. – Vol. 21. – P. 225–235.

240. Schönborn W., 1989. The topophenetic analysis as a method to elucidate the phylogeny of testate amebas (Protozoa, Testacealobosia and Testaceafilosia) // Arch. Protistenk. – Vol. 137 (3). – P. 223–245.

241. Schönborn W., 1992. Adaptive polymorphism in soilinhabiting testate amebas (Rhizopoda) – its importance for delimitation and evolution of asexual species // Arch. Protistenk. – Vol. 142 (3–4). – P. 139–155.

242. Schönborn W., Foissner W., Meisterfeld R., 1983. Licht- und rasterelektronenmokroskopische Untersuchungen zur Schalenmorphologie und Rassenbildung bodenbewohneneder Testaceen (Protozoa: Rhizopoda) sowie vorschlage zur biometrischen Characterisierung von Testaceen-schalen // Protistologica. – Vol. 19 (4). – P. 553–566.

243. Schönborn W., Peschke T., 1988. Biometric studies on species, races, ecophenotypes and individual variations of soil-inhabiting testacea (Protozoa, Rhizopoda), including *Trigonopyxis minuta* n.sp. and *Corhytion asperulum* n.sp. // Arch. Protistenk. – Vol. 136. – P. 345–363.

244. Schouteden H., 1905. Note sur les organismes inferieurs; 2eme note // Ann. Soc. Royal. Malacologie (Bruxelles). – Vol. 40. – P. 3–16.

245. Schröter D., Wolters V., De Ruiter P.C., 2003. C and N mineralisation in the decomposer food webs of a European forest transect // Oikos. – Vol. 102. – P. 294–308.

246. Scott D., Medioli F., Schafer C., 2001. Monitoring in coastal environments using foraminifera and thecamoebian indicators. – Cambridge : Cambridge University Press.

247. Smet de W.H., Gibson J.A.E., 2009. On a new species of euglyphid testate amoeba, *Scutiglypha cabrolae*, from the Licancabur caldera lake, Central Andes // Acta Protozool. – Vol. 48. – P. 119–126.

248. Snegovaya N., Alekperov I., 2005. Fauna of testate amoebae of western Azerbaijan rivers // Protistology. – Vol. 4 (2). – P. 149–183.

249. Steinecke F., 1927. Leitformen und Leitfossilien des Zehlaubruches // Botanische Archiv; zeitschrift fur die gesamte Botanik (Koenigsberg). – Vol. 19. – P. 328–343.

250. Štěpánek M., 1952. Testacea of the pond of Hradek at Kunratice (Prague) // Acta Mus. Nat. Pragae, ser.b. – Vol. 8. – P. 1–55.

251. Štěpánek M., 1963. Die Rhizopoden aus Katanga (Kongo-Africa) // Ann. Mus. Roy. Afr. Centrale. Sciences Zoologiques. – Vol. 117. – P. 9–91.

252. Štěpánek M., 1967. Testacea des Benthos der Talsperre Vranov am Thayafluss // Hydrobiologia. – Vol. 29. – P. 1–66.

253. Sullivan M.E., Booth R.K., 2011. The potential influence of short-term environmental variability on the composition of testate amoeba communities in *Sphagnum* peatlands // Microb. Ecol. – Vol. 62. – P. 80–93.

254. Szelecz I., Fournier B., Seppey C., Amendt J., Mitchell E.A.D., 2014. Can soil testate amoebae be used for estimating the time since death? A field experiment in a deciduous forest // Forensic Science International. – Vol. 236. – P. 90–98.

255. Tarnogradsky D.A., 1959. Microflora and microfauna of peats in the Caucasus. 8. A sedge-sphagnum lake in the upper reaches of the Balkarian river Terek // Raboty severo kavkazskoj Gidrobiologiceskoj Stantsii (Trudy Severo-osetinskogo Sel'skokhozyaistvennogo Instituta). – Vol. 6. – P. 3–59.

256. Thomas R., 1958a. Le genre *Plagiopyxis* Penard // Hydrobiologia. – Vol. 10. – P. 198–214.

257. Thomas R., 1958b. Sur quelques *Euglypha* nouvelles ou peu connues observees en Afrique // Bull. Soc. Hist. Nat. Afr. Nord. – Vol. 49.

258. Thomas R., 1961. Note sur quelques Rhizopodes de France // Cah. Natur., Bull. Nat. Parisiens, New series. – Vol. 17. – P. 74–80.

259. Thomas R., 1962. Kystes et Enkystment chez les Thecamoebiens // Bull. Soc. Zool. France. – Vol. 87. – P. 276–280.

260. Thomas R., Chardez D., 1958. Etude critique de *Trinema penardi* nom.nov. (Thecamoebiens) // Cah. Nat., Bull. Nat. Parisiens, New series. – Vol. 14. – P. 101–104.

261. Thomas R., Gauthier-Lièvre L., 1959a. Note sur quelques Euglyphidae d'Afrique // Bull. Soc. Hist. Nat. Afr. Nord. – Vol. 50. – P. 204–221.

262. Thomas R., Gauthier-Lièvre L., 1959b. Le genre *Lesquereusia* Schlumberger 1845 (Rhizopodes testaces) // Bull. Soc. Hist. Nat. Afr. Nord. – Vol. 50. – P. 34–83.

263. Todorov M., Golemansky V., 2003. Morphology, biometry and ecology of *Arcella excavata* Cunningham, 1919 (Rhizopoda: Arcellinida) // Acta Protozool. – Vol. 42. – P. 105–111.

264. Tolonen K., Warner B.G., Vasander H., 1992. Ecology of testaceans (Protozoa, Rhizopoda) in mires in Southern Finland. 1. Autecology // Arch. Protistenk. – Vol. 142. – P. 119–138.

265. Tolonen K., Warner B.G., Vasander H., 1994. Ecology of Testaceans (Protozoa, Rhizopoda) in Mires in Southern Finland. 2. Multivariate-Analysis // Arch. Protistenk. – Vol. 144. – P. 97–112.

266. Valkanov A., 1962. *Euglyphella delicatula* n.g., n.sp. (Rhizopoda Testacea) und ihre Kopulation // Doklady Bolgarkoi Akademii Nauk. – Vol. 15. – P. 207–209.

267. Valkanov A., 1963. Über die Kopulation der Testaceen (Rhizopoda, Tetstacea) // Dokl. Bolg. Akad. Nauk. – Vol. 15 (3). – P. 305–308.

268. Virieux J., 1916. Recherches sur le plankton des lacs du Jura Central // Ann. Biol. Lacustr. – Vol. 8. – P. 5–192.

269. Vucetich M.C., 1976. Tecamebianos del Lago San Roque y de un ambiente vinculado al mismo (Cordoba, Argentina) // Limnobios. – Vol. 1. – P. 29–34.

270. Wailes G.H., 1912. Freshwater Rhizopoda and Heliozoa from the States of New York, New Jersey, and Georgia, U.S.A.; with supplemental note on Seychelles species // J. Linn. Soc. London, Zoology. – Vol. 32 (214). – P. 121–161.

271. Wailes G.H., 1913. Freshwater Rhizopoda from North and South America // J. Linn. Soc. London, Zoology. – Vol. 32 (216). – P. 201–218.

272. Wailes G.H., 1925. Some new or rare Protozoa from British Columbia // Ann. Mag. Nat. Hist. – Vol. 16. – P. 40–48.

273. Wailes G.H., 1927. Rhizopoda and Heliozoa from British Columbia // Ann. Mag. Nat. Hist. – Vol. 20. – P. 153–156.

274. Wailes G.H., Penard E., 1911. Rhizopoda // Proceedings of the Royal Irish Academy. 31 "A biological survey of Clare Island in the County of Mayo, Ireland, and of the adjoining district", sect. 3, "Zoology (Oligochaeta to Protozoa). Marine Ecology. Summary", Part 65. – P. 1–64.

275. Wallich G.C., 1864. On the extent, and some of the principal causes, of structural variation among the Difflugian Rhizopods // Ann. Mag. Nat. Hist. Ser. 3. - Vol. 13. - P. 215-245.

276. West G. S., 1901. On some British freshwater Rhizopods and Heliozoa // J. Linn. Soc. London, Zoology. – Vol. 28 (183). – P. 308–342.

277. West G. S., 1903. Observations on freshwater Rhizopods with some remarks on their classification // J. Linn. Soc. London, Zoology. – Vol. 29. – P. 108–117.

278. Wilkinson D., Mitchell E.A.D., 2010. Testate amoebae and nutrient cycling with particular reference to soils // Geomicrobiol Journal. – Vol. 27. - P.520-533.

279. Wylezich C., Meisterfeld R., Meisterfeld S., Schlegel M., 2002. Phylogenetic Analyses of Small Subunit Ribosomal RNA Coding Regions Reveal a Monophyletic Lineage of Euglyphid Testate Amoebae (Order Euglyphida) // J. Euk. Microbiol. – Vol. 49. – P. 108–118.

Научное издание

Цыганов Андрей Николаевич, **Бабешко** Кирилл Владимирович, **Мазей** Юрий Александрович

Определитель родов раковинных амеб = A Guide to Testate Amoebae with the Keys to Genera

На английском языке

Корректор М. М. Кузнецова Компьютерная верстка М. Б. Жучковой Дизайн обложки А. А. Стаценко

Подписано в печать 23.11.2016. Формат 60×84¹/₁₆. Усл. печ. л. 7,67. Тираж 500. Заказ № 009094.

Издательство ПГУ 440026, Пенза, Красная, 40. Тел./факс: (8412) 56-47-33; e-mail: iic@pnzgu.ru